



Development of a Simple and Efficient Clinical Protocol for Identifying and Quantifying Bacterial Cells in Liquid Suspensions with Laser-Induced Breakdown Spectroscopy

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Presented in Symposium #001: Laser-Induced Breakdown Spectroscopy (LIBS) for the Benefit of the Global Community

Sunday, December 20, 2021

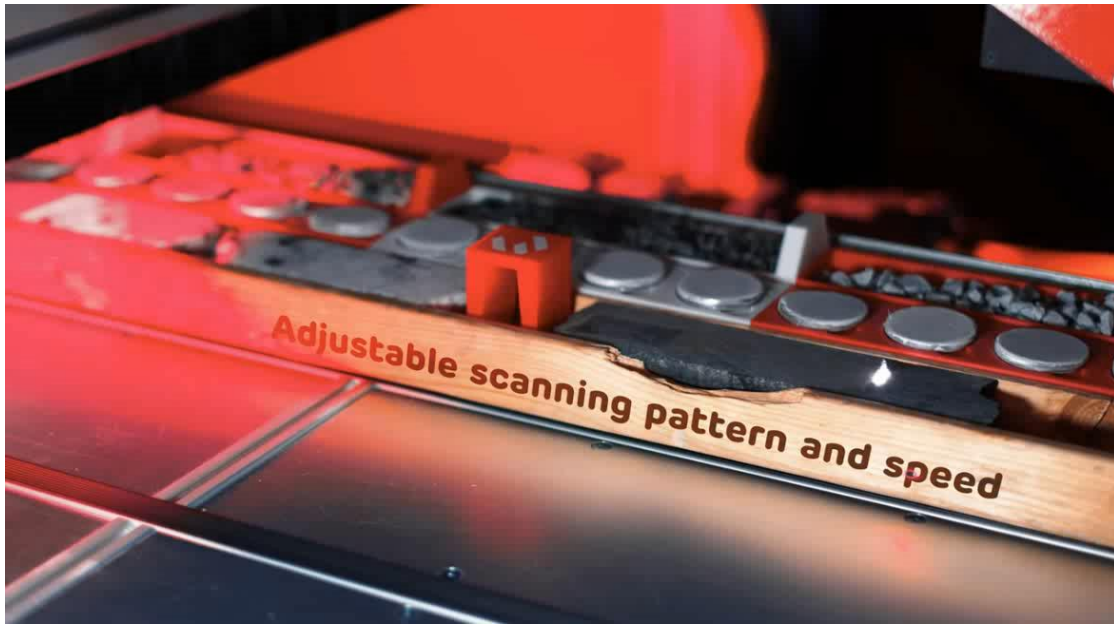


University of Windsor

Motivation



We all know the advantages that LIBS brings to an elemental analytical analysis...specifically speed and autonomous classification.



ECORE drill core scanner, by Elemission



SHREDDERSORT, by Lenz Instruments

Motivation

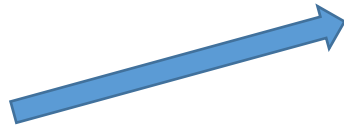


A similar analysis of specimens for the detection and identification of pathogenic bacteria is badly needed.



✓ Current “standard” methods of diagnosis takes ~1-3 days

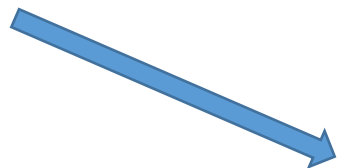
Delaying the initiation of appropriate antibiotic therapy can lead to:



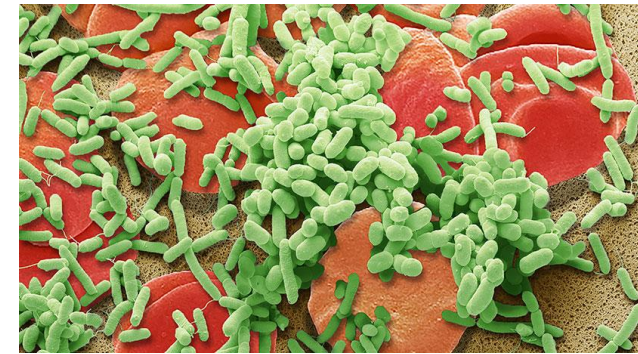
Worsening patient outcomes, including death (sepsis, meningitis)



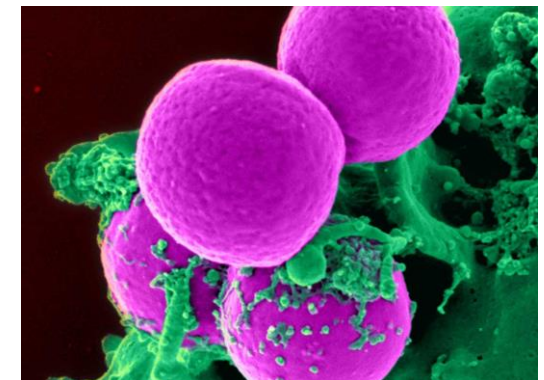
Increased expense due to treatment time, hospitalization (hospital-acquired infections)



Overuse/abuse of broad-spectrum antibiotics leading to antibiotic resistant strains (MRSA)



Composite coloured scanning electron micrograph (SEM) of sepsis. (Steve Gschmeissner/ Science Photo Library)



<https://www.airius.bg/all-resources/how-can-medical-settings-reduce-the-risk-of-mrsa-transmission-with-airius-pureair/>

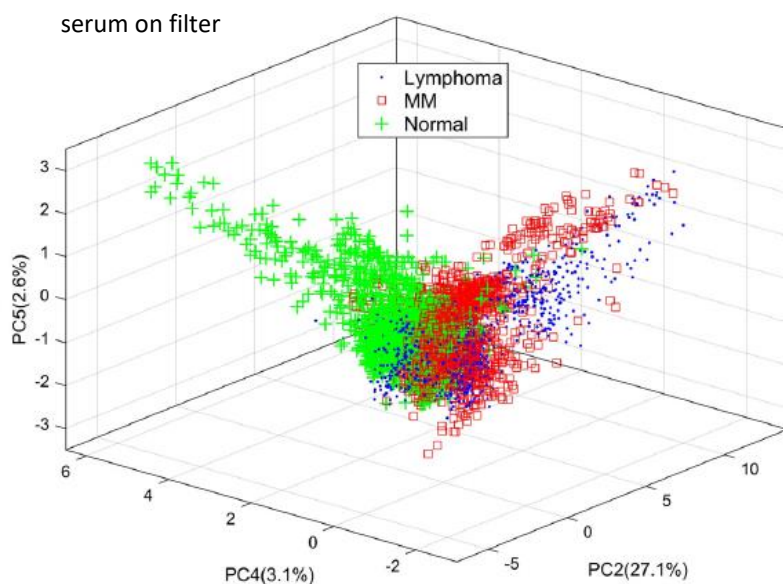
The idea to use LIBS on medical specimens is very reasonable:

Analysis of Biofluids

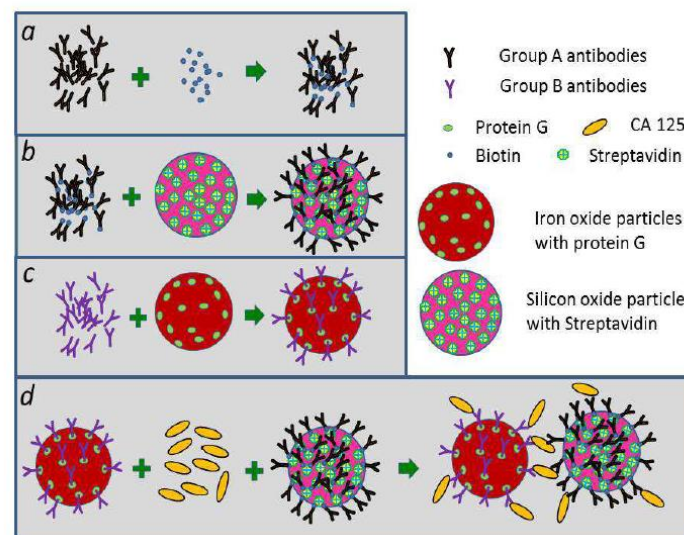
Biomarkers in fluid specimens

With a combination of appropriate substrate and algorithm, melanoma biomarkers in blood showed discrimination between healthy and diseased mice with accuracy up to 96%, but *direct analysis of LIBS spectra did not provide any conclusive results.* (Gaudio et al. 2018)

Multi-element micro- and nanoparticles labelling approach an attractive alternative (Markushin et al. 2012)



Chen et al., 2018



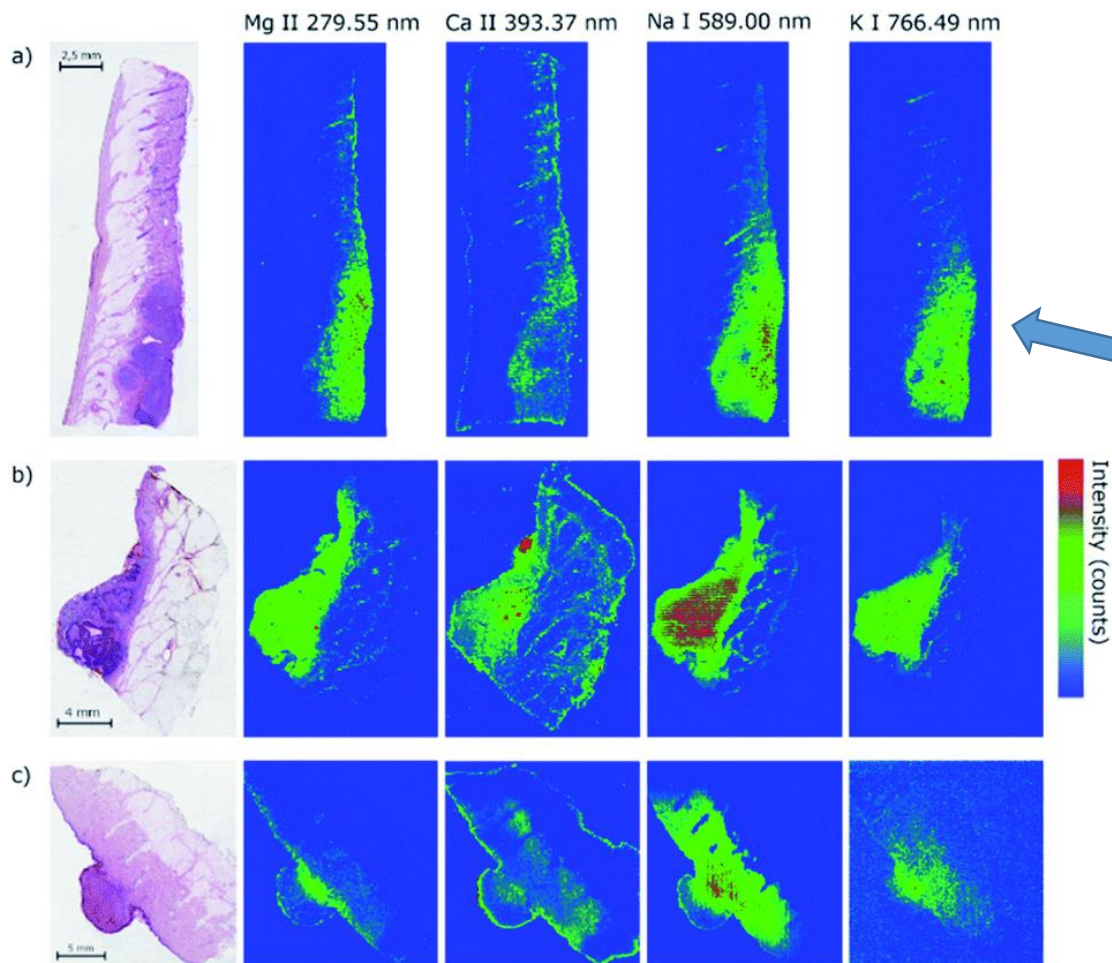
Markushin et al., 2012

CA125 ovarian cancer biomarker detected at the level of 10 U/mL (estimated LOD ~ 1 U/mL)

The idea to use LIBS on medical specimens is very reasonable:

Analysis of Tissues

Histological sections



Tissues

Higher concentrations of major and trace elements such as **Mg**, **Fe**, **Ca**, **Na**, and **K** in the neoplastic tissues.

Unfortunately a lot of work done *ex vivo* and in less-than-realistic experiments (caution)!

Our next speaker:

The imaging of biotic elements (Mg, Ca, Na, and K) provides the elemental distribution within the tissue.

The elemental images were correlated with the tumor progression and its margins, as well as with the difference between healthy and tumorous tissues

Bacteria



Can all-optical technique of LIBS really identify bacteria in under a second?



LIBS spectral fingerprint

- growth-medium independent
- independent of sample (are)
- independent of v (inactivated by UV light)
- obtainable even when other types of bacteria or contaminants are present (mixed samples)
- obtainable from urine specimens
- capable of strain discrimination
- obtainable from about 500 bacteria

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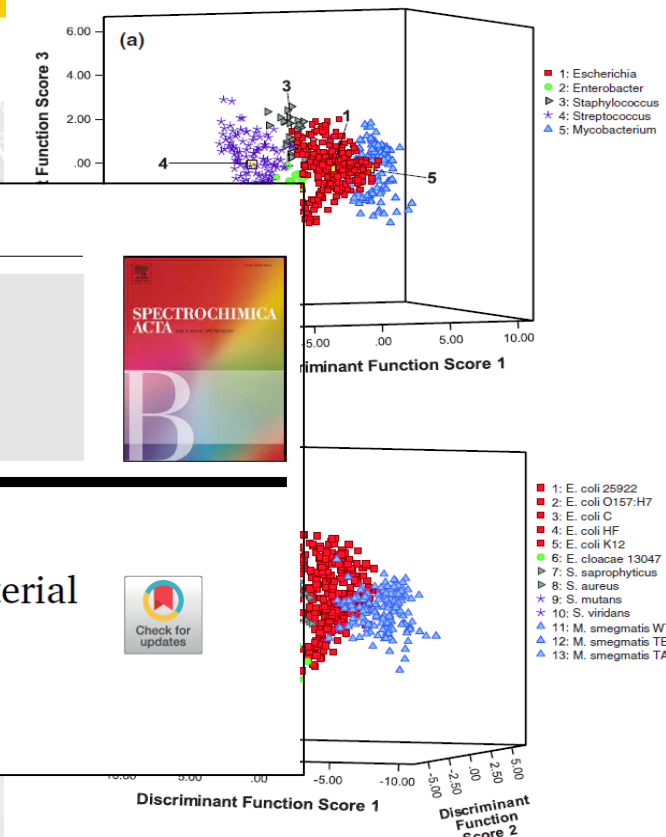
ELSEVIER

Invited Review

A review of the use of laser-induced breakdown spectroscopy for bacterial classification, quantification, and identification

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PLSDA			DFA		
E. COLI	True	False	E. COLI	True	False
Positive	95.65%	9.17%	Positive	89.63%	15.95%
Negative	90.83%	4.35%	Negative	84.05%	10.37%
STAPHYLOCOCCUS	True	False	STAPHYLOCOCCUS	True	False
Positive	54.05%	0.51%	Positive	86.49%	5.85%
Negative	99.49%	45.95%	Negative	94.15%	13.51%
STREPTOCOCCUS	True	False	STREPTOCOCCUS	True	False
Positive	95.59%	1.02%	Positive	99.26%	13.32%
Negative	98.98%	4.41%	Negative	88.68%	0.74%
MYCOBACTERIUM	True	False	MYCOBACTERIUM	True	False
Positive	88.31%	1.06%	Positive	96.10%	4.08%
Negative	98.94%	11.69%	Negative	95.92%	3.90%

DFA: Sensitivity: 91.37 ± 16.39 % Specificity: 97.46 ± 9.35 %
 PLSDA: Sensitivity: 93.13 ± 10.25 % Specificity: 90.60 ± 21.33 %

Bacteria

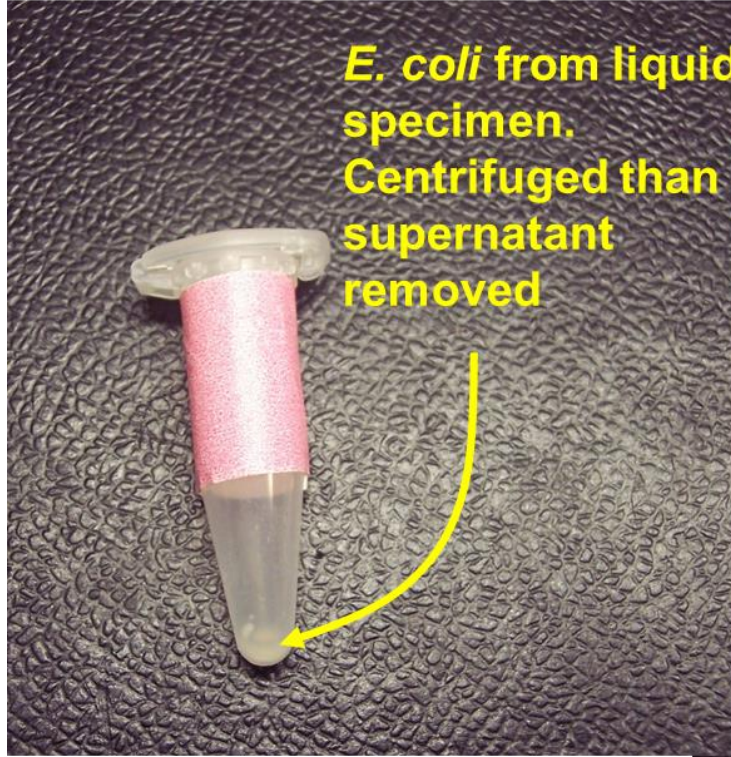


2011-2021

Can we translate this to be a convenient and easy-to-use (for clinicians) test?



Future

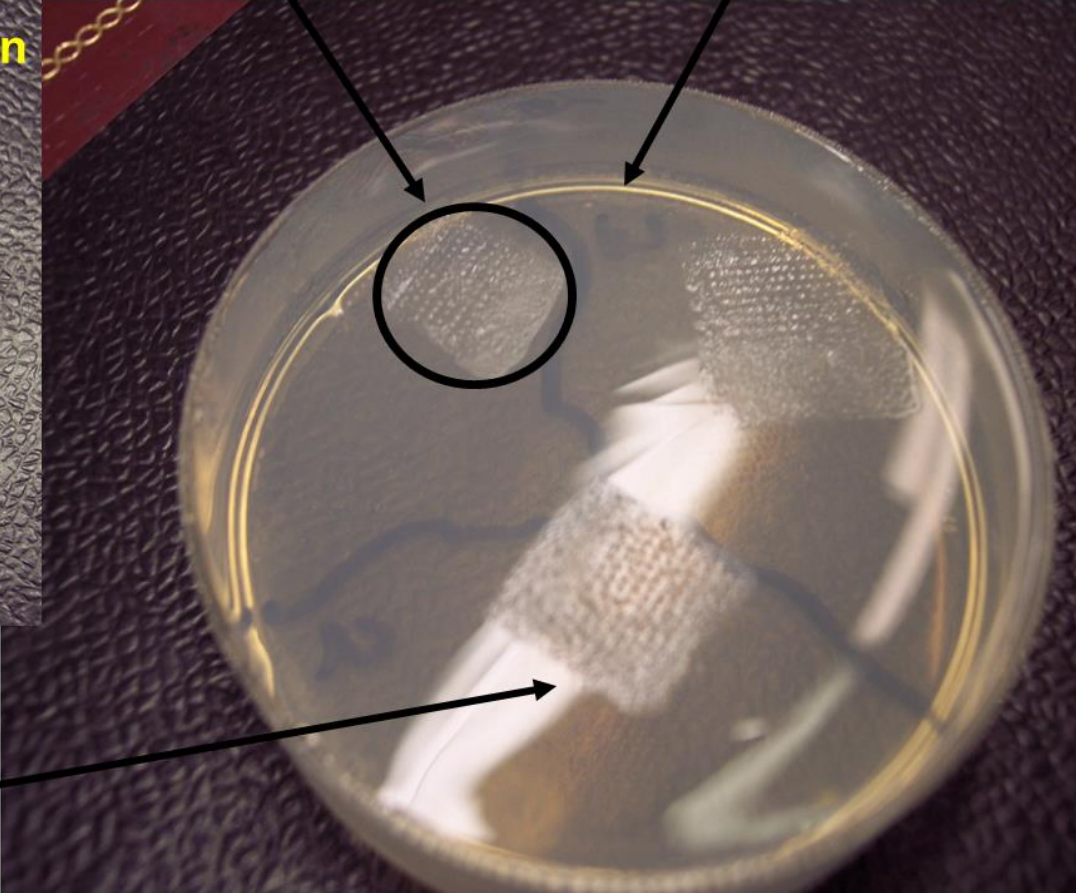


E. coli from liquid specimen. Centrifuged than supernatant removed

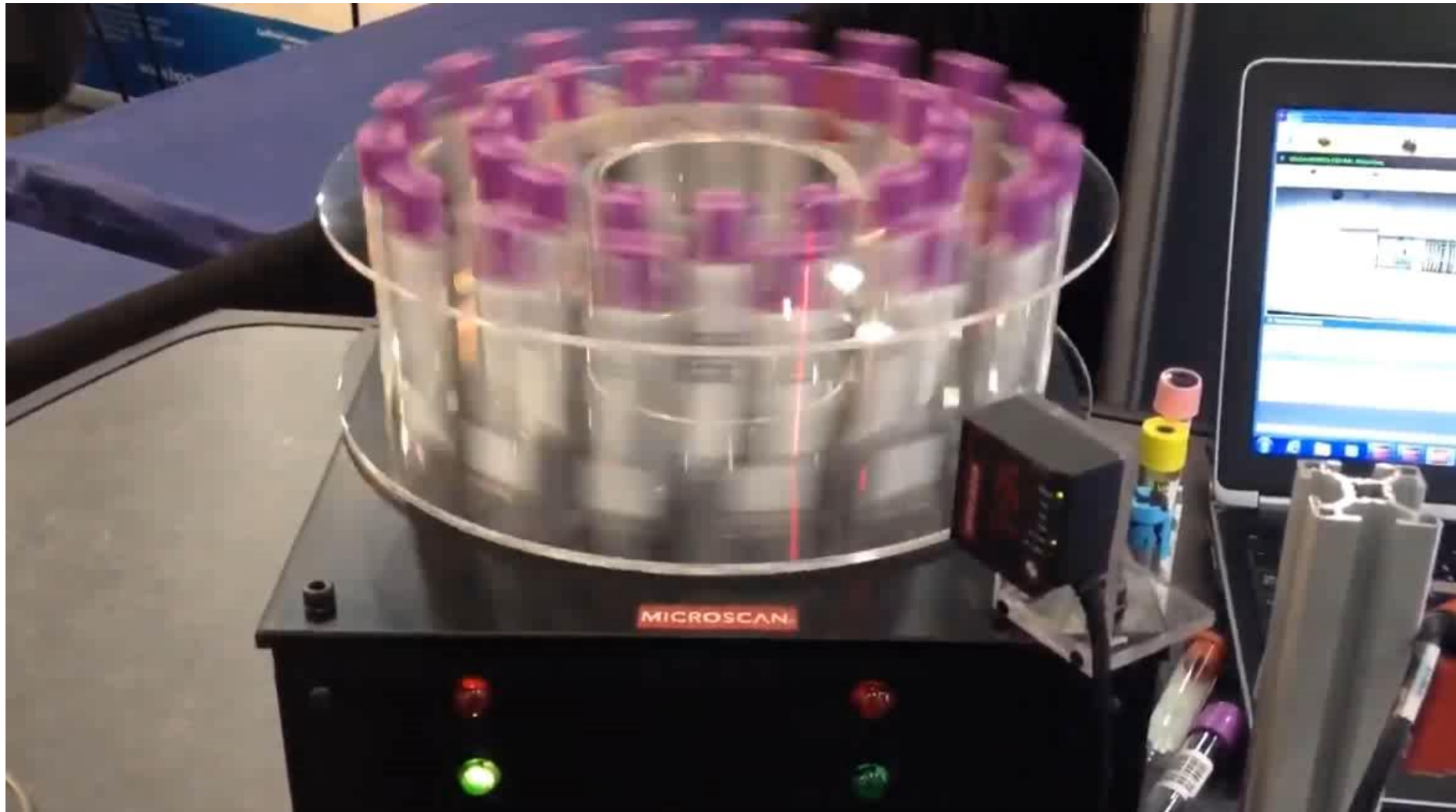
about 500-1500 bacteria per sampling location

10 microliter of bacteria pellet

bacto-agar (99% water)



What We Need

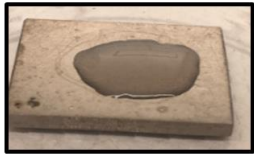


Our Approach

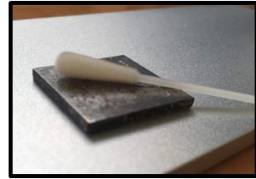


Simulating bacteria on a surface

Deposit 100 μL of bacteria suspended in DI water onto metal plate and heat for 2 min and 20 s.



Deposit 10 μL water onto swab tip and swab metal plate.



Place swab in centrifuge tube with 1 mL of water and vortex tube for 15 s.

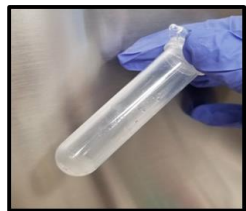


Place a 0.45 μm filter on the base piece. Connect the base. Place the cone in the insert.

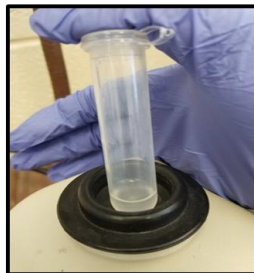


Simulating infection in clinical specimen

Pipette 100 μL of liquid contaminant containing bacteria suspension into a centrifuge tube.



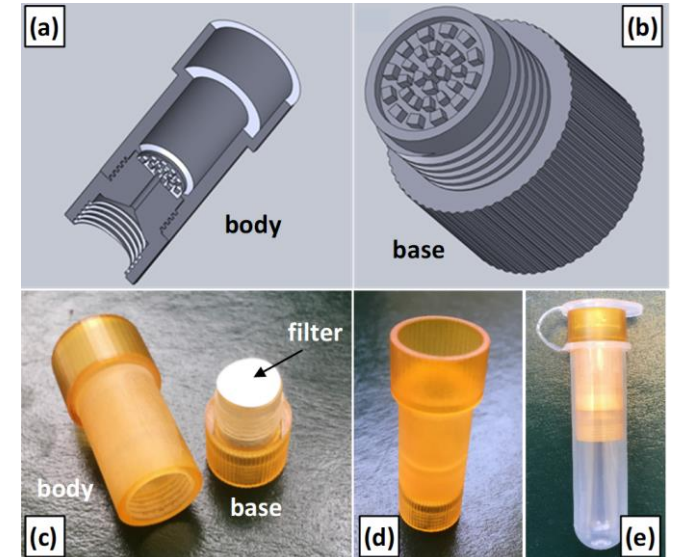
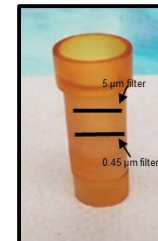
Add 1 mL water to the tube and vortex for 15 s.



Mount a 5 μm filter and a 0.45 μm filter on two separate insert base pieces.



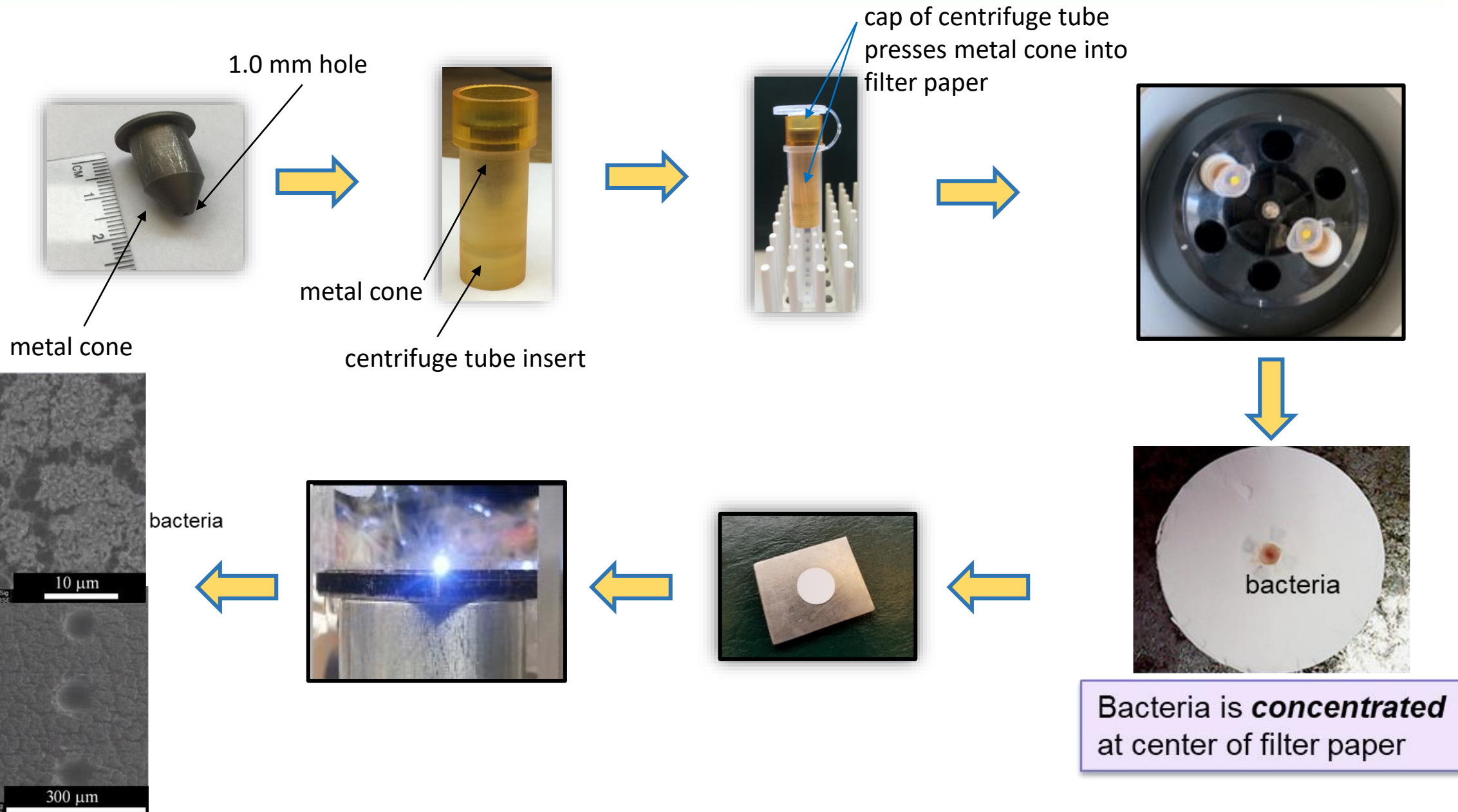
Connect both base pieces to the insert, with the 5 μm filter on top. Place the cone in the insert.



Bacterial LOD ~ 90 000 CFU per laser ablation event

These LOD's are *not clinically relevant*. Bacterial LOD with LIBS MUST be lowered.

Our Approach

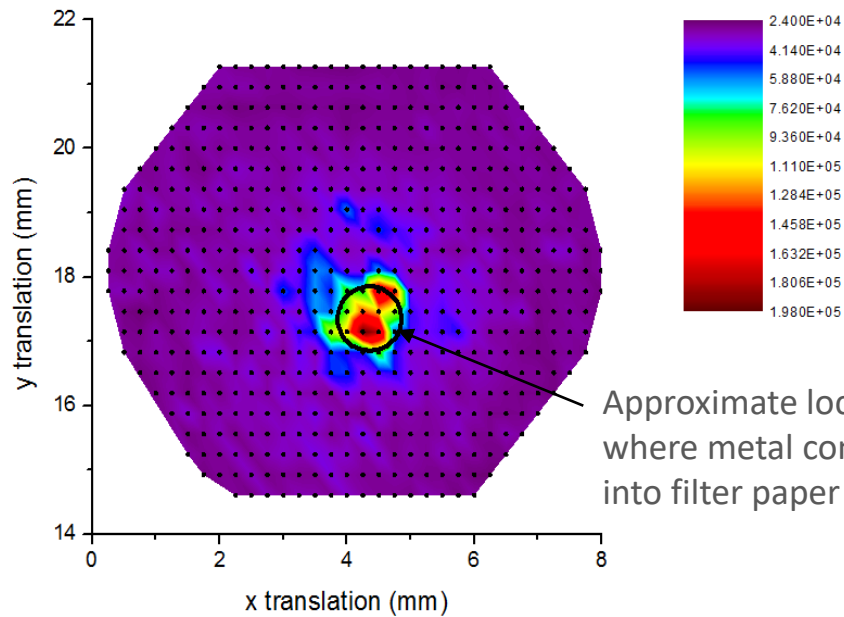


Results



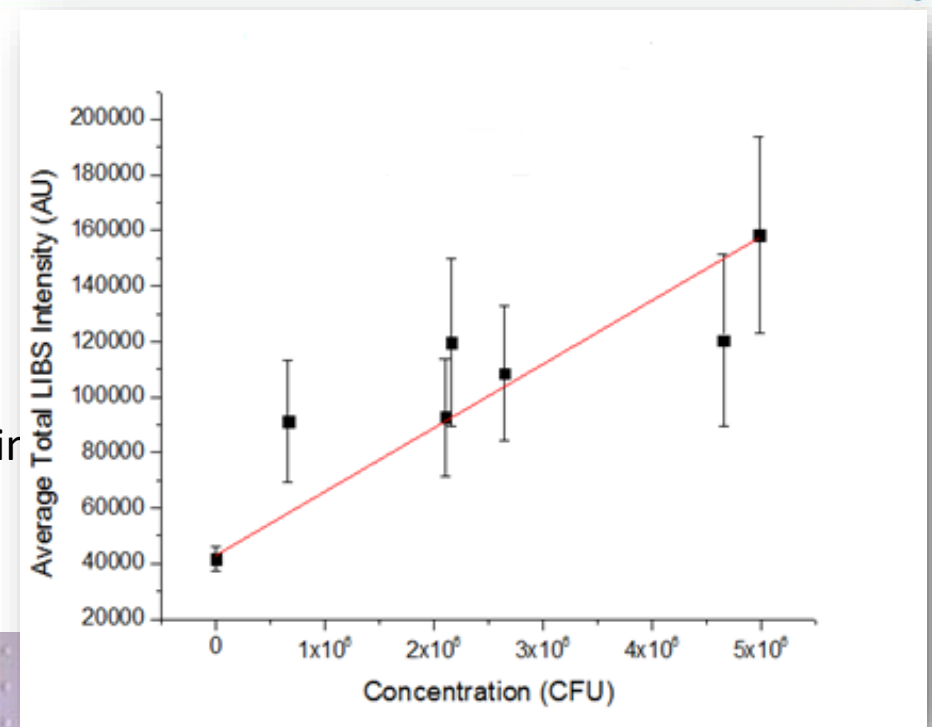
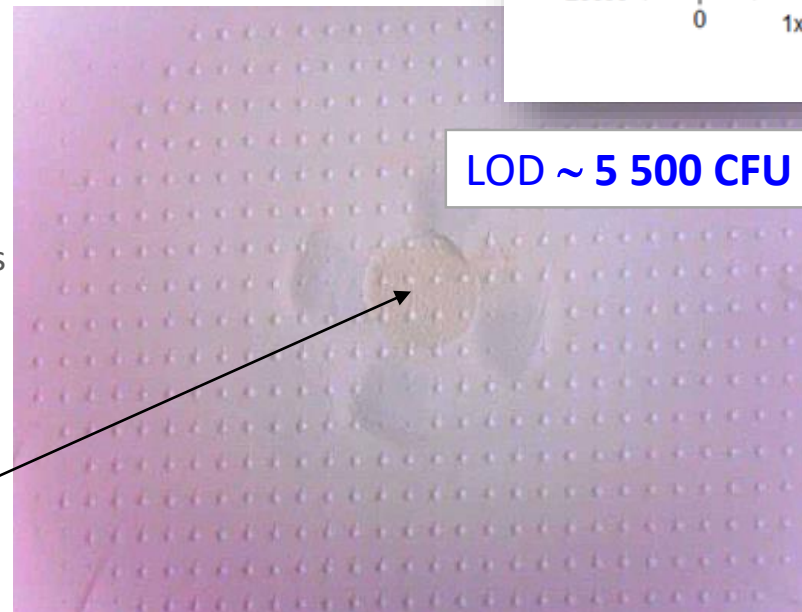
- *E. coli* deposited on filter paper with metal cone
- 569 LIBS spectra acquired across filter

Intensity map depicting bacterial deposition on filter paper for bacteria deposited with metal cone



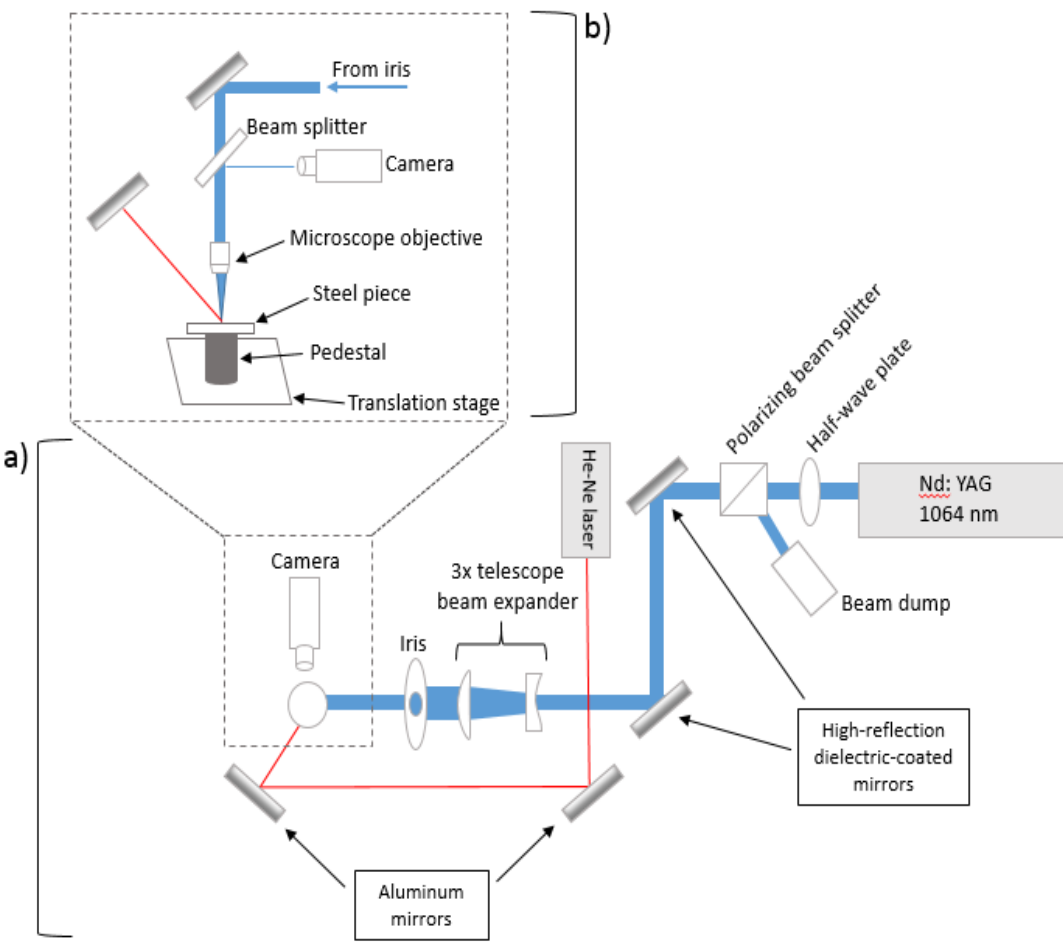
Approximate location of where metal cone presses into filter paper

Discolouration due to presence of bacteria

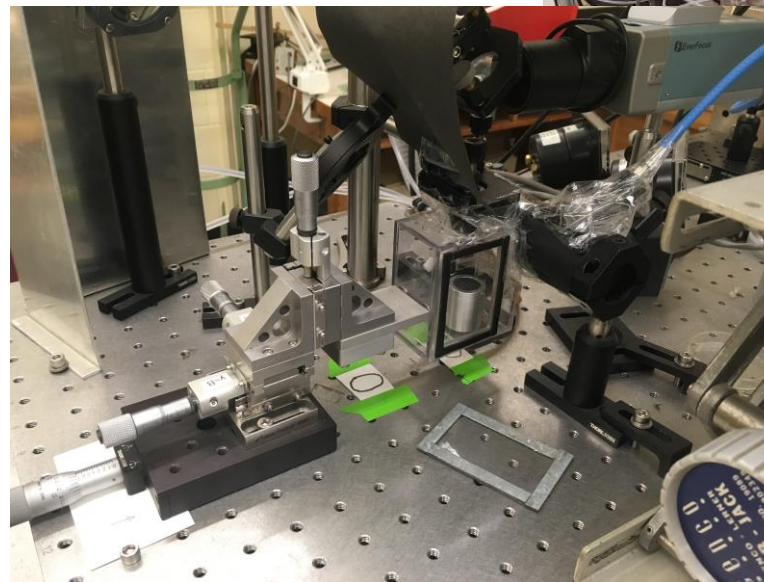
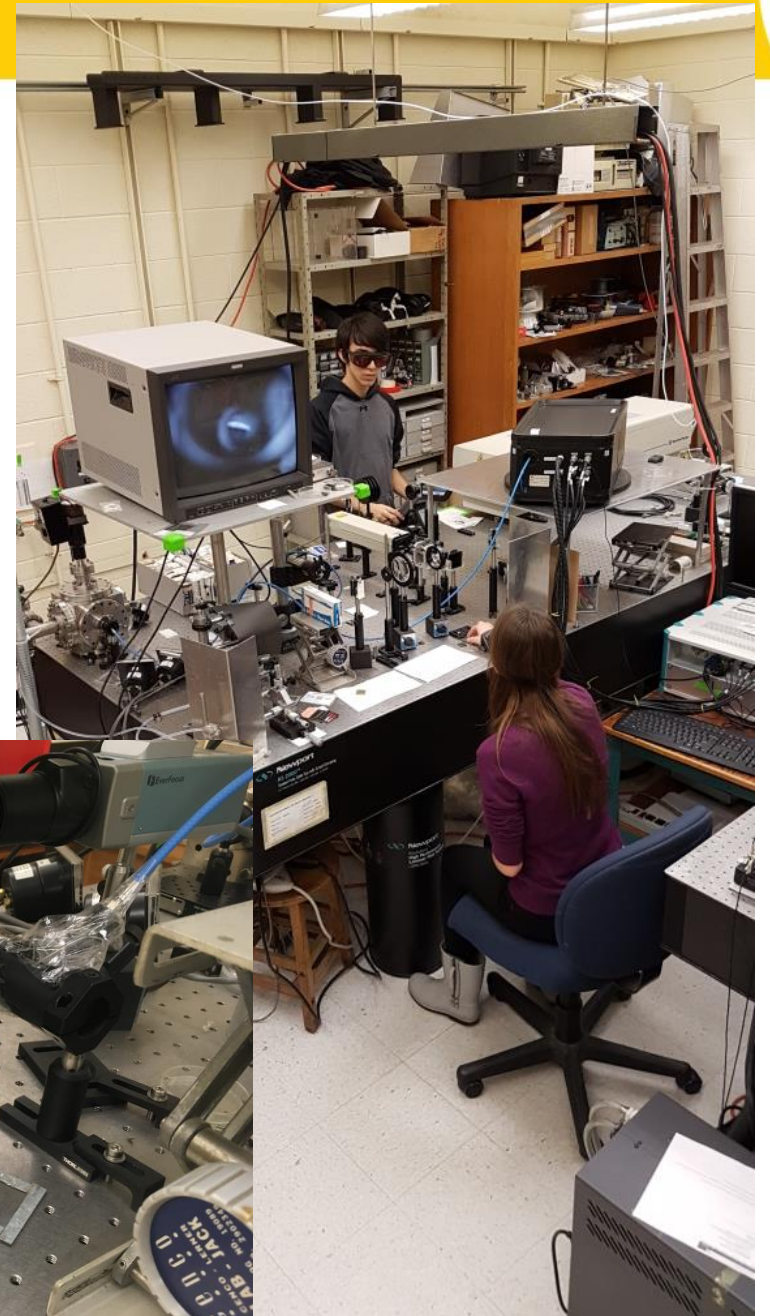


LOD ~ 5 500 CFU per laser ablation event

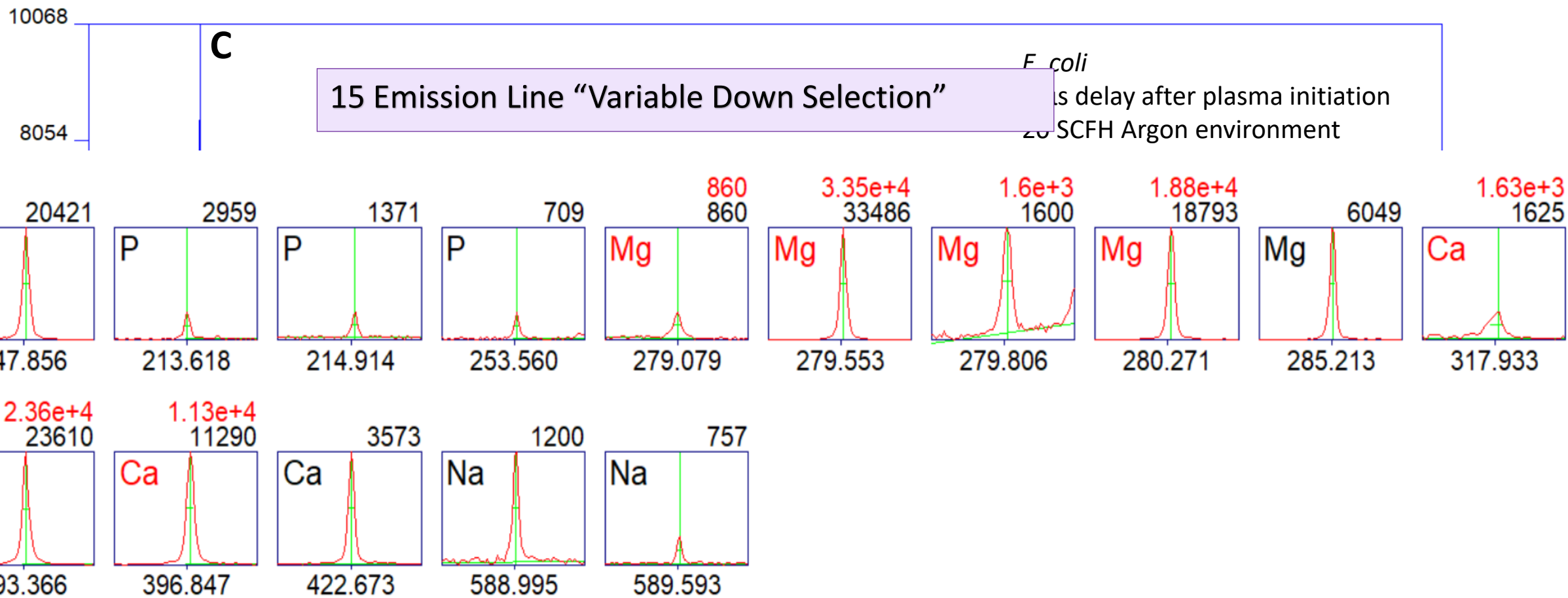
Apparatus



(a) Overhead schematic of the optical train used to direct laser pulses to the target. (b) Schematic side view of laser pulses emerging from the iris and directed to a target which is mounted on a steel piece

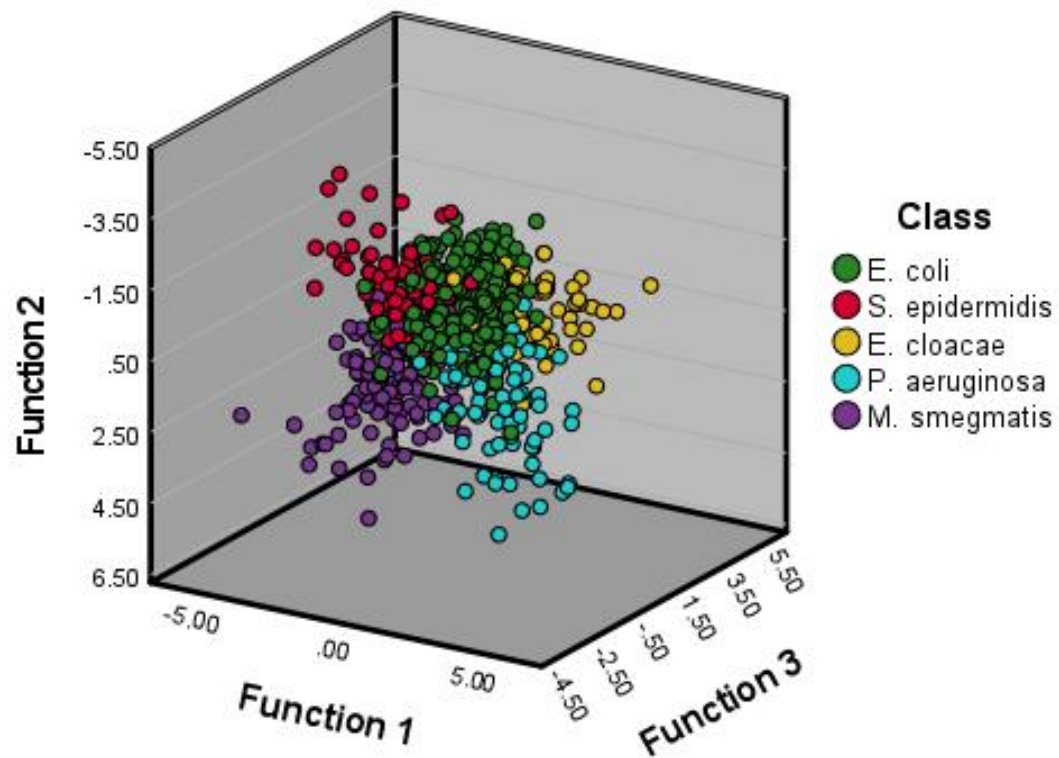


Typical LIBS Bacteria Spectrum

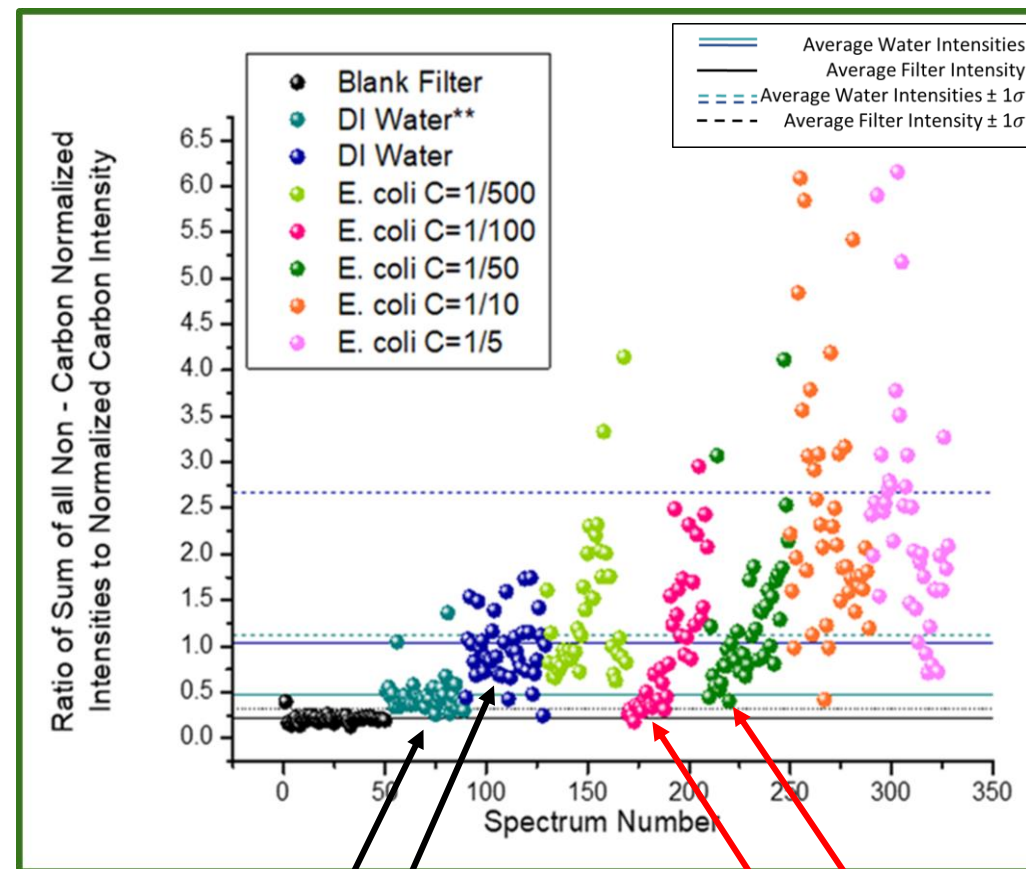




Canonical Discriminant Functions



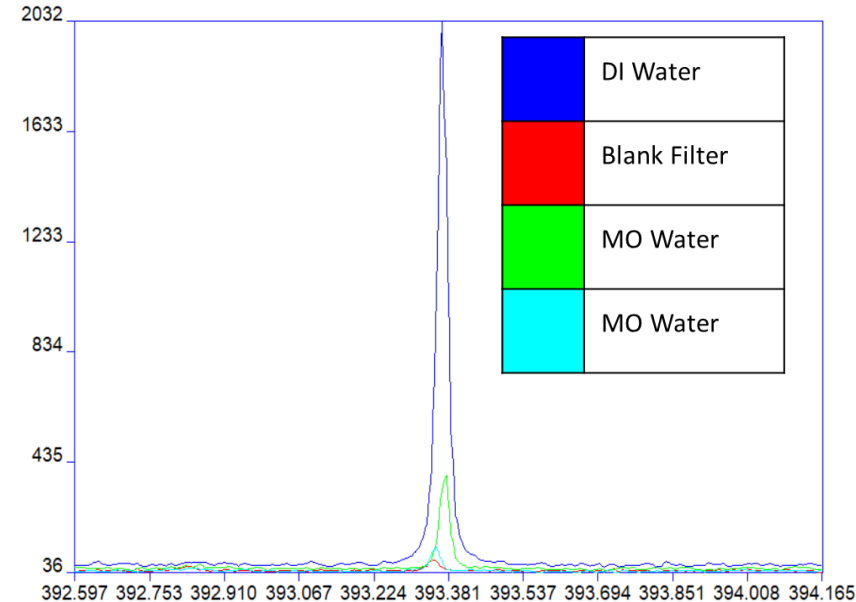
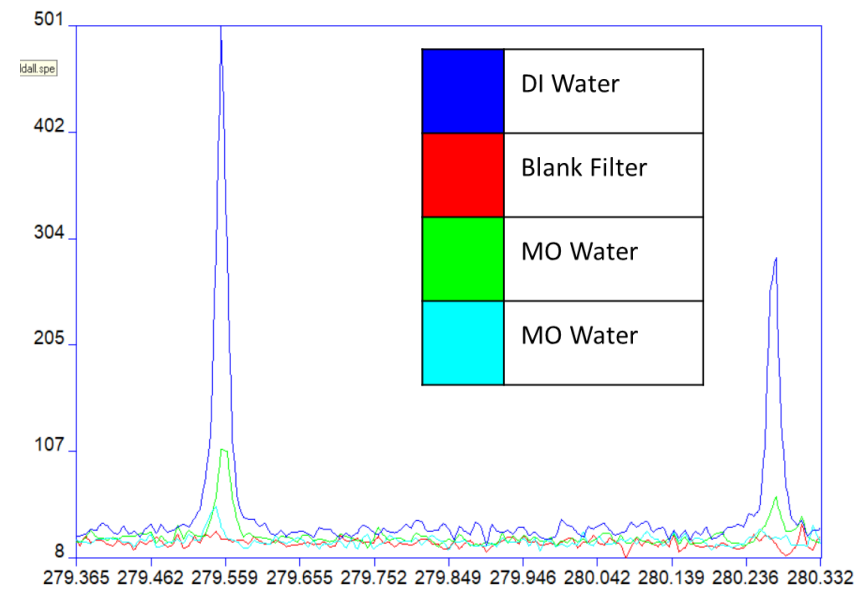
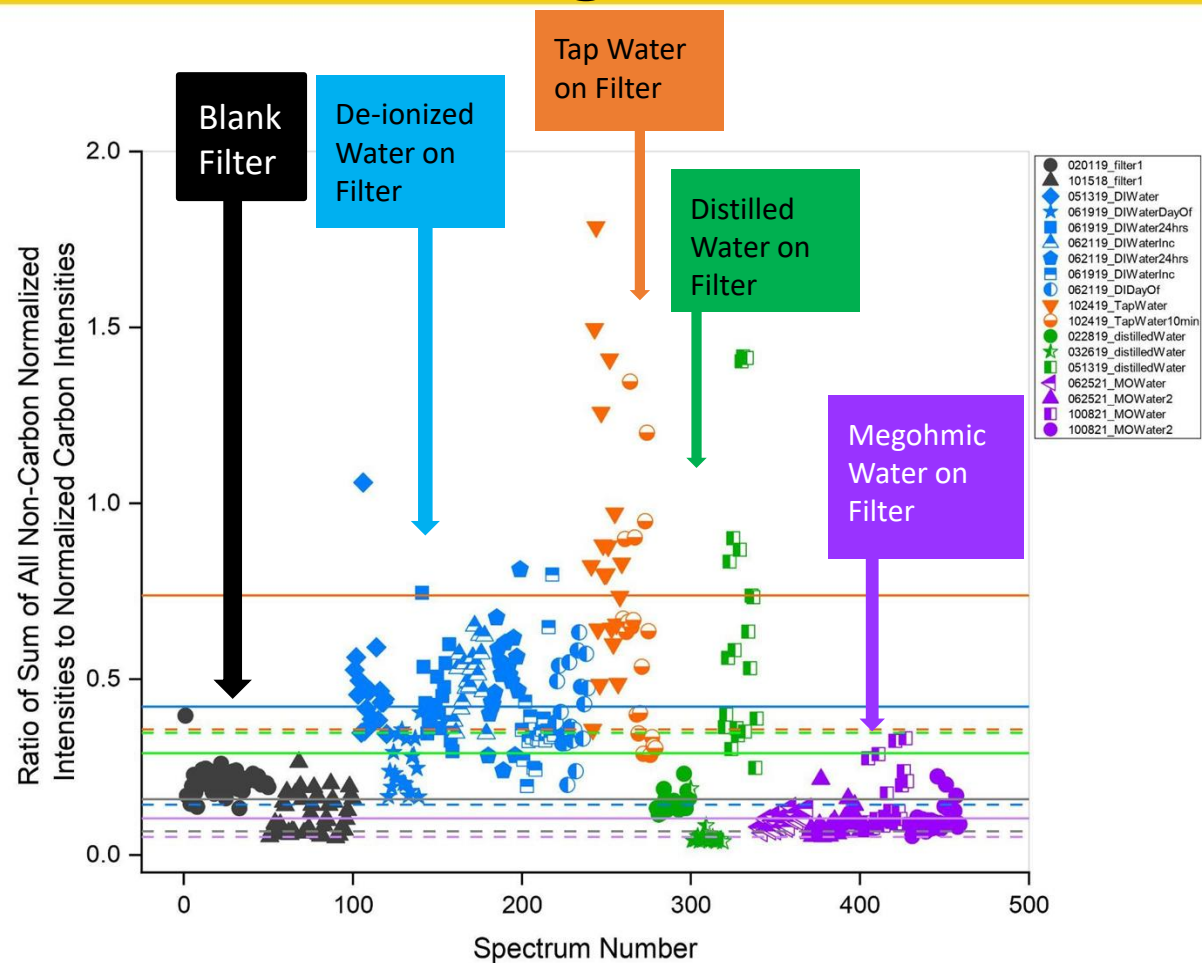
Bacteria	Sensitivity	Specificity	Classification error
<i>E. coli</i>	60 %	79 %	31 %
<i>S. epidermidis</i>	64 %	91 %	23 %
<i>E. cloacae</i>	50 %	91 %	29 %
<i>P. aeruginosa</i>	66 %	94 %	20 %
<i>M. smegmatis</i>	65 %	82 %	27 %



Why are sterile specimens (no bacteria) non-zero?

Why is this zero? What can we do about it?

To Reduce Background, Use Better Water

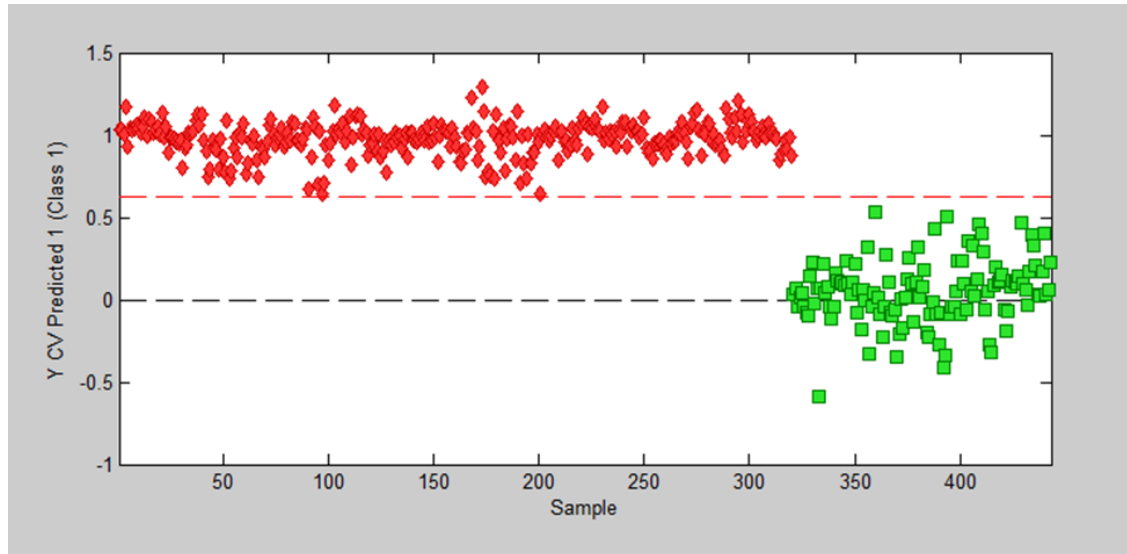


- Megohmic Water is consistent with filter
- Megohmic Water adds no extra background signal (as opposed to DI Water)

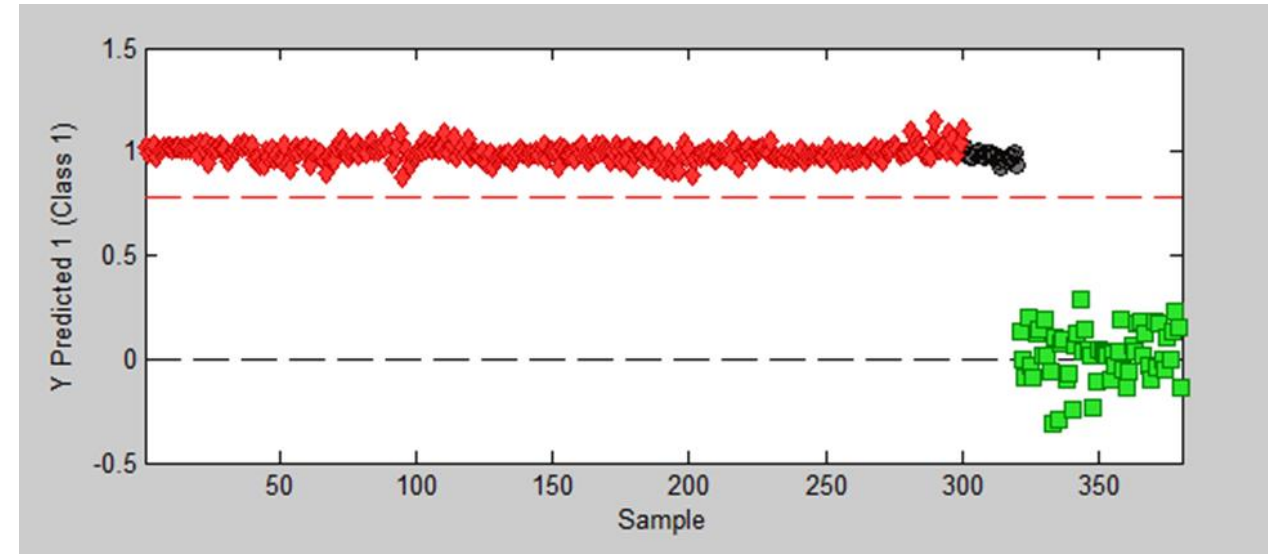
Water Studies (Bacterial Control)



PLS-DA: *E. coli* 1/5 vs. MO-Water



PLS-DA: All *E. coli* concentrations vs. MO-Water

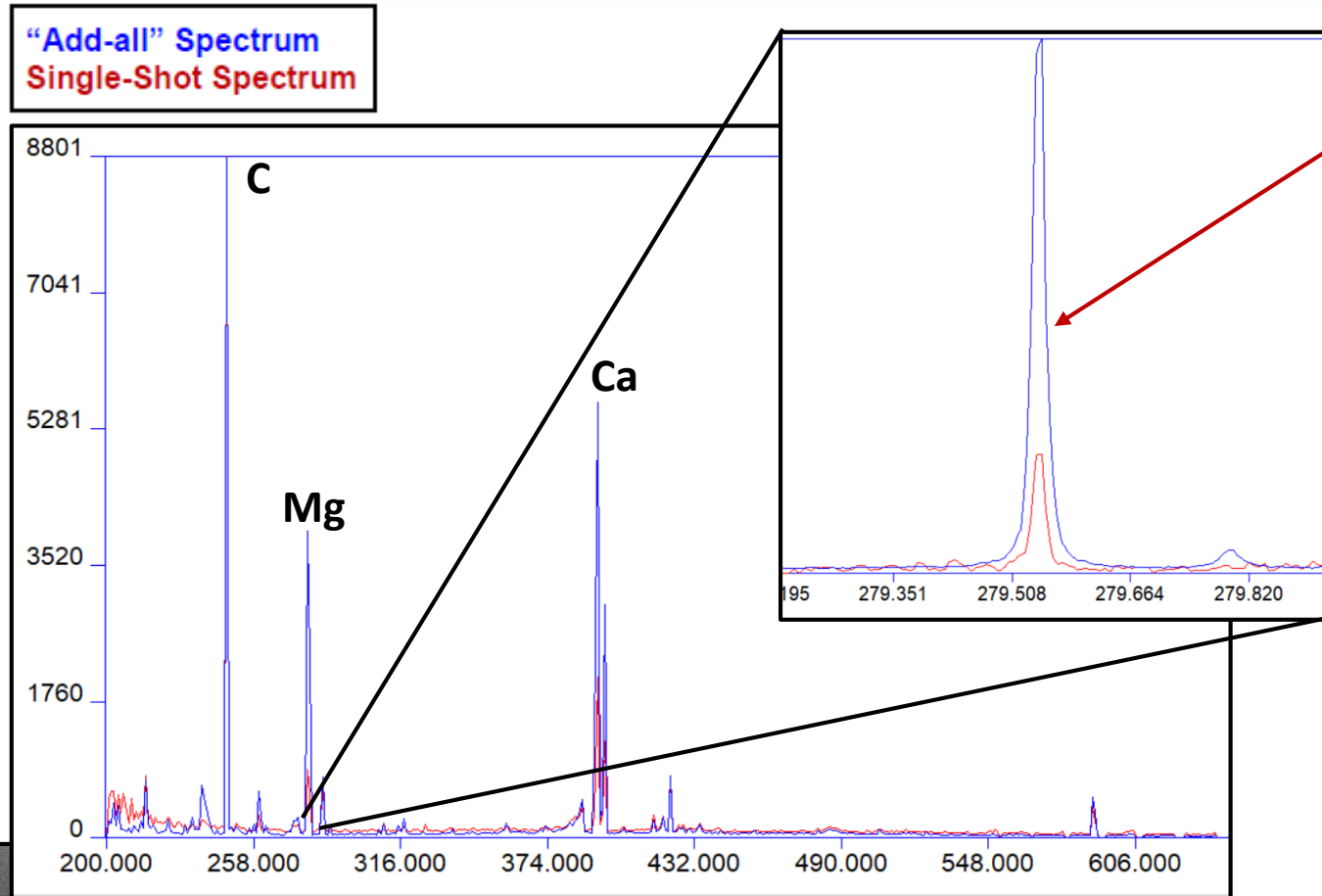


- Megohmic water allows for high discrimination accuracy for all bacterial species we work with
- Megohmic water is indistinguishable from filter background

Averaging / Summation of Shots to Eliminate “Empty” Shots

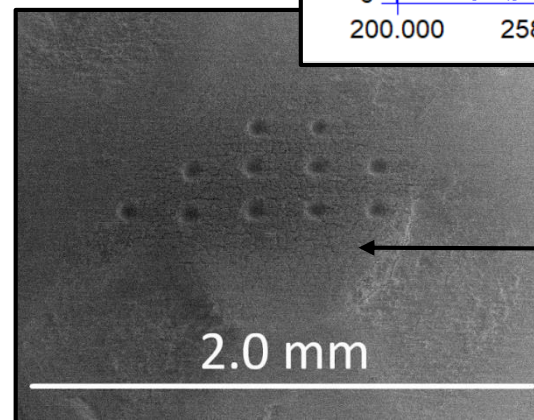


- We need lots of data “shots” to build our chemometric model, but we only need ONE measurement per filter (patient sample)
- We can sum all the spectra using the “add all” feature of the spectrometer.
- Eliminates many “empty” spectra, but then data is very slow to accumulate and publish.



Magnesium
279 nm line
has greater
intensity

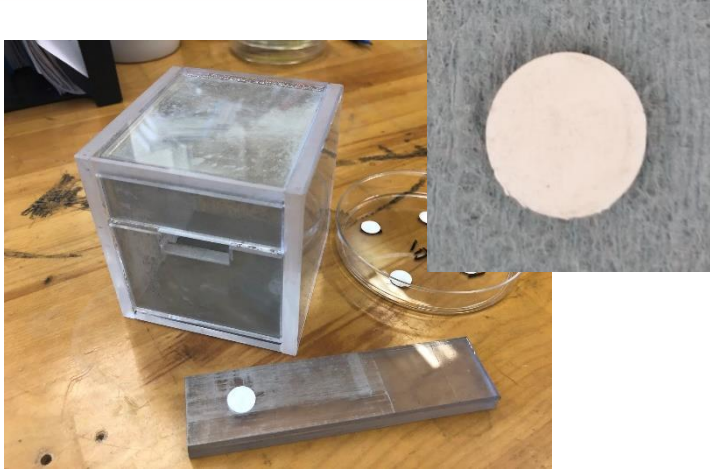
The Add-all
spectra
increases the
signal to noise
ratio



Circular area of filter
containing 10
single-shot spectra.

2.0 mm

Using silver to amplify the LIBS spectrum: microparticles



Method for filter preparation

0.5 - 1 micron spherical silver (99.9%) powder:

Trace uniform spread (with custom chamber) *approx. 1 – 50 ng deposited*

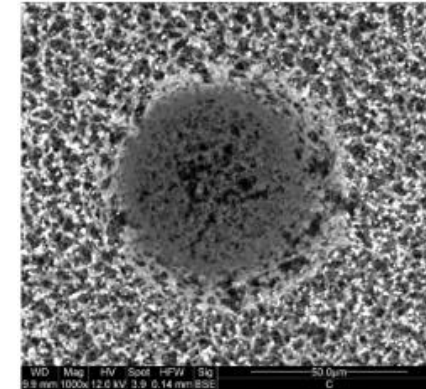
Surface coverage density rate $2.6 \times 10^{-8} \mu\text{g}/\mu\text{m}^2 \cdot \text{s}$

110 pg of Ag ablated per shot

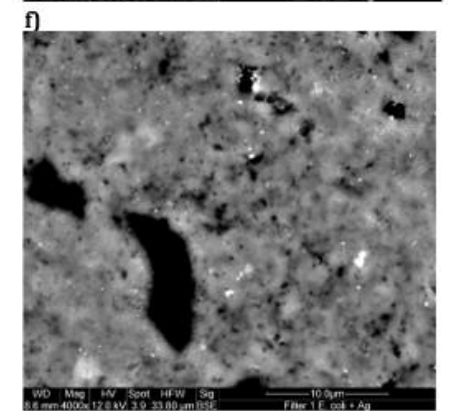
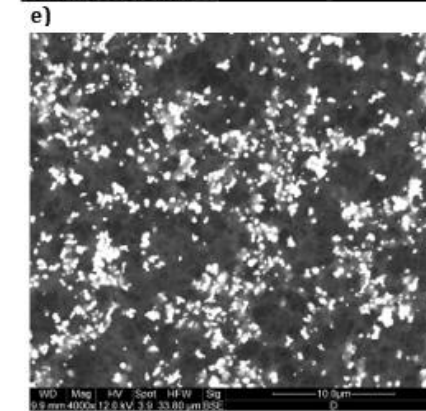
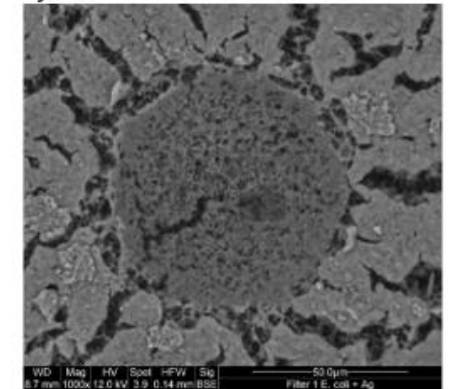
Average Elemental Enhancement of 4 Bacteria Species with the Addition of Silver Microparticles

Bacteria Species	Elemental LIBS Spectral Emission Enhancement				
	C	P	Mg	Ca	Na
<i>Escherichia coli</i>	1.3	4.6	3.9	5.4	3.9
<i>Mycobacterium smegmatis</i>	1.2	1.7	2.7	8.4	6.7
<i>Pseudomonas aeruginosa</i>	1.3	1.1	6.9	27.3	1.0
<i>Enterobacter cloacae</i>	1.2	4.4	6.9	2.2	1.3

c) Silver on filter



d) Silver on filter under bacteria

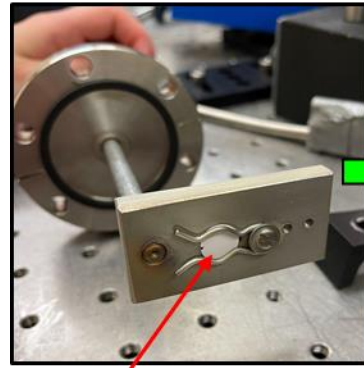


Using silver to amplify the LIBS spectrum: Ag sputtered filters

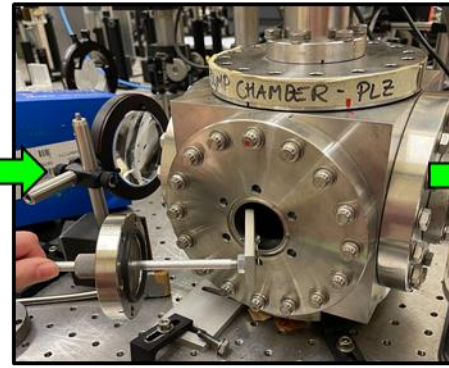


Method for filter preparation

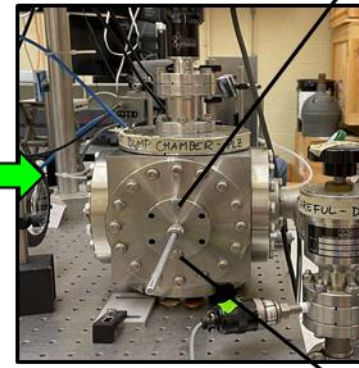
- 99.9% pure Ag foil on rotating target
- 1064 nm pulsed laser sputtering (*approx. 60 mJ / pulse*)
- Vacuum environment
- Blank filters places in various geometries around sputtering source
- Sputtering time variable (*20 s – 12 min*)



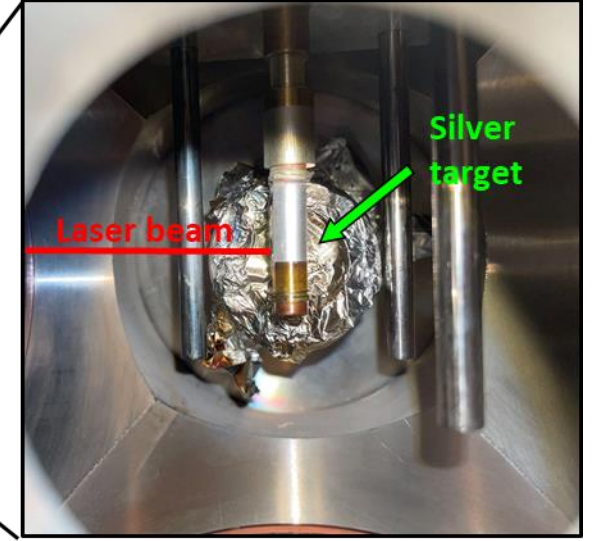
Filter on custom **aluminum filter holder**



Filter holder being inserted into **10 mTorr evacuated chamber**

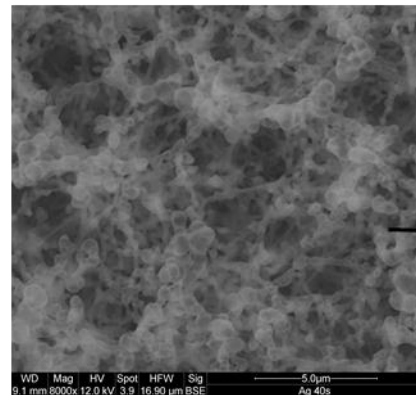


Closed vacuum chamber apparatus

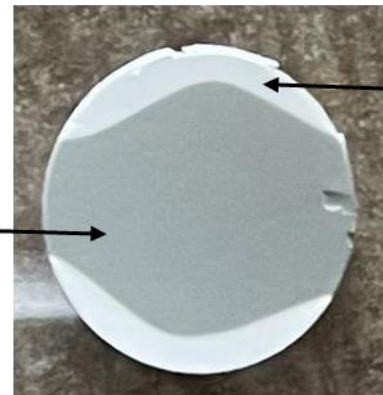


Inside the chamber

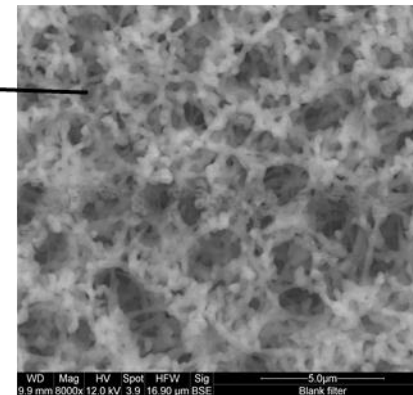
The deposition is so uniform and thin it appears the same as a blank filter



Filter with 40s Ag

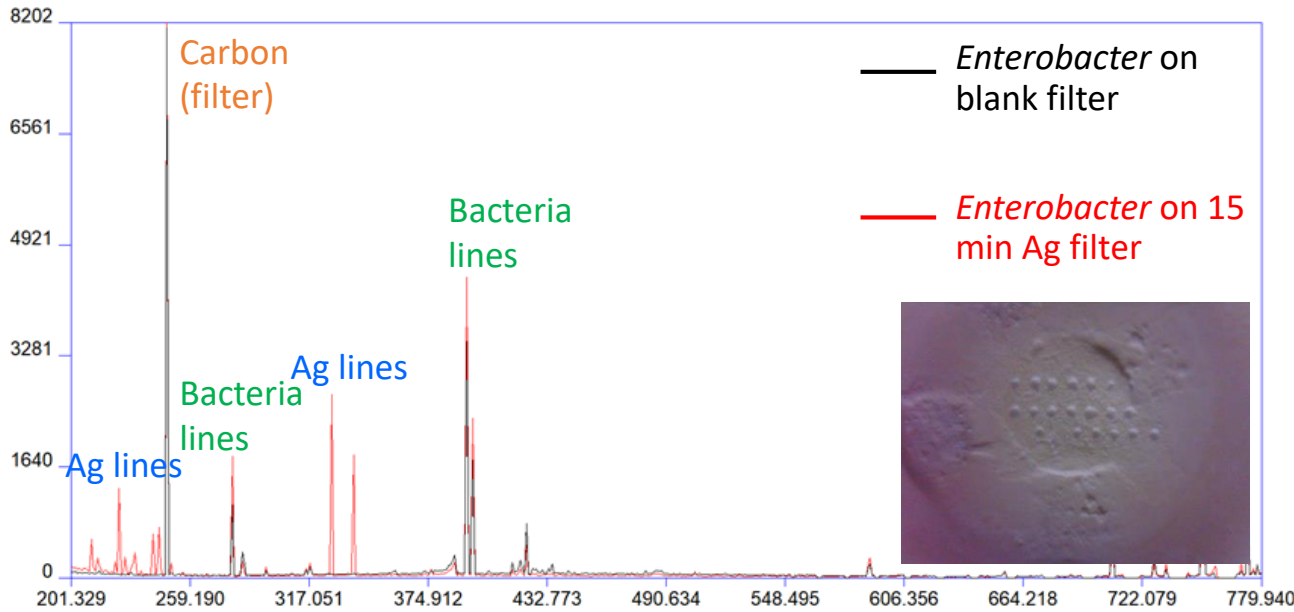


40 s deposition time Ag filter

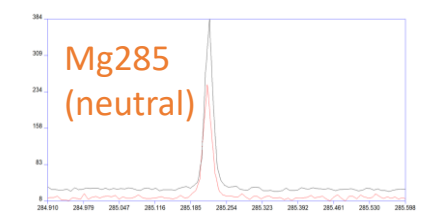
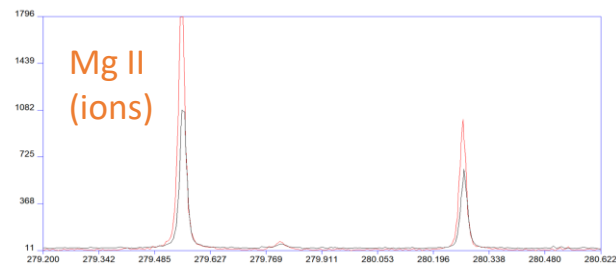
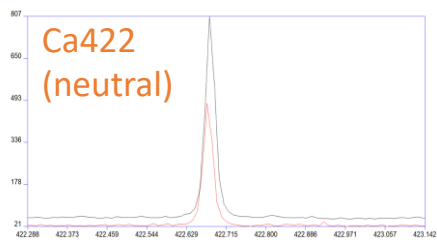
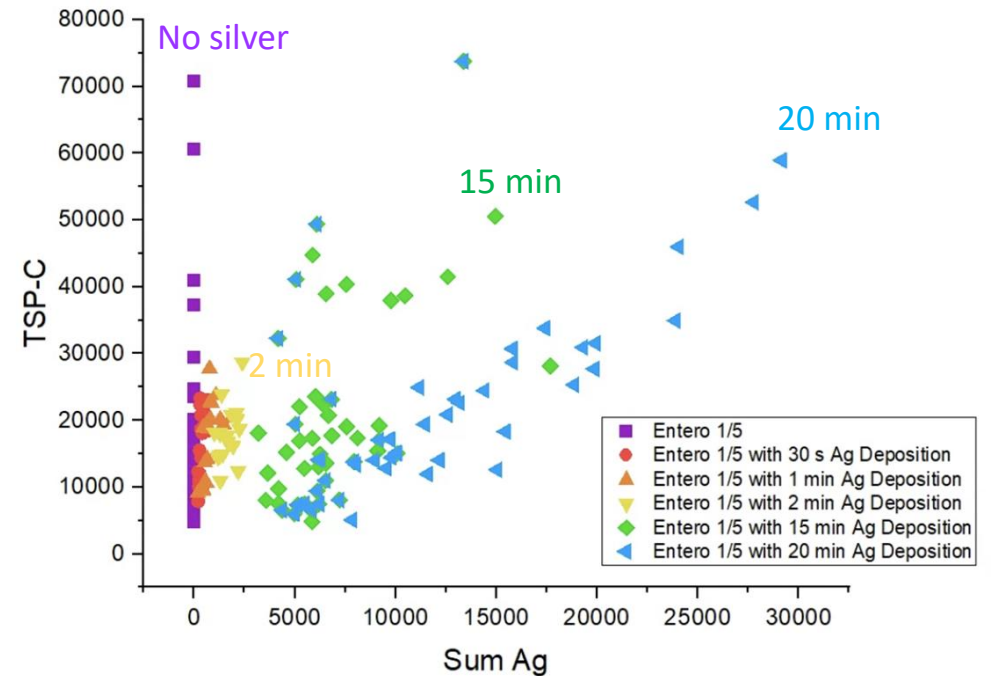


Blank Filter

Using silver to amplify the LIBS spectrum: Ag sputtered filters

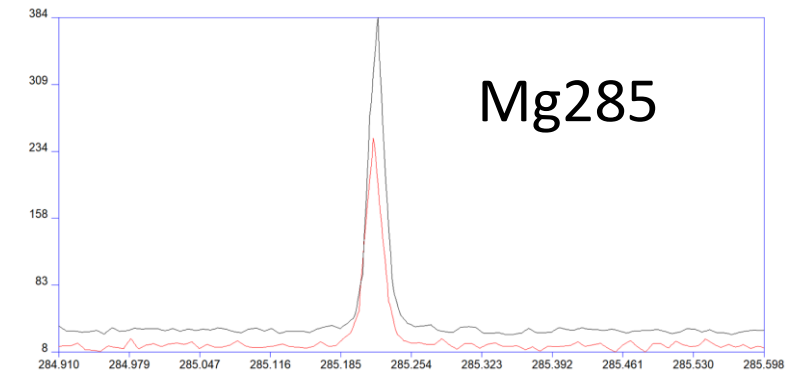
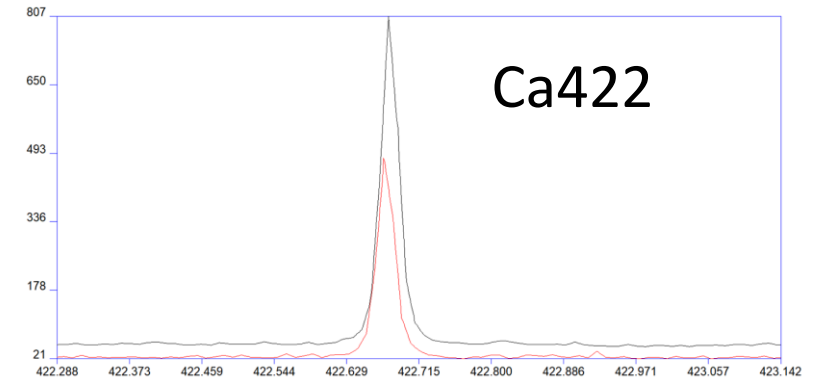
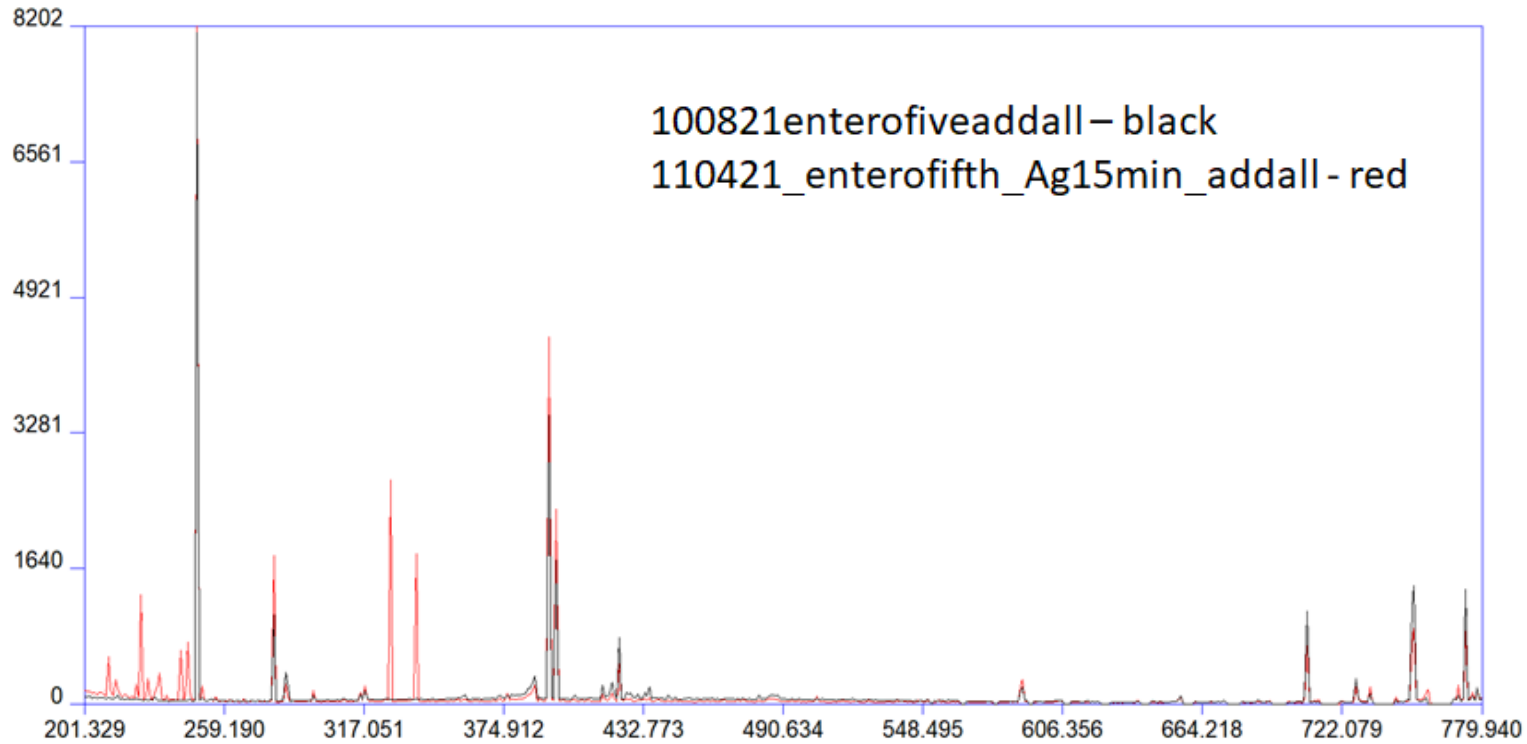


- Bacterial lines strongly correlate with Ag lines
- But they are not enhanced relative to use of “blank filter” (purple)

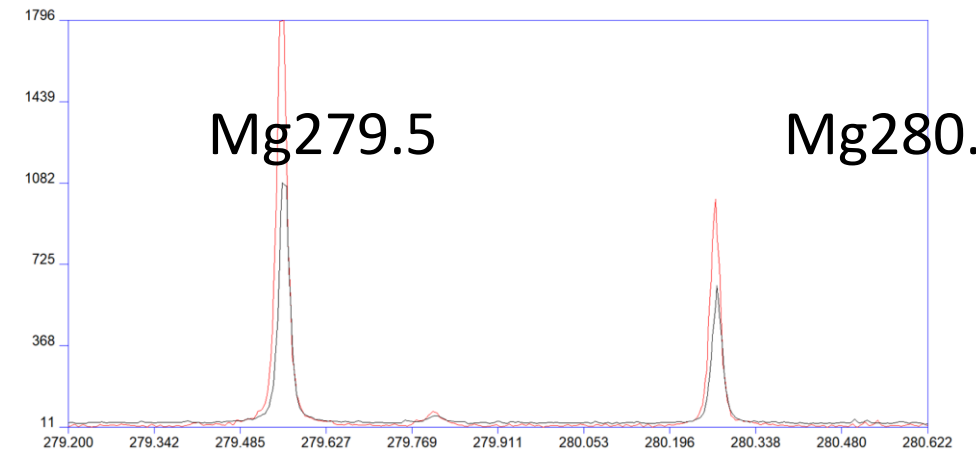


- Plasma temperature is increased.
- Enhancement was not observed until 15 min deposition time.
- 20 min gave the same amount of enhancement as 15 min.

The x-axis is the sum of all Ag lines and the y is sum of all lines other than silver excluding carbon.

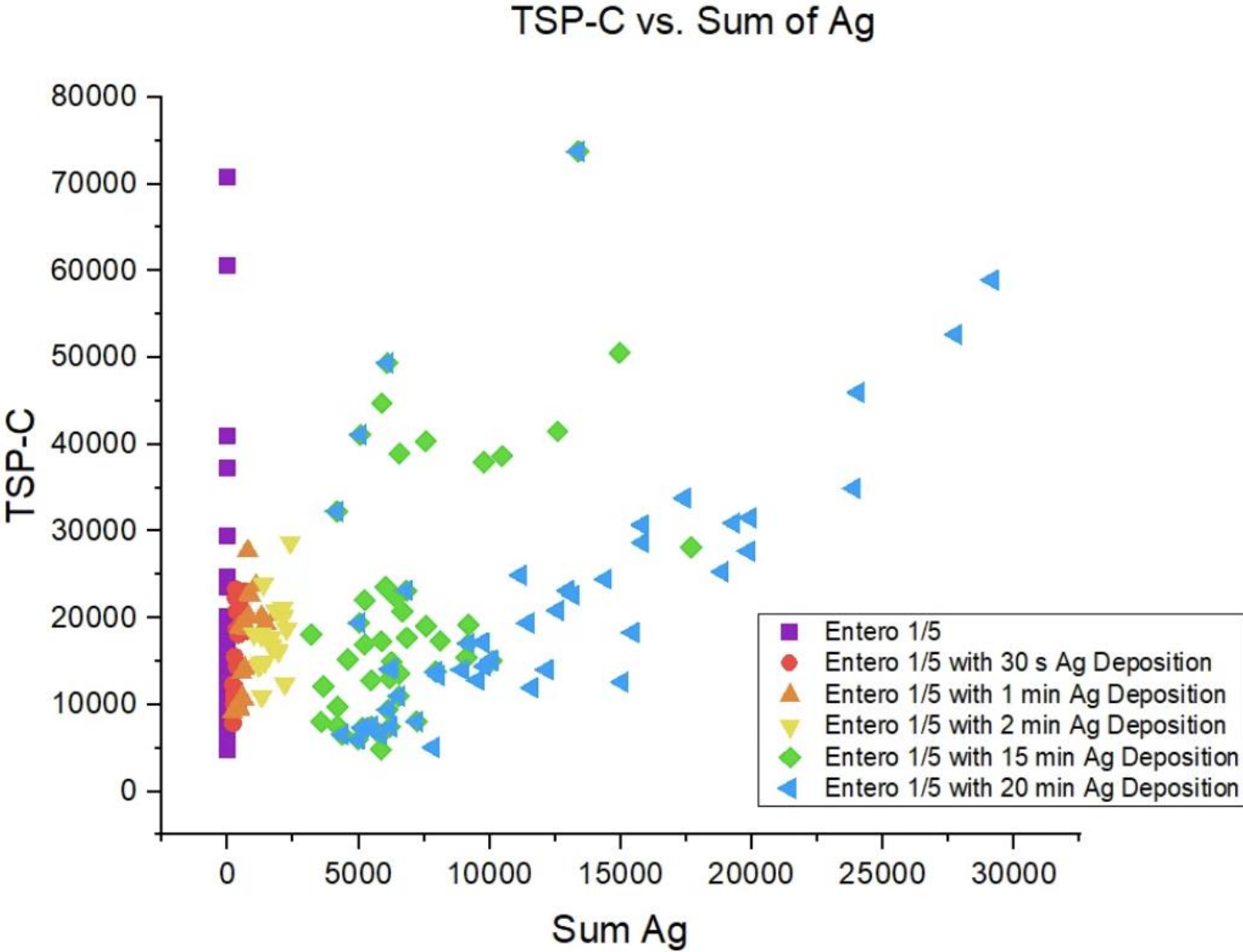


Intensity is enhanced for all lines except C247, which is the same, and Mg 285 and Ca422 which are lower because they are neutrals. This proves that our plasma temperature is increased. Enhancement was not observed until 15 min deposition time. 20 min gave the same amount of



Silver Enhancement vs. TSP-C

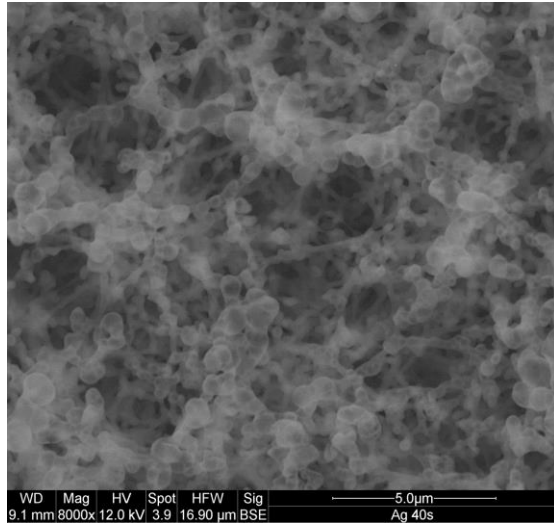
The x-axis is the sum of all ag lines and the y is sum of all lines other than silver except for carbon.



Using silver to amplify the LIBS spectrum: nanoparticles

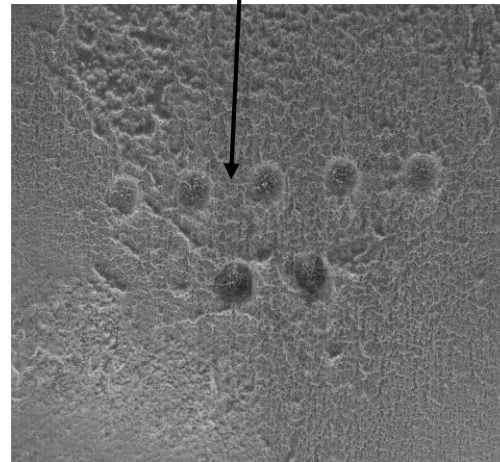
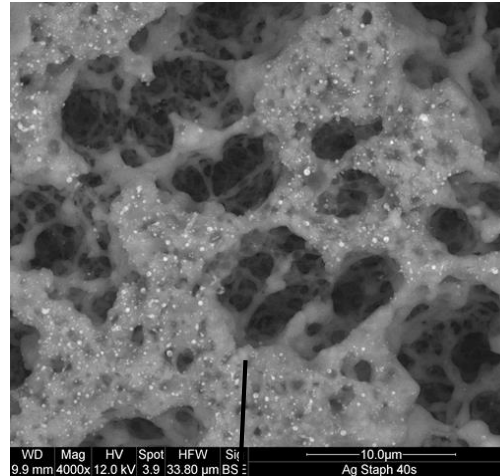


Silver is present, but not visible



Filter with 40s Ag

White specks = Silver nanoparticles



Filter with 40s Ag, then *Staphylococcus* deposited on top

Sputter deposition in vacuum produces no nanoparticles.

LIBS on the sputter targets DOES produce nanoparticles.

Subsequent shots on areas coated with nanoparticles shows enhancement (NPELIBS).

Using silver to “boost” signals



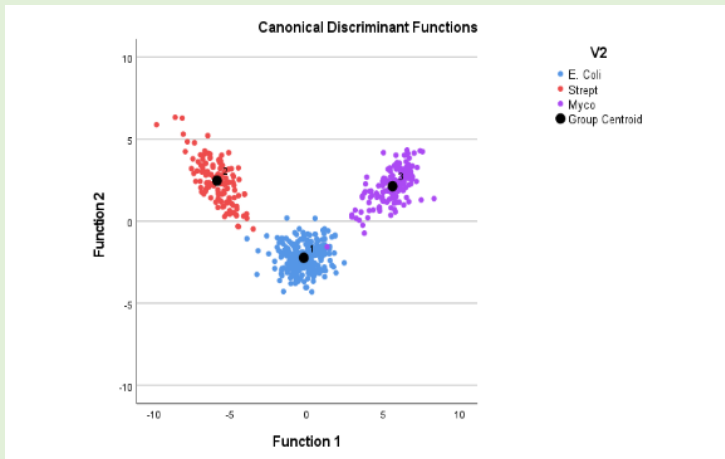
- We might get rid of carbon



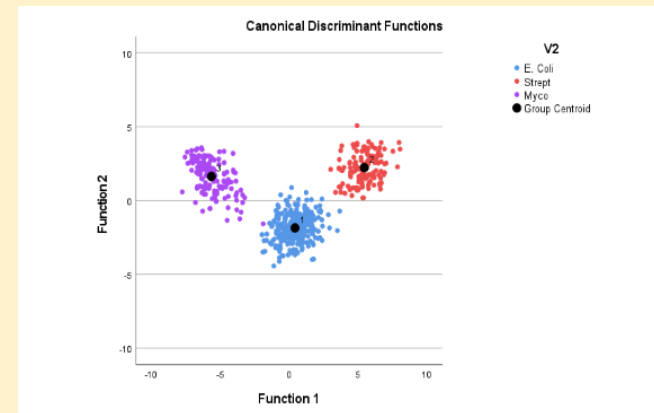
We Are Also...

- Investigating the removal of the C247 line from all analysis (most of all the carbon comes from the filter)
 - Preliminary results show this will not significantly reduce sensitivity or specificity

Includes carbon line and three bacteria classes



Carbon line excluded and three bacteria classes



All results obtained from analysis of previously-obtained data (not from filters)

Sensitivity and Specificity results:

E. Coli		Myco	
Sensitivity with Carbon:	0.983277	Sensitivity with Carbon:	0.902597
Specificity with Carbon:	0.944827	Specificity with Carbon:	0.990804
Strept			
Sensitivity with Carbon:	0.992647		
Specificity with Carbon:	0.997792		

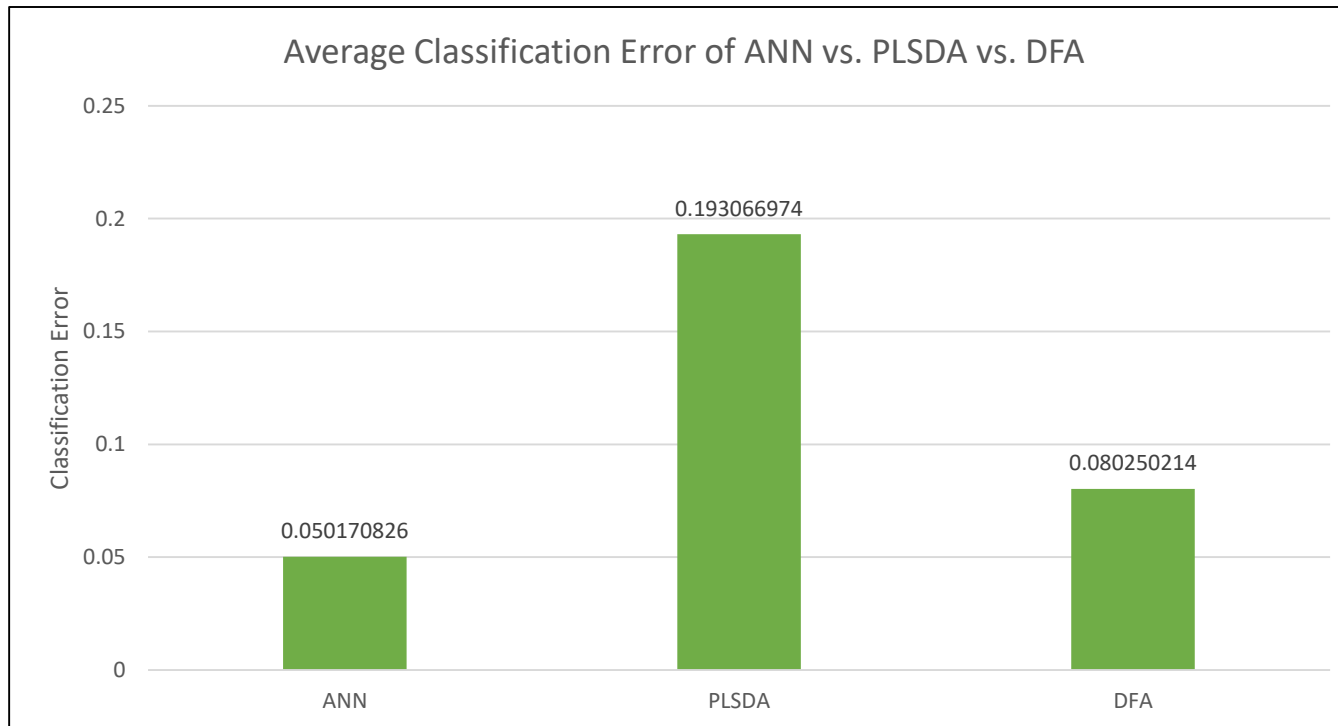
Sensitivity and Specificity results:

E. Coli		Myco	
Sensitivity:	0.969899	Sensitivity:	0.896103
Specificity:	0.931034	Specificity:	1
Strept			
Sensitivity:	0.970588		
Specificity:	0.980132		

We Are Also...



- Investigating the use of ANN* to replace PLS-DA or DFA
 - Most groups have shown improved performance when using SVM, ANN, or KNN as opposed to DFA, LDA, PLS-DA
 - We need much more training data to properly develop this model (number of epochs, number of nodes in hidden layer, use of validation, batch size, etc.)



Using our previously obtained data (not on filter)

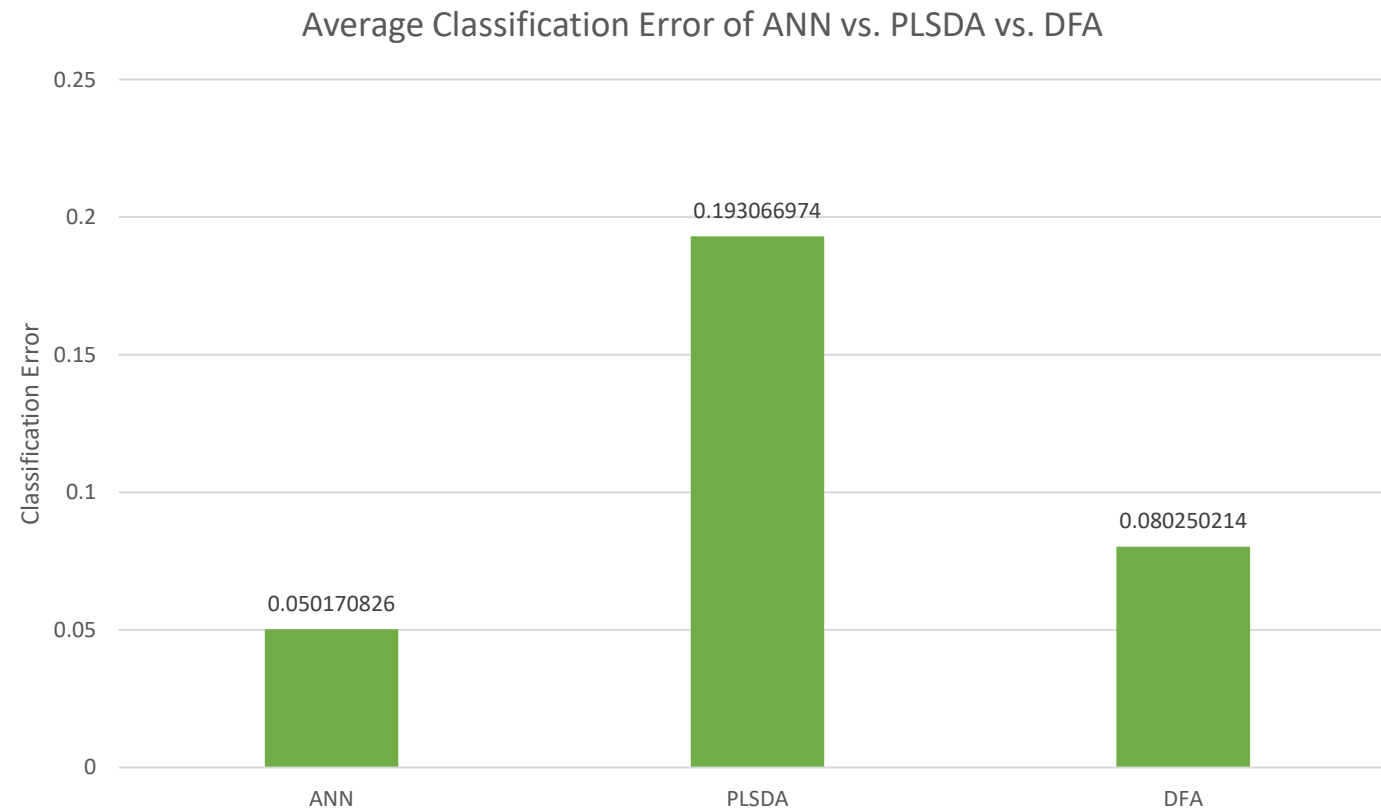
5 Class test

- Escherichia
- Enterobacter
- Staphylococcus
- Streptococcus
- Mycobacterium

*Built an artificial neural network (ANN) in python using Keras which acts as an interface of Tensorflow (open-source platform for machine learning)



- Using silver





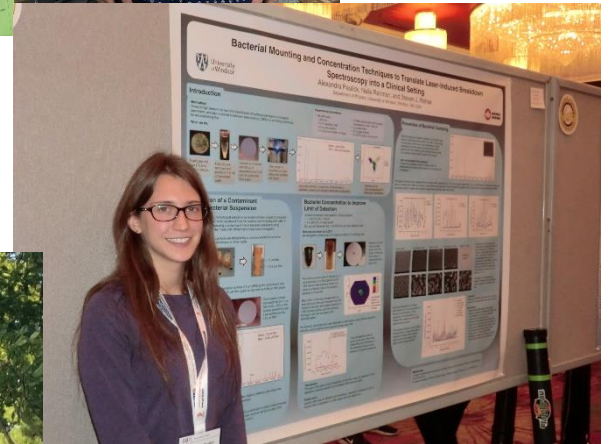
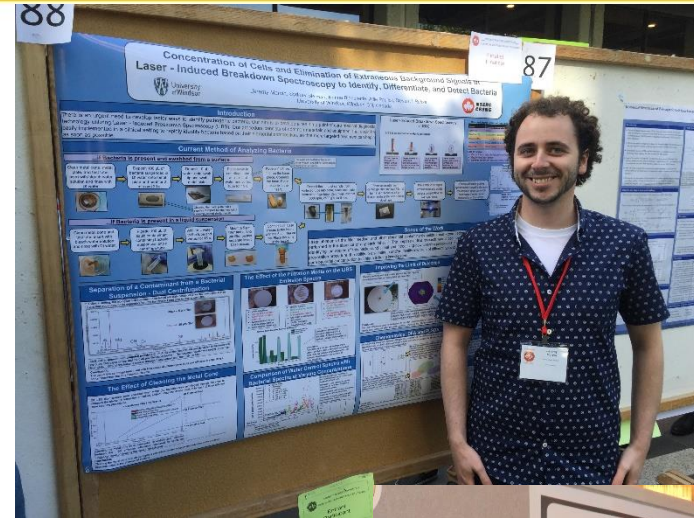
- We have a very flexible, rapid way to deposit bacteria on a testable disposable filter
- Use of ultra-pure water improves specificity
- The modification of filters with metallic silver shows promise
- The removal of carbon from the elemental analysis may improve performance
- The use of ANN may improve performance
- We are now beginning to test blood and urine clinical specimens obtained from our clinical collaborators at Windsor Region hospital.

Thanks To

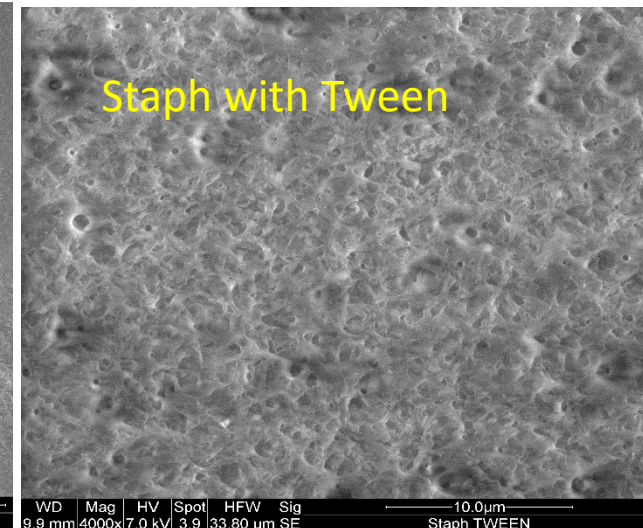
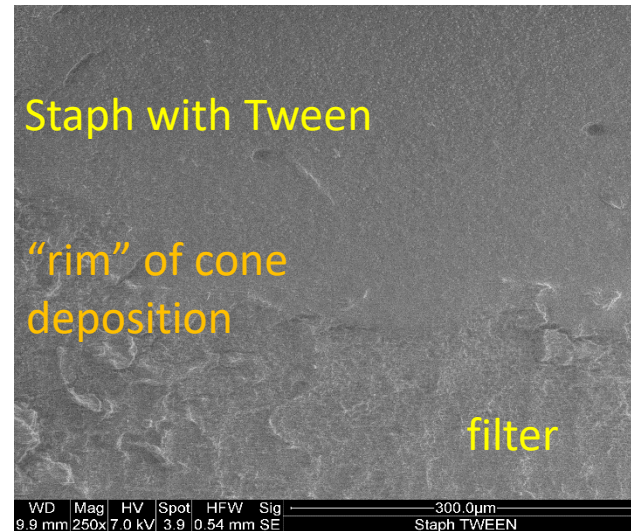
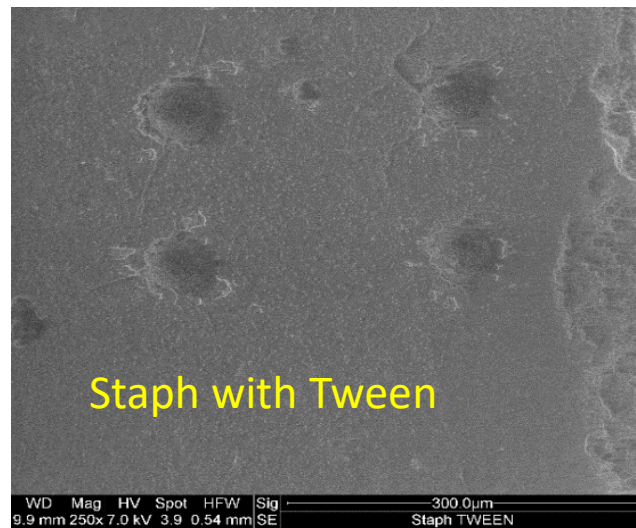
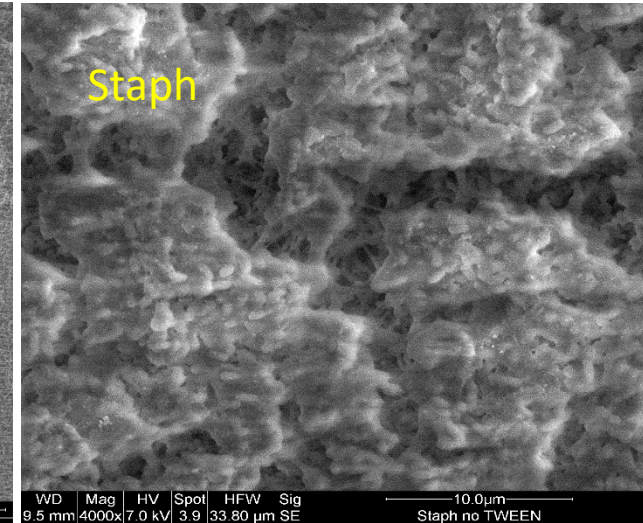
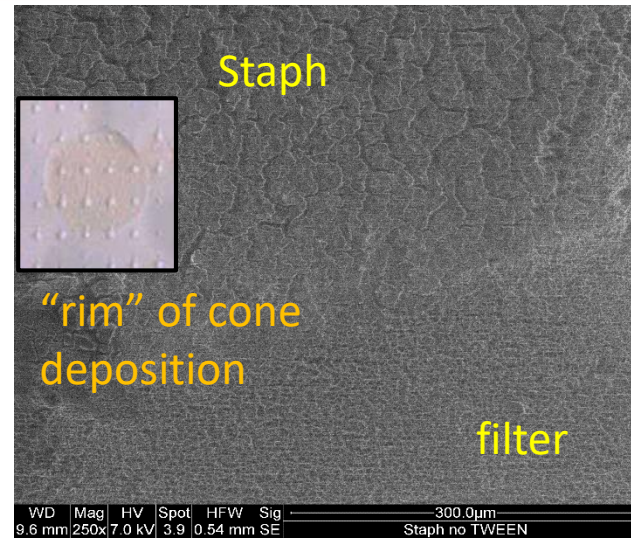
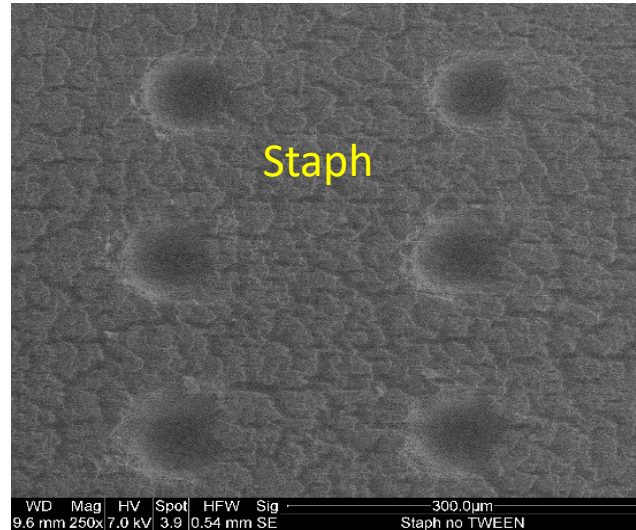


NSERC
CRSNG

Thanks To

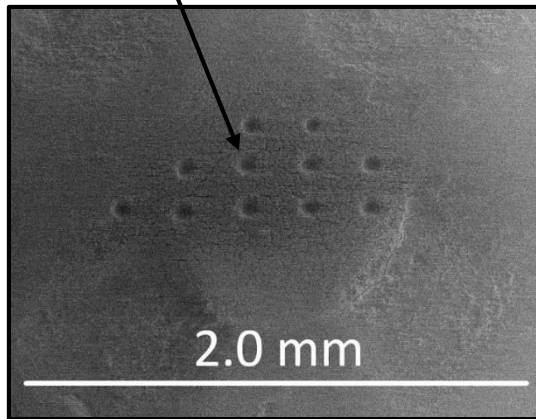


The Scatter in the Data

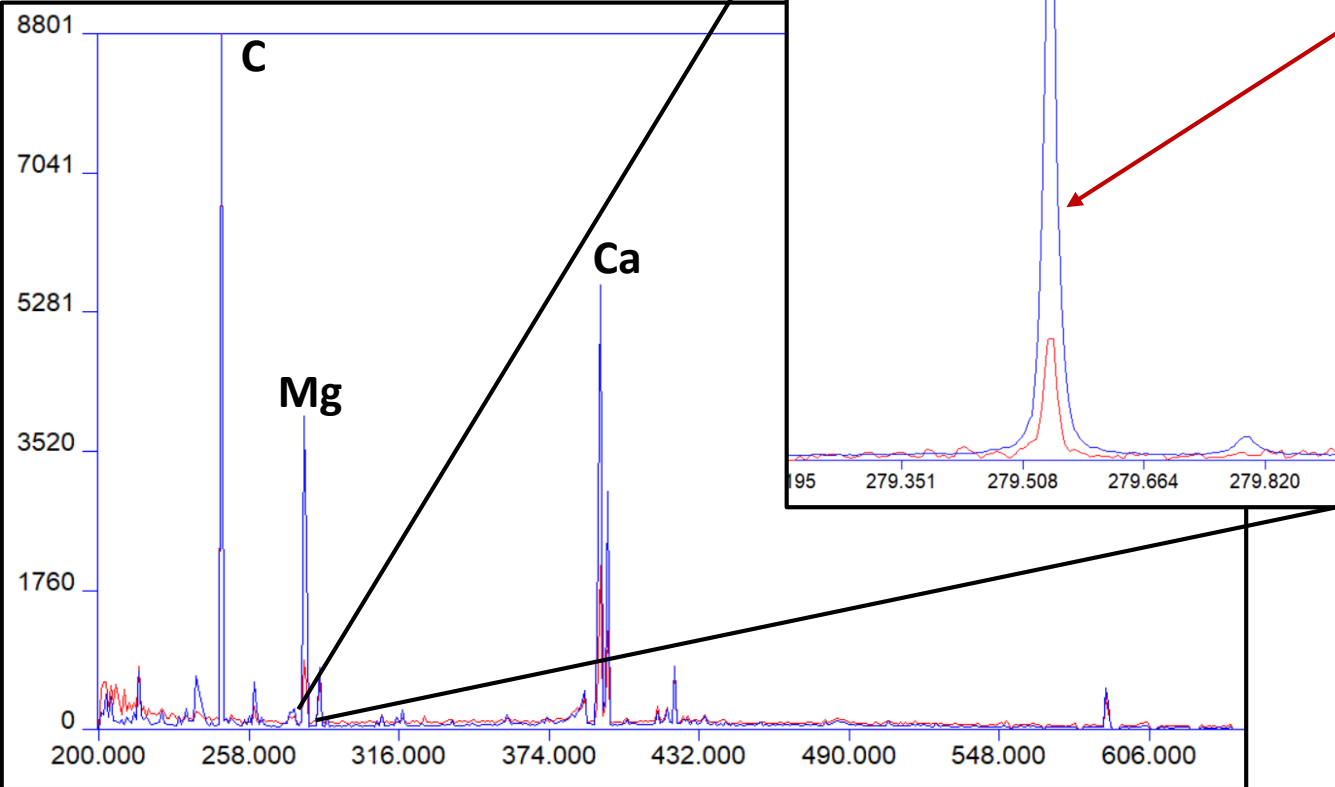


Effect of Adding Spectra

Circular area of filter
containing 10
single-shot spectra.



“Add-all” Spectrum
Single-Shot Spectrum



Magnesium
279 nm line
has greater
intensity

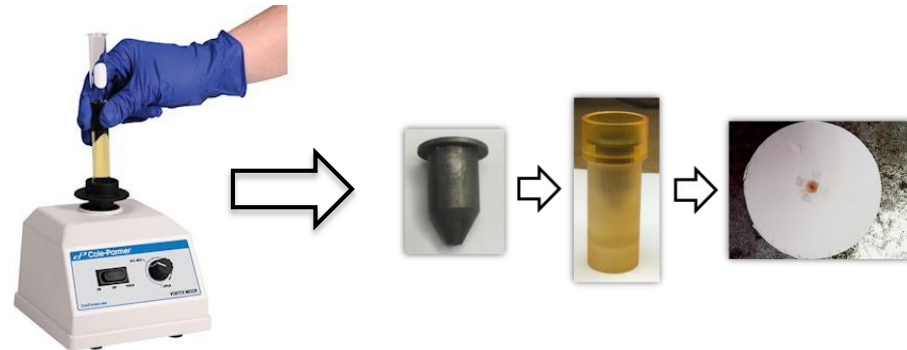
The Add-all
spectra
increases the
signal to
noise
ratio

Method 4



Cannot shoot right on the swab

- Far too irregular (almost no plasma)
- Cells not concentrated



Method 4



Cannot shoot right on the swab

- Far too irregular (almost no plasma)
- Cells not concentrated

