Laser-Induced Breakdown Spectroscopy as a Rapid Diagnostic Tool for Bacterial Detection and Discrimination

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# Motivation

- Current methods of bacterial identification in a clinical setting
  - require expertise in microbiology
  - labor-intensive
  - slow

For example: standard culturing techniques for bacterial identification take 1-3 days

- Patients are treated with broad-spectrum drugs that have given rise to the crisis of antibiotic resistant bacteria
- Rapid and accurate diagnosis of bacterial infection are required so that more targeted treatment can begin as soon as possible



#### GOAL:

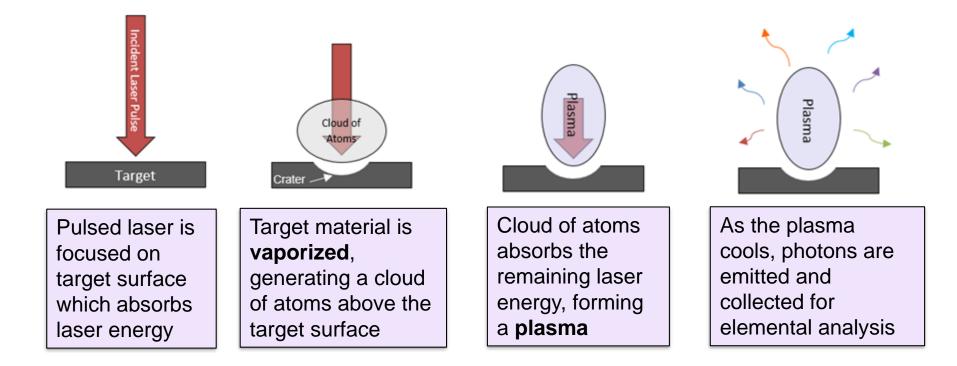
**Rapidly identify bacteria** based on their elemental composition using laser-induced breakdown spectroscopy (LIBS)

This includes developing a **quick bacterial preparation method** prior to testing that utilizes equipment and methods that are **common in a clinical setting** 



## Laser-Induced Breakdown Spectroscopy (LIBS)

#### LIBS is an **elemental analysis technique**





### LIBS Advantages

- Can be done on **solids**, **liquids**, **gases** and <u>bacteria</u>
- Little to no sample preparation
- Requires only  $\mu$ **g** of sample
- **Fast:** elemental composition can be determined in under 1 second
- Simultaneously detects all elements in periodic table
- The use of the laser allows for point sampling & elemental mapping



### **Overview of Methodology**



Bacteria is cultured on TSA plates





Bacterial suspension is vortexed and deposited on nitrocellulose filter paper





Bacterial cells are removed and suspended in 1.5 mL deionized water

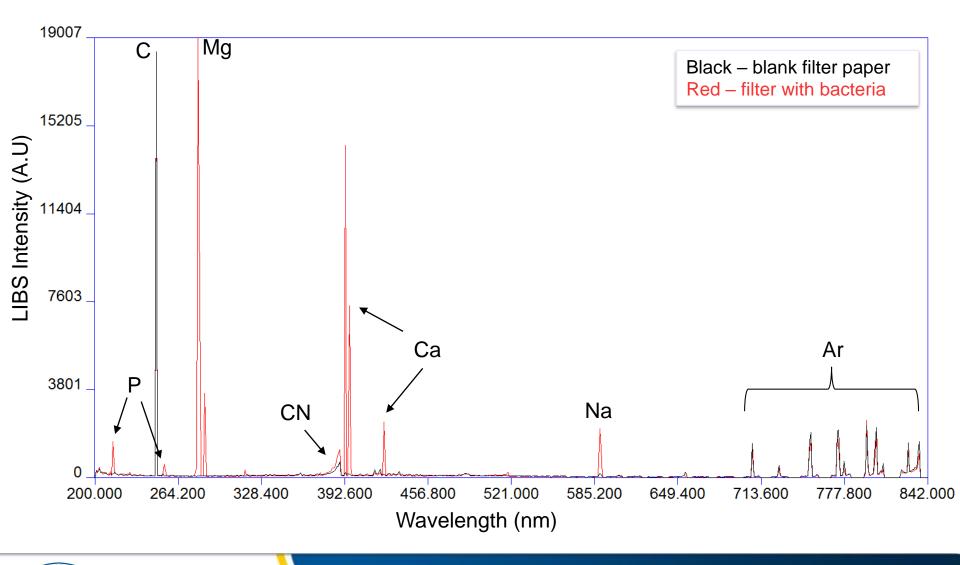


Filter paper is mounted on a steel piece and ablated with laser





After laser ablation, light from the plasma is dispersed, revealing the sample's elemental composition



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Bacteria are discriminated based on elemental composition

C, P, Mg, Ca, and Na can be used to identify a bacterium's species

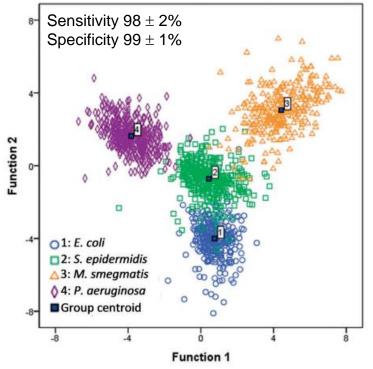
Bacterial spectra are classified using discriminant function analysis (**DFA**)

Unknown spectra are classified against a precompiled library of known spectra

#### **Bacterial library:**

- 164 independent variables (intensities of elemental lines and ratios of these lines to each other)
- ~ 1500 spectra acquired over 3 months from 4 species of bacteria (*E. coli, S. epidermidis, M. smegmatis, P. aeruginosa*)

### Bacterial classification based on elemental composition measured by LIBS





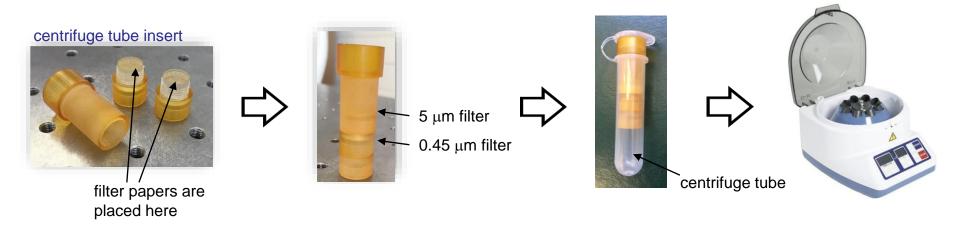
### Preparation Method to Separate a Contaminant from a Bacterial Suspension

Biological samples (blood sample, swab sample, etc.) will likely contain **unwanted cells** that would need to be separated from the bacteria before testing with LIBS

#### **Cell sizes:**

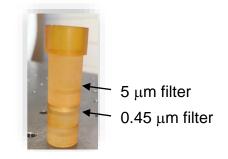
- Bacteria ~ 1 μm
- > Red blood cell ~ 6-8  $\mu$ m
- Eukaryotic cells ~ 10-100 μm

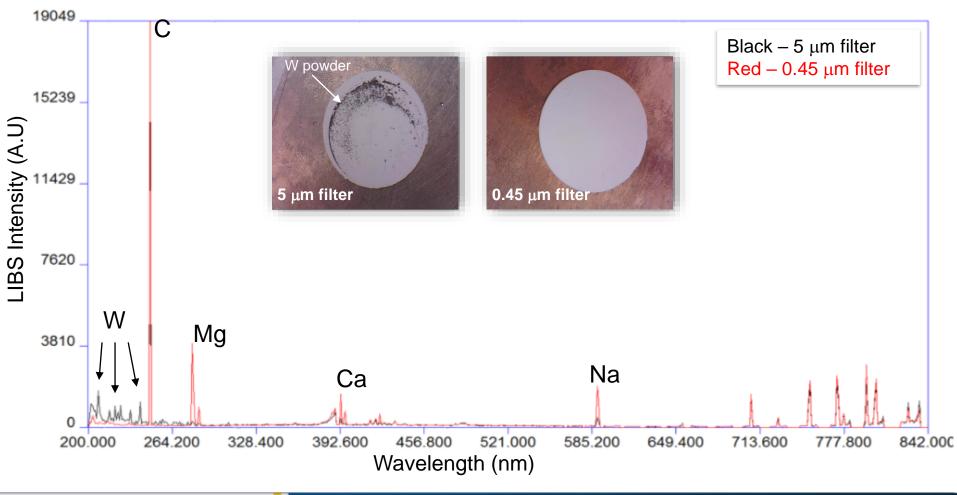
Isolate the bacteria using filter papers with different pore sizes (5  $\mu$ m and 0.45  $\mu$ m)





Tungsten powder (12  $\mu$ m average particle size) added to *E. coli* suspension to simulate unwanted cells in a biological sample



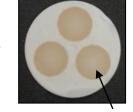


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### **Previous Bacterial Deposition Procedures**

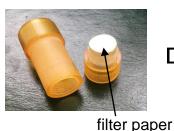
#### 1) Well-plate



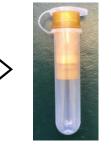


**Bacterial lawn** 

#### 2) Centrifuge tube insert







Bacterial LOD ~ **50 000 CFU** per laser ablation event

Bacterial LOD ~ 90 000 CFU per laser ablation event

Number of bacterial cells present in clinical samples:

- < 100 CFU/mL in blood</li>
- 0-200 CFU in typical nasal swab

These LOD's are *not clinically relevant*. Bacterial LOD with LIBS MUST be lowered.

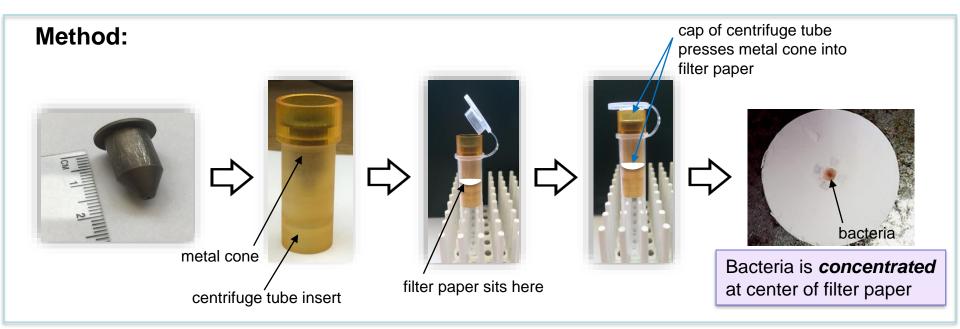


### New Deposition Procedure: Metal Cone

Metal cone to force deposition of bacteria onto *smaller region* at center of filter paper

#### Why do this?

Increases the number of bacterial cells per unit area, leading to *more bacterial cells ablated in a laser shot* compared to previous deposition procedures

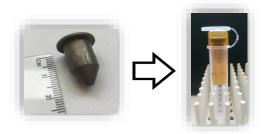




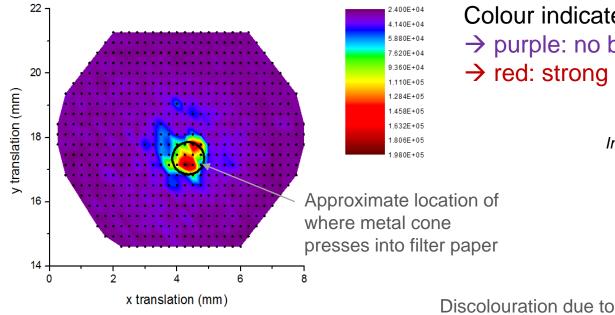
### Metal Cone: Bacterial Concentration

presence of bacteria

- E. coli deposited on filter paper with metal cone
- 569 LIBS spectra acquired across filter



Intensity map depicting bacterial deposition on filter paper for bacteria deposited with metal cone



→ purple: no bacterial signal
→ red: strong bacterial signal

Image of filter paper after data acquisition



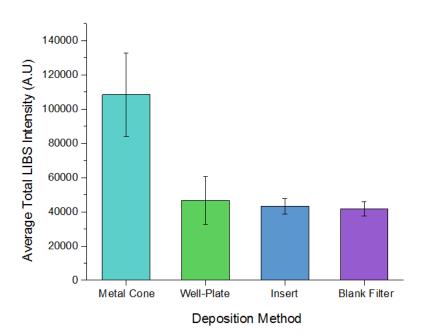


#### Comparison of LIBS Signal to **Previous Deposition Methods**



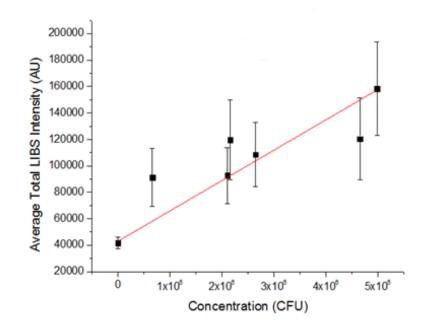
metal cone well-plate

insert



Suggestive of a **lower LOD** for bacteria deposited with metal cone

## Metal Cone Limit of Detection



#### LOD ~ 5 500 CFU per laser ablation event

#### **Recall:**

Well-plate → LOD ~ 50 000 CFU per laser ablation event Insert → LOD ~ 90 000 CFU per laser ablation event



## Conclusions

 Preparation method to separate unwanted material from bacterial suspension was effective

<u>Future work</u>: test this method using a contaminant that more closely simulates biological cells

- Metal cone:
  - effective at concentrating bacterial cells to a small region of the filter paper
  - significantly lowered bacterial LOD with LIBS compared to previous methods of bacterial deposition
- Future work: LIBS analysis on bacteria collected with swabs that are used
  in hospitals

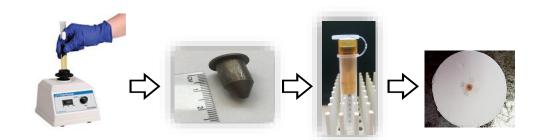






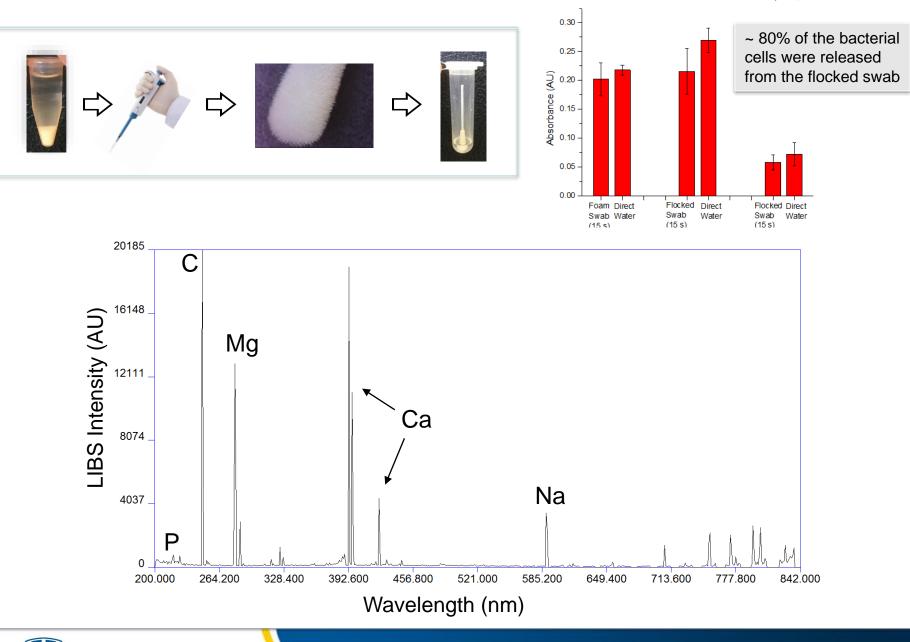
#### Cannot shoot right on the swab

- $\rightarrow$  Surface is too irregular
- $\rightarrow$  Bacterial cells are not concentrated





Absorbance Measurements of E. coli Cells - 50 µL Pipetted



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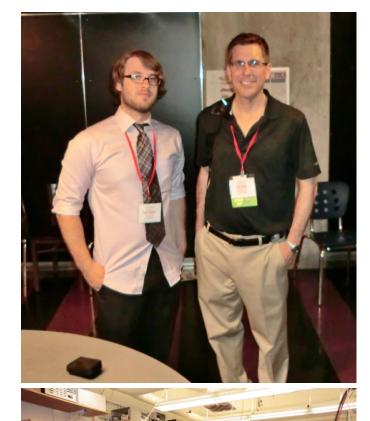
### Acknowledgments



• University of Windsor Student Life Enhancement Fund

University of Windsor Faculty of Science







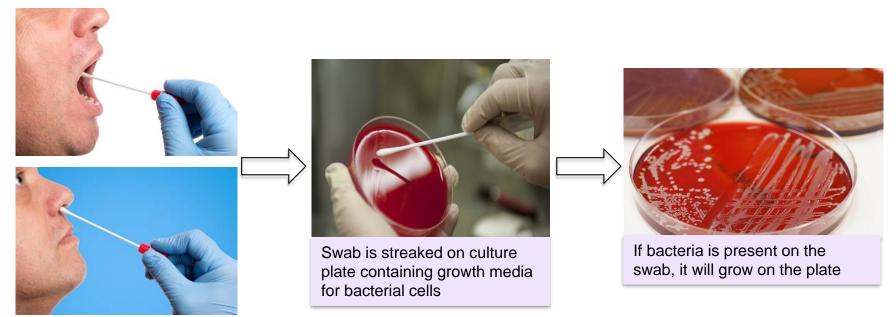
New students welcome!

If interested, contact Dr. Steven Rehse rehse@uwindsor.ca



### **Bacterial Collection with Swabs**

Some clinical specimens are collected with swabs



time consuming & require microbiology expertise

#### Can we use LIBS instead?

