

Concentration of Cells and Elimination of Extraneous Background Signals in Laser-Induced Breakdown Spectroscopy to Identify, Differentiate and Detect Bacteria



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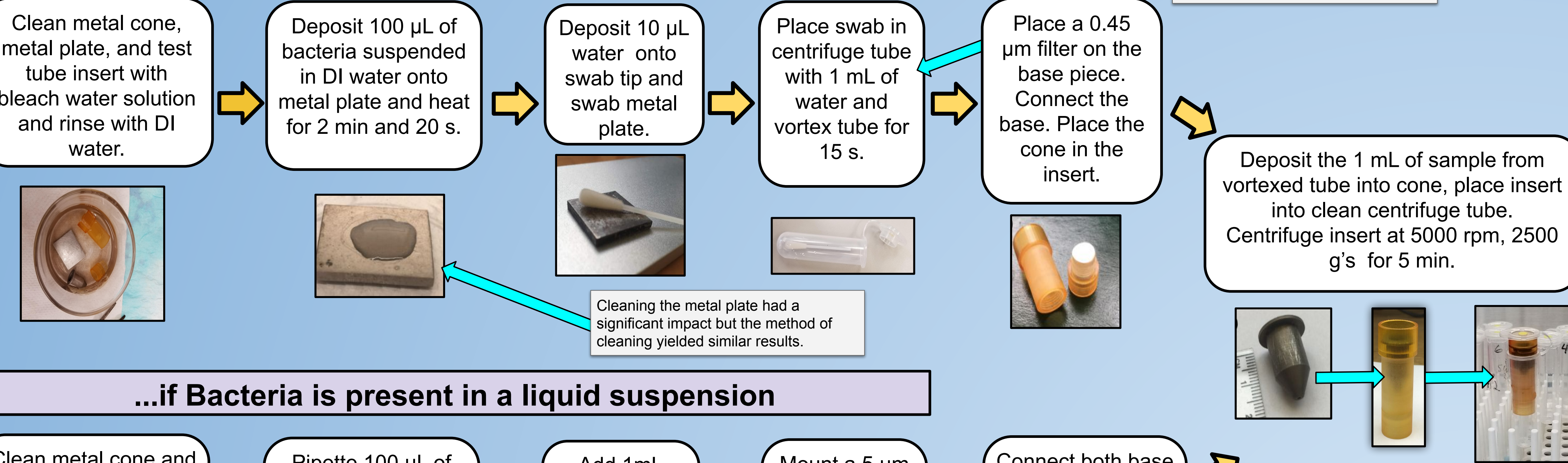


Introduction

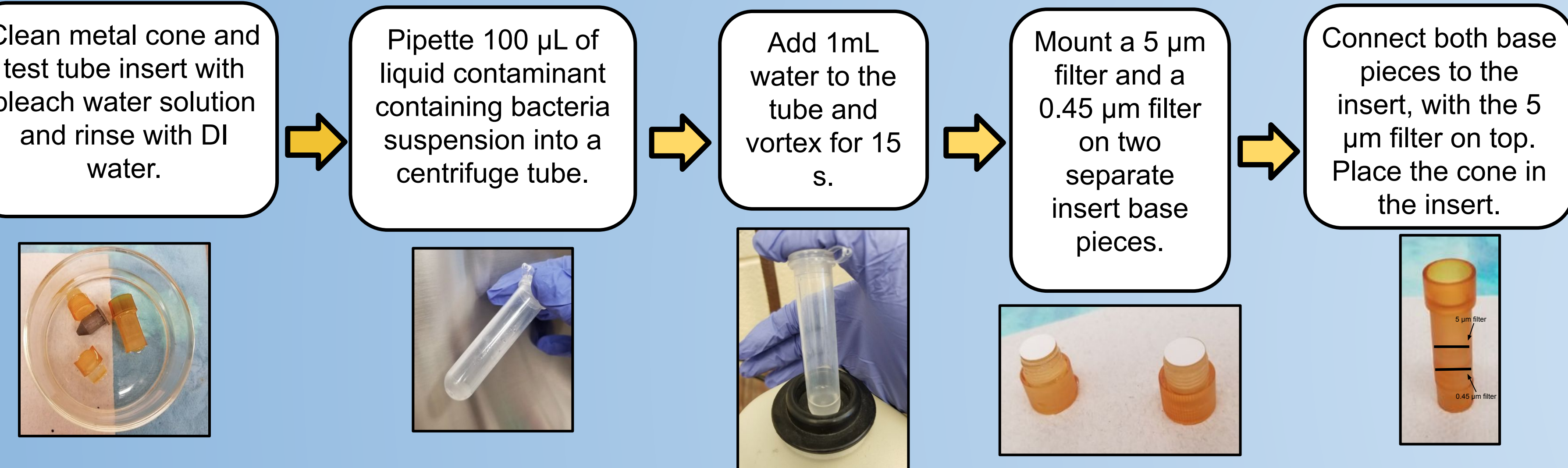
There is an urgent need to develop faster ways to identify pathogenic bacteria. Our aim is to develop a real-time point-of-care medical diagnostic technology utilizing Laser - Induced Breakdown Spectroscopy (LIBS). Our procedure consists of common materials and equipment that could be easily implemented in a clinical setting to rapidly identify bacteria based on their elemental composition, so that more targeted treatment can begin as soon as possible.

Current Method of Analyzing Bacteria

...if Bacteria is present and swabbed from a surface

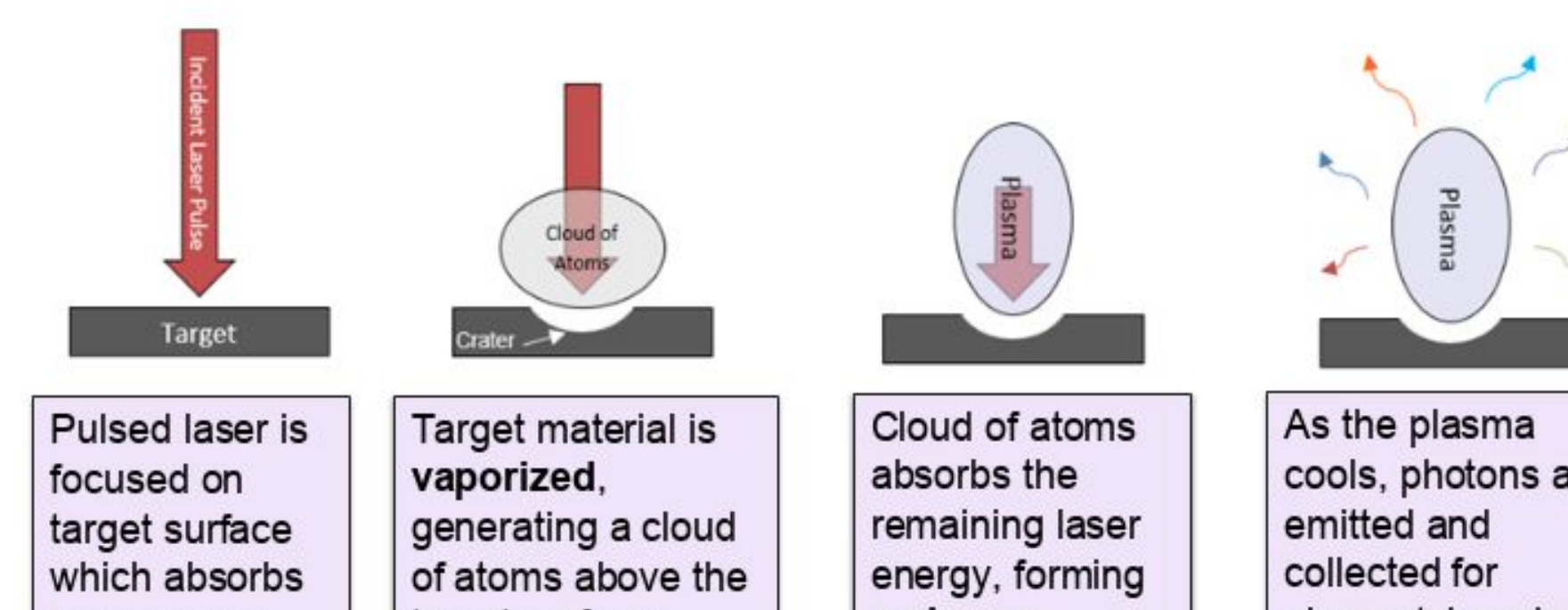


...if Bacteria is present in a liquid suspension



Laser-Induced Breakdown Spectroscopy (LIBS)

LIBS is an elemental analysis technique

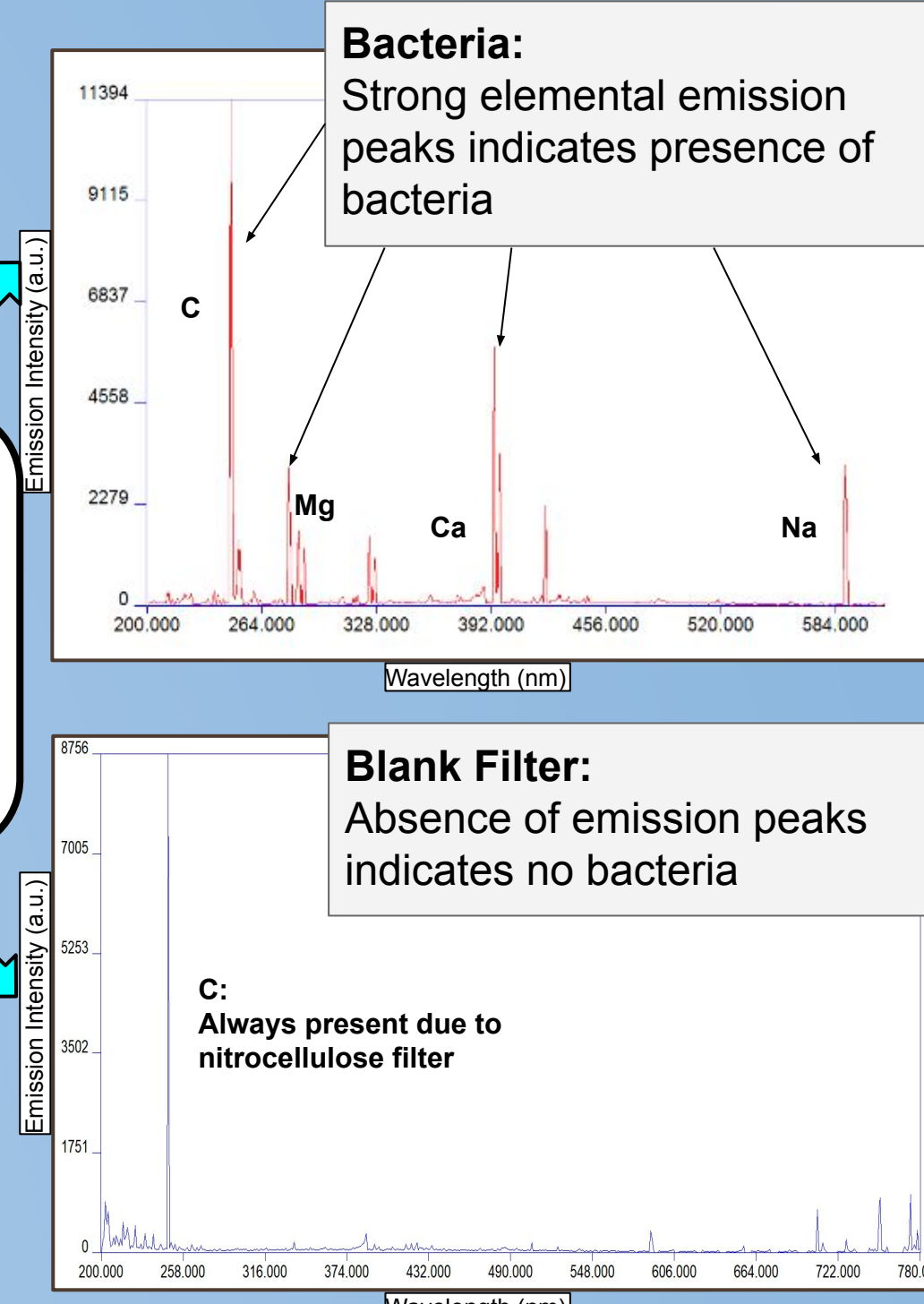


Measured Atomic Emission Lines In Bacterial LIBS Spectra

Element	Emission Line Wavelength (nm)
Carbon (C)	247.856
Phosphorus (P)	213.618, 214.914, 253.398, 253.560, 255.326, 255.491
Magnesium (Mg)	277.983, 279.079, 279.553, 279.806, 280.271, 285.213
Calcium (Ca)	317.933, 393.366, 396.847, 422.673
Sodium (Na)	588.995, 589.593

LIBS Advantages

- Can be done on solids, liquids, gases and bacteria
- Requires only µg of sample
- Elemental composition can be determined in seconds

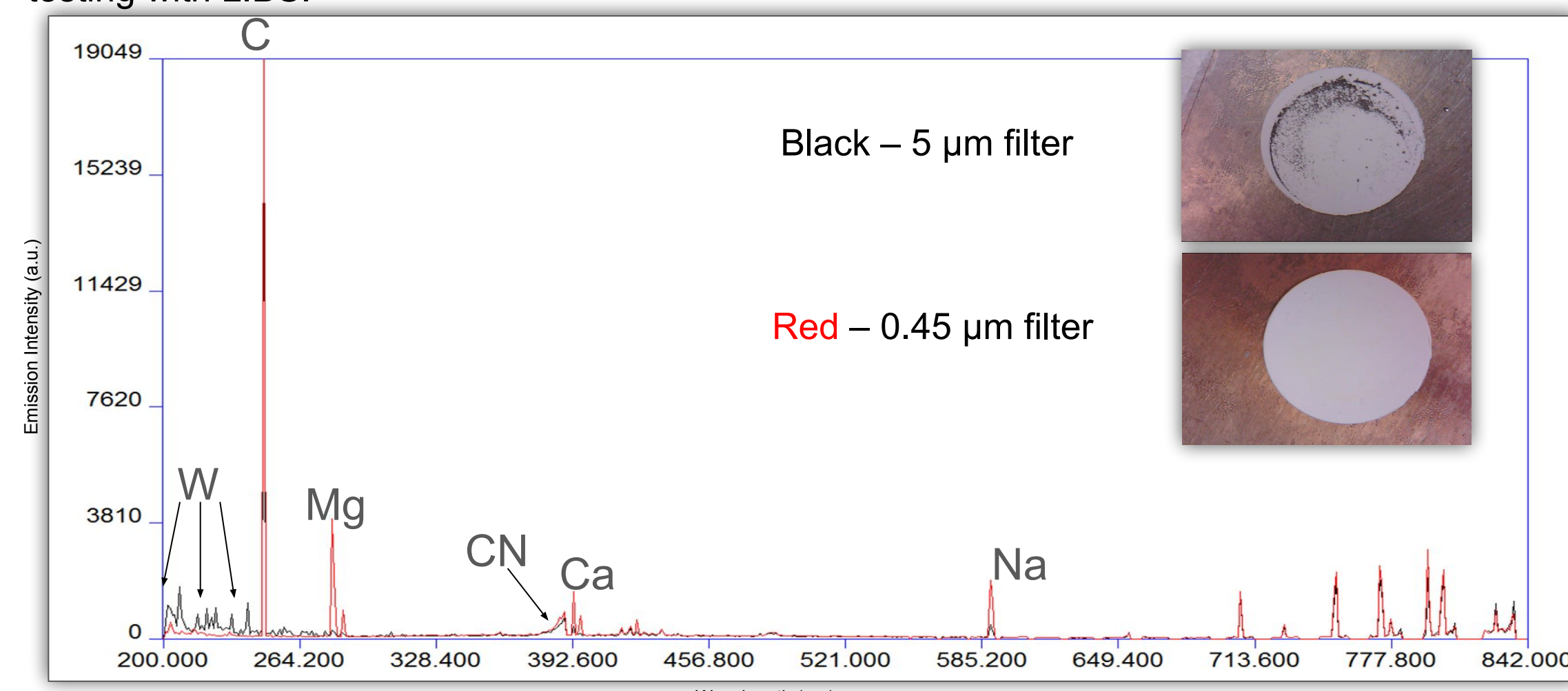


Scope of the Work

Laser ablation of the filter medium and other elemental contaminants yields a non - zero background signal when a control experiment is performed in the absence of any bacterial cells. The purpose of this research was to optimize the bacterial sample preparation protocol by identifying the source of this background signal and introducing new cleaning procedures that reduce this background signal. Following this preparation procedure, the ability to discriminate and/or identify low titers of different types of pathogenic bacteria using the centrifuge insert / cone bacterial concentration technique is being investigated.

Separation of a Contaminant from a Bacterial Suspension - Dual Centrifugation

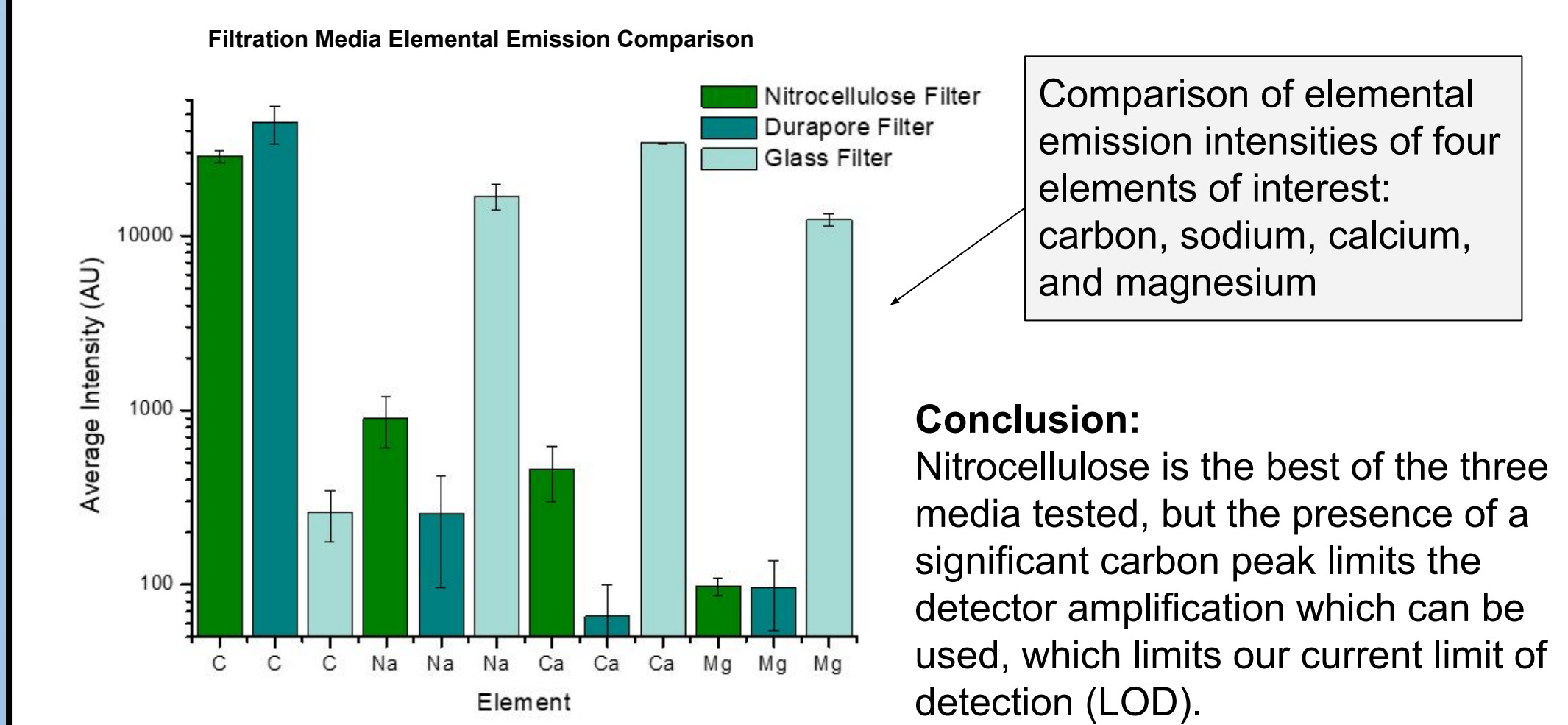
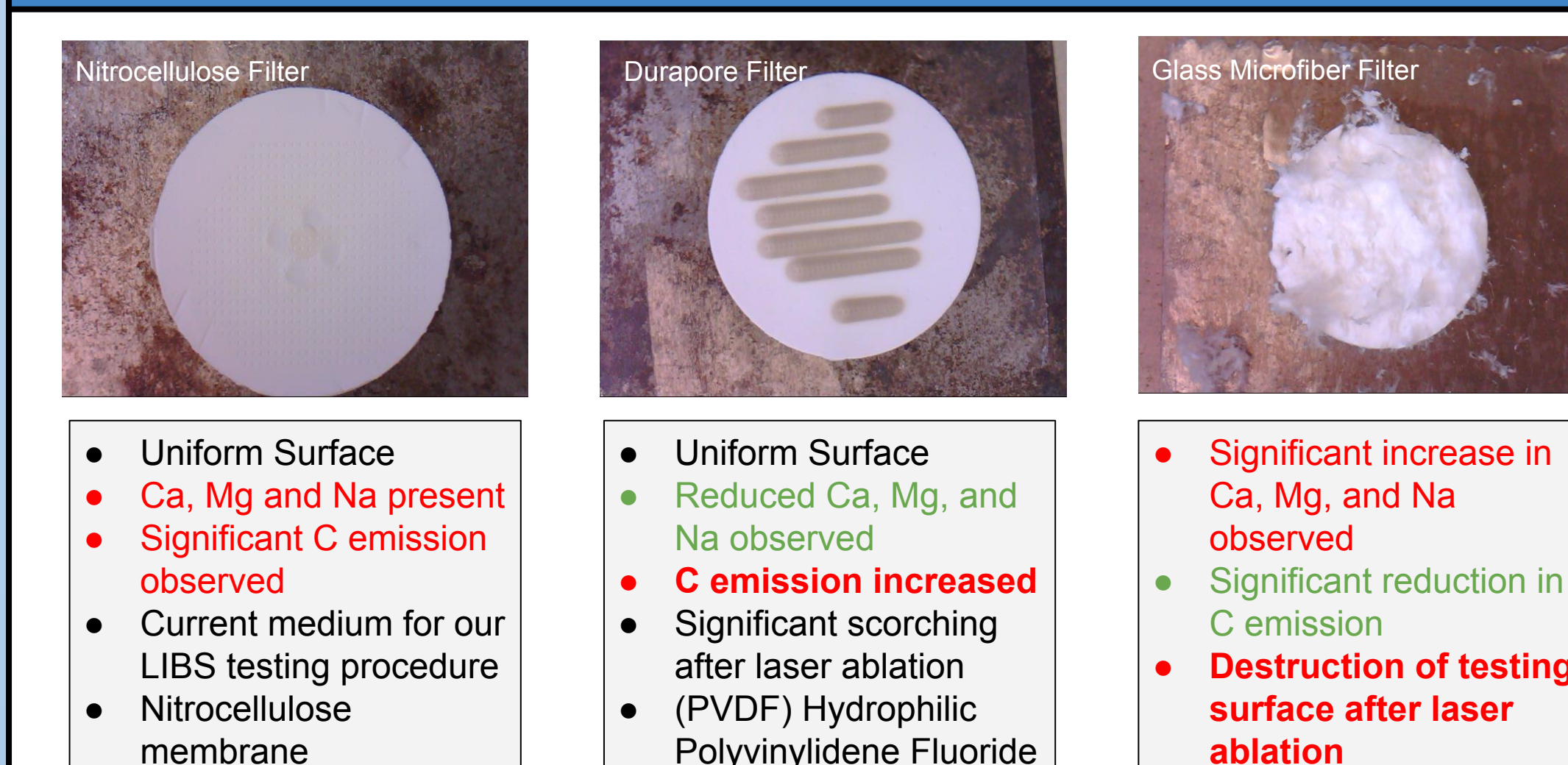
In a clinical setting, the biological fluid specimen to be tested will likely consist of unwanted cells (greater than 10 µm) that would need to be separated from the bacteria (~1 µm) prior to testing with LIBS.



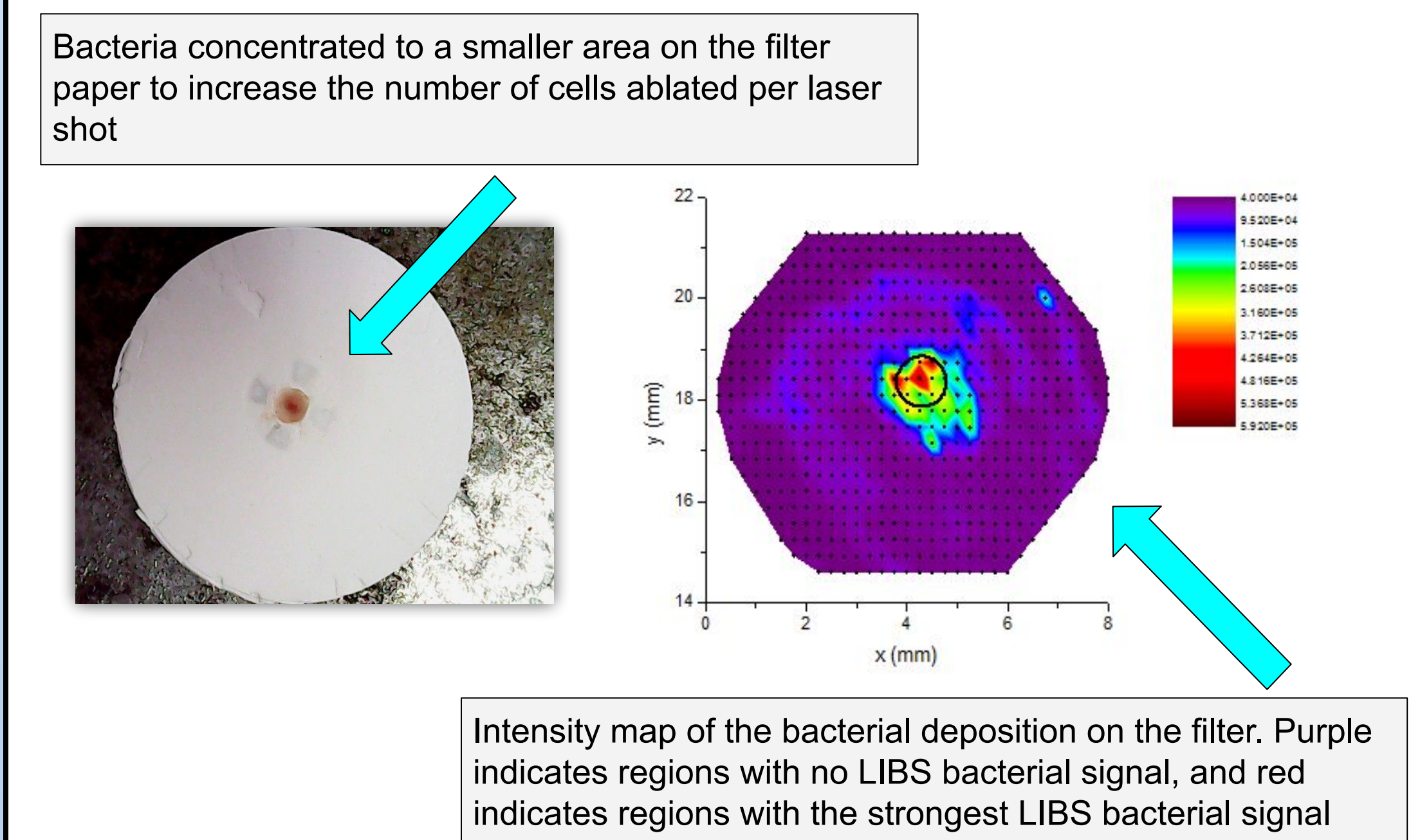
An *E. coli* suspension with tungsten powder (12 µm APS) as the contaminant was deposited in the insert with the 5 µm filter paper on top and the 0.45 µm filter paper below it. The tungsten powder was caught by the 5 µm filter while - 90% of the bacteria passed through it and settled onto the 0.45 µm filter.

Conclusion: The use of this centrifuge tube insert with filter media of different pore sizes was effective at separating a contaminant from bacteria based on its size.

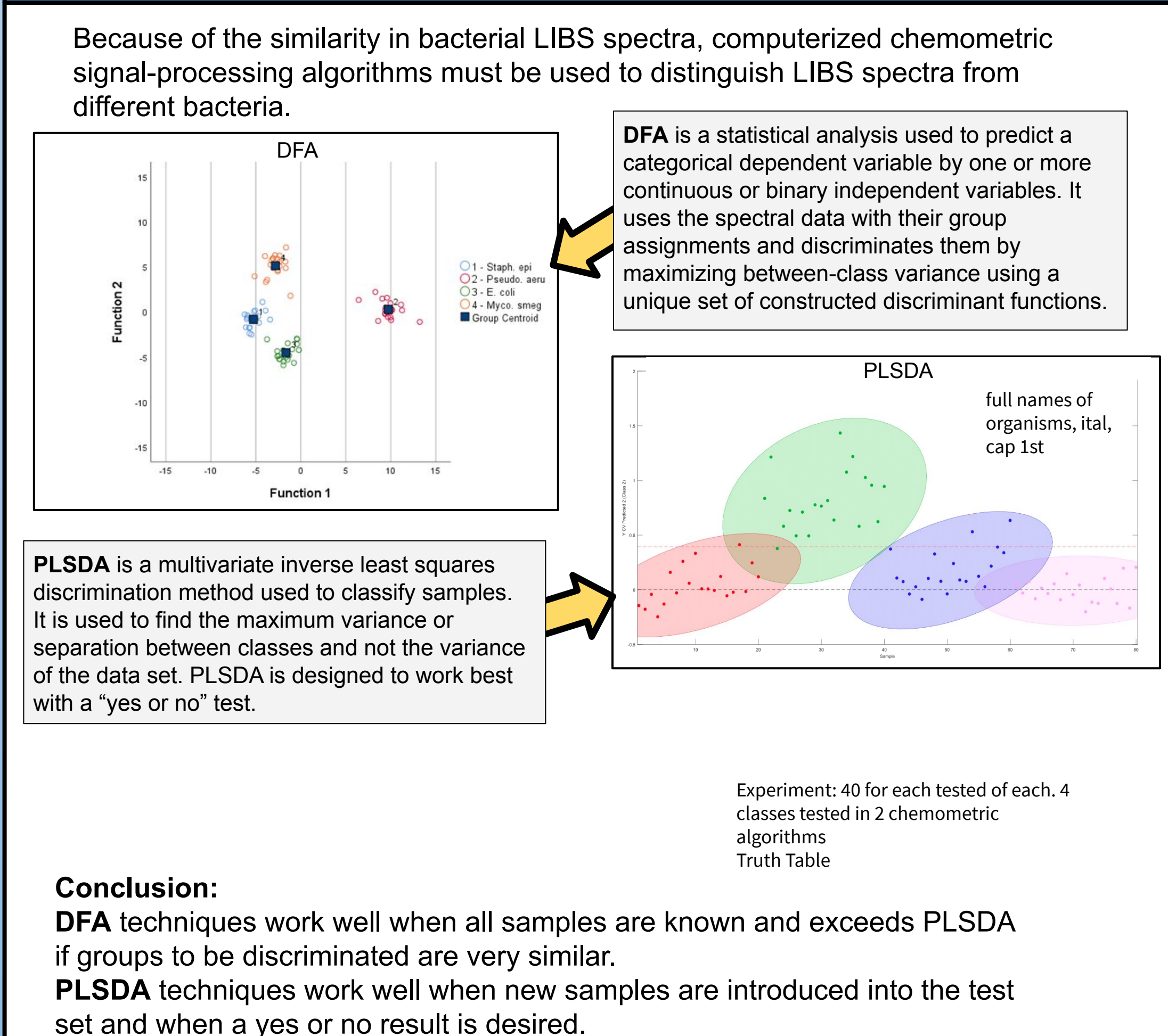
The Effect of the Filtration Media on the LIBS Emission Spectra



Improving the Limit of Detection

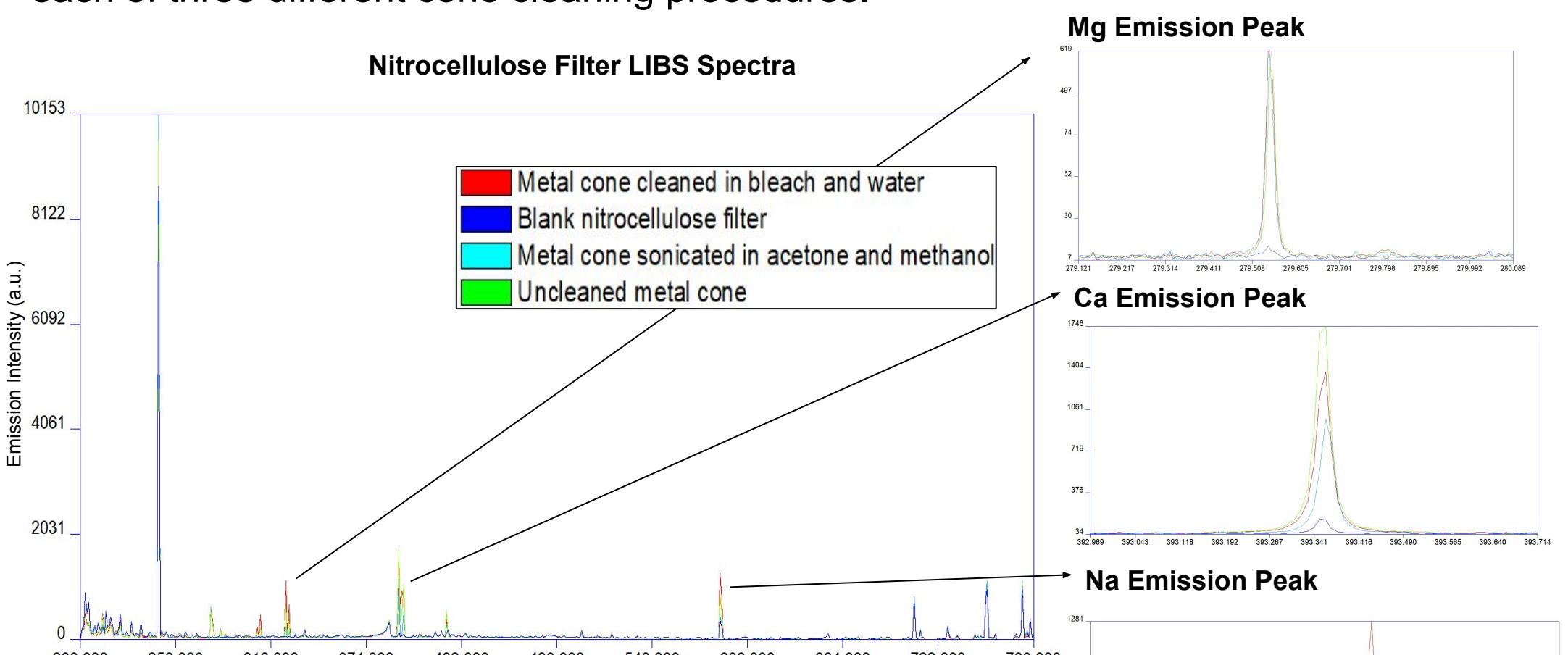


Chemometrics: DFA and PLSDA

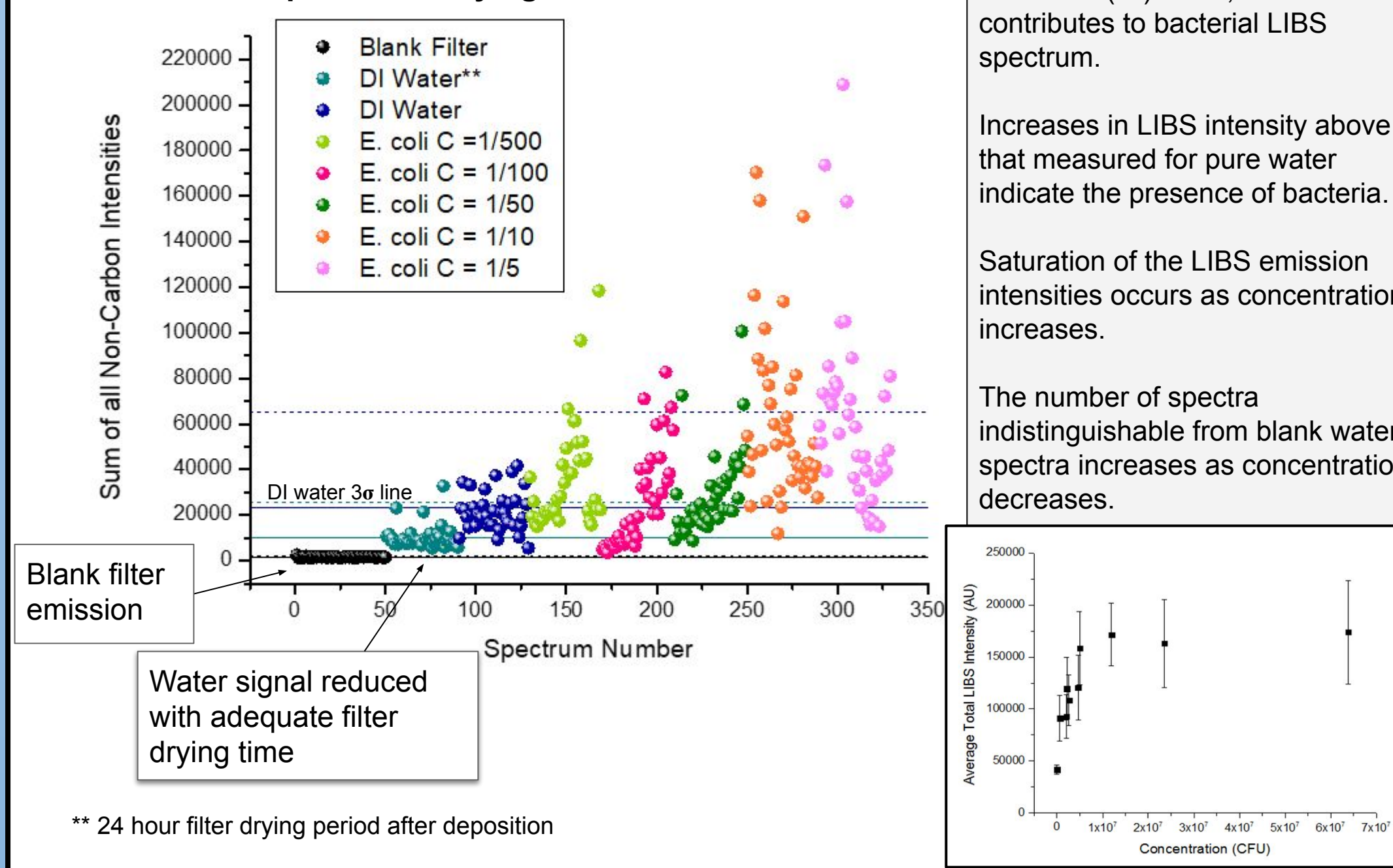


The Effect of Cleaning the Metal Cone

20 LIBS filter spectra were acquired when water (no bacteria) was centrifuged through the cone to compare the elemental intensities of carbon, sodium, magnesium and calcium for each of three different cone cleaning procedures.



Comparison of Water Control Spectra with Bacterial Spectra at Varying Concentrations



Conclusions and Future Work

The protocols developed in this experiment (cleaning of cone, cleaning of plate) will be integrated as a part of the current testing protocol, which will reduce the limit of detection of bacteria with LIBS.

Dual centrifugation and the use of two filters of different pore sizes will be tested on yeast cells to simulate a fluid suspension with contaminant through the metal cone.

The addition of a 'drying stage' will be investigated.

The limits of detection will be determined for bacteria swabbed off of surfaces as well as deposited in fluid suspensions (both simulating clinical tests).

The discrimination of different bacteria when swabbed off of surfaces and deposited in fluid suspensions in the cone will continue to be investigated.

Testing of actual clinical specimens obtained from pathology labs will be initiated including bacteria suspended in complex mixtures (i.e. blood, saliva, cerebral spinal fluid). Other areas of interest include automation techniques to make the technique more usable by non-experts and more safe, without loss of sensitivity or specificity.

Acknowledgements

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References

- [1] - A.E. Paulick, *Development of Laser-Induced Breakdown Spectroscopy as a Rapid Diagnostic Tool for Bacterial Infection*, University of Windsor, Windsor, 2018.
- [2] - S.J. Rehse, A Review of the Use of Laser-Induced Breakdown Spectroscopy for Bacterial Classification, Quantification, and Identification, *Spectrochim. Acta B* 154 (2019) 50-69.
- [3] - D.J. Malenfant, *Influences on the Emissions of Bacterial Plasmas Generated through Nanosecond Laser-Induced Breakdown Spectroscopy*, University of Windsor, Windsor, 2016.