

# Recent Advances in the Use of Laser-Induced Breakdown Spectroscopy to Classify Pathogens in Clinical Specimens

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University of Windsor

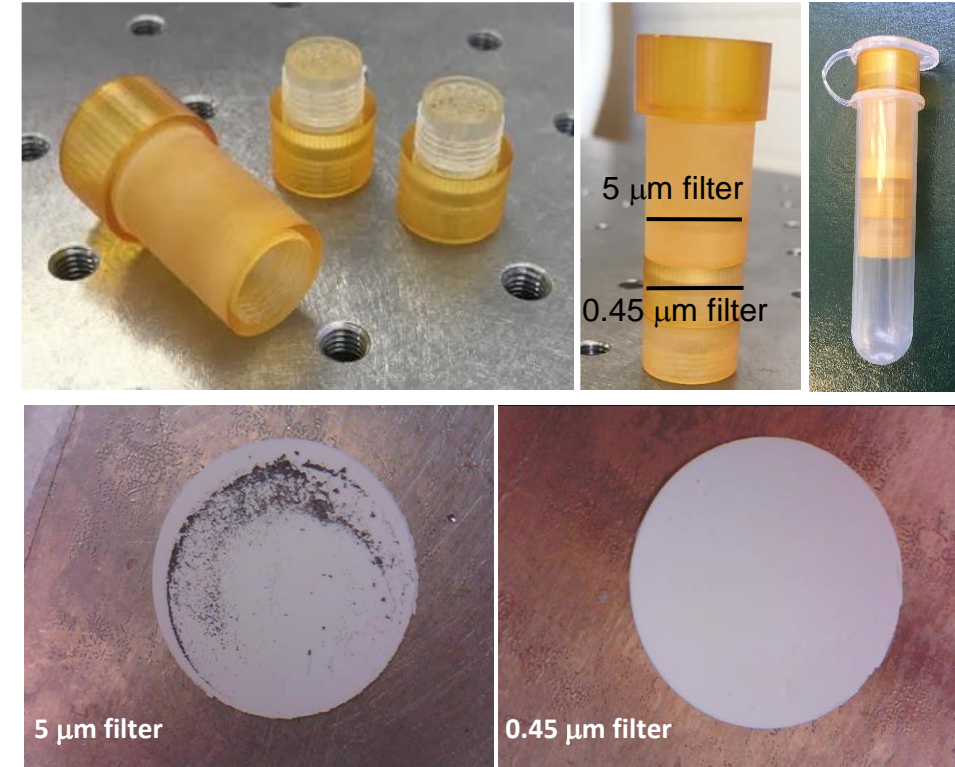
# History and Motivation

- Since I have been at the University of Windsor, we have invented a new way to analyze bacterial cells in liquid suspension.
- This method was used for all the studies described in this talk.





# Sample Centrifugation

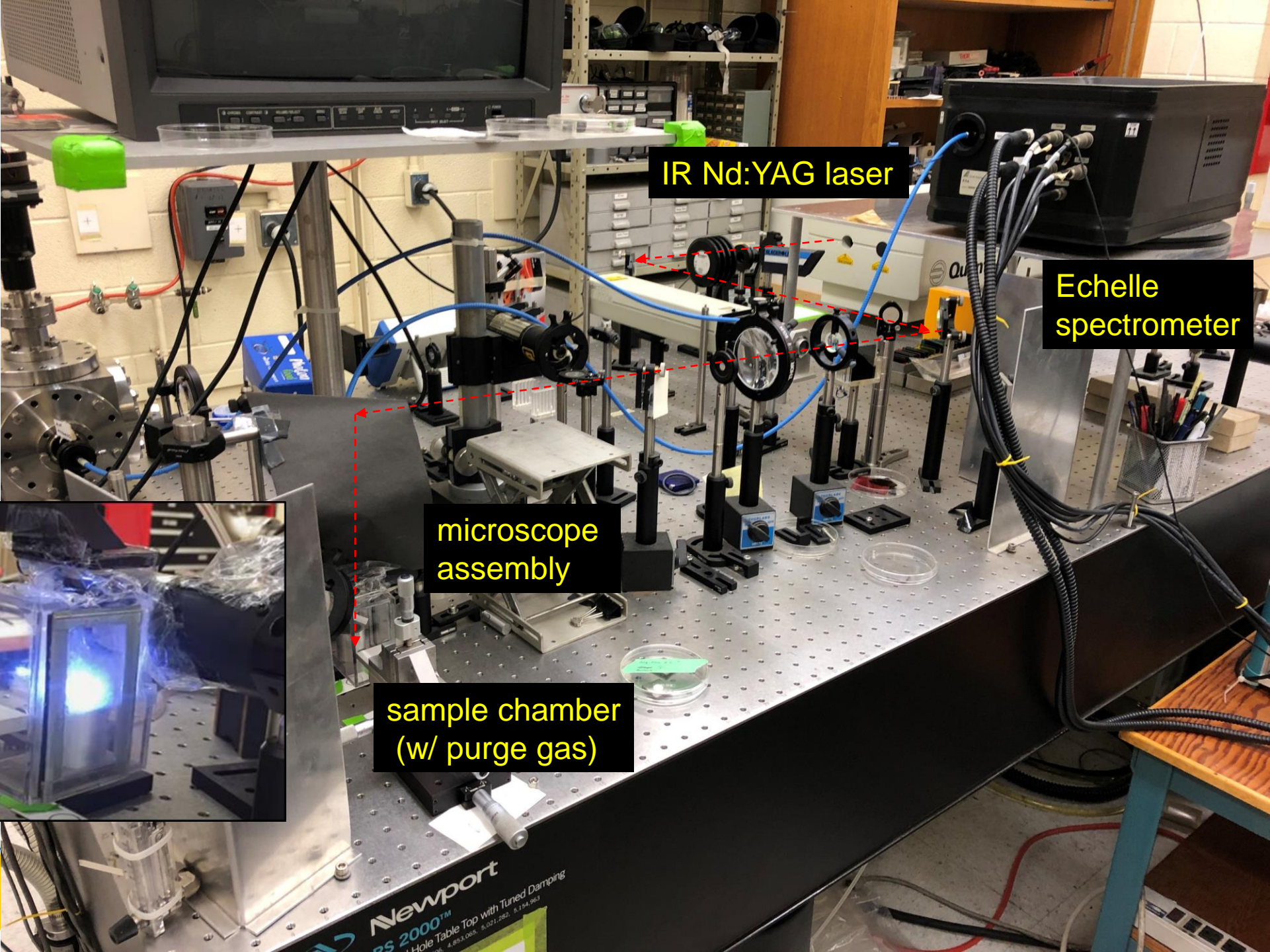


- We culture our own bacteria
- 3D printed “centrifuge insert”
- Nitrocellulose filters (Millipore. 0.45 and 0.22  $\mu\text{m}$ )
- 1 mL of bacterial suspension centrifuged at 5000 rpm
- Filter removed, bacteria on filter ablated
- Second base constructed to allow dual stage centrifugation







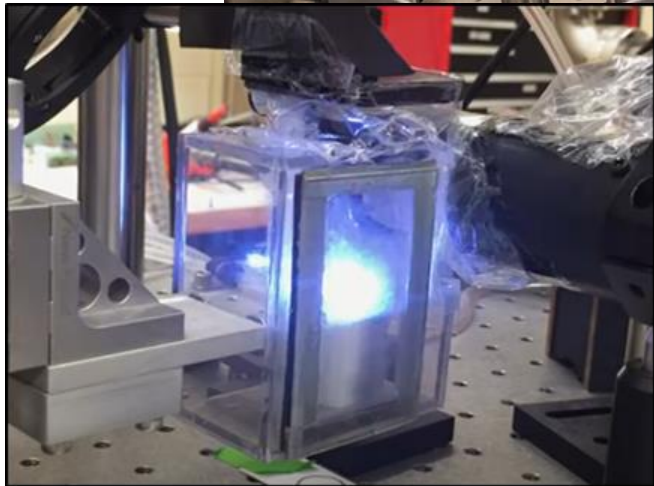


IR Nd:YAG laser

Echelle spectrometer

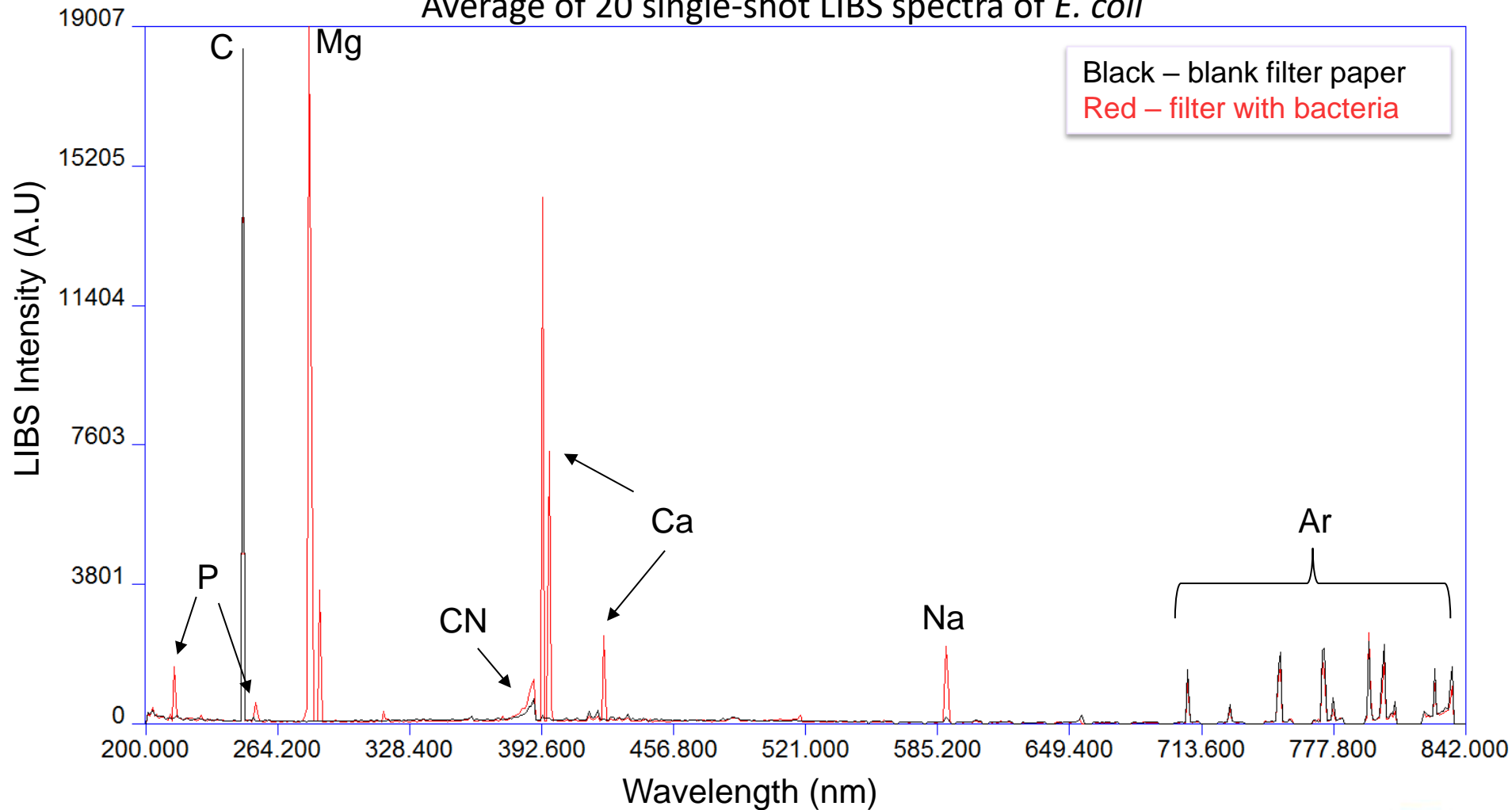
microscope assembly

sample chamber (w/ purge gas)



# Sample Centrifugation

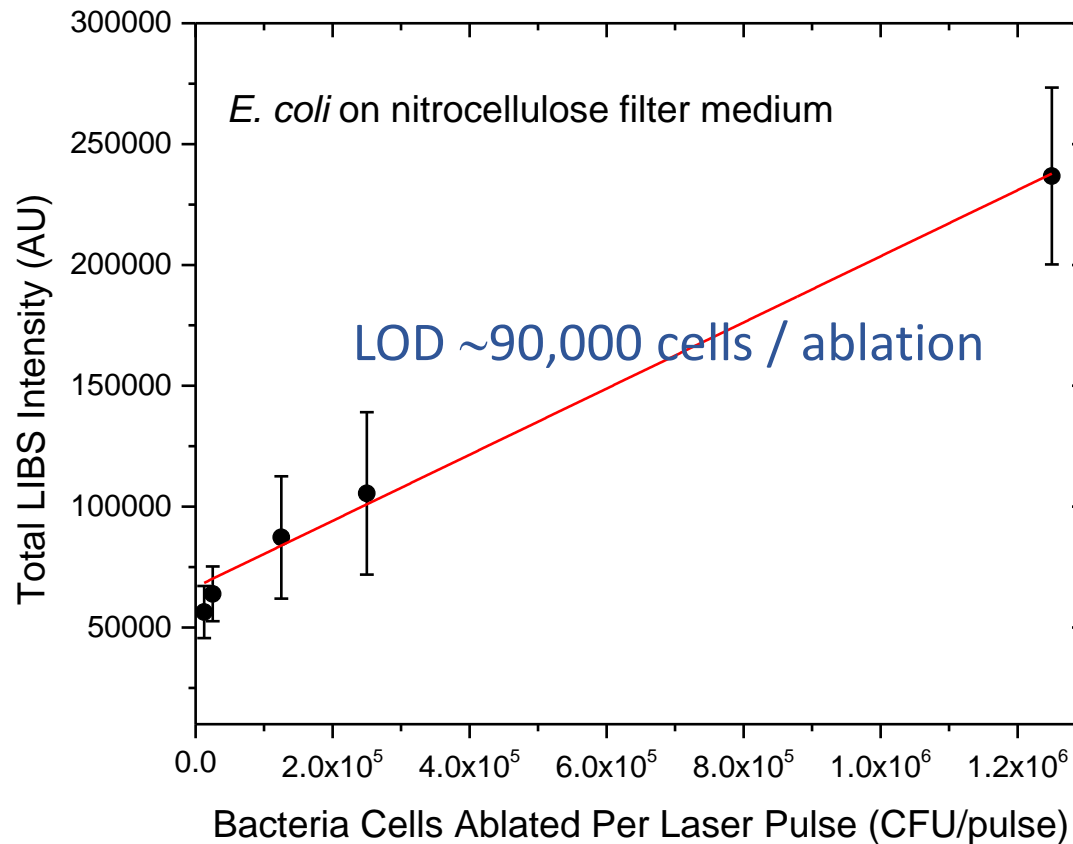
Average of 20 single-shot LIBS spectra of *E. coli*



- 1064 nm, 10 ns laser
- 8 mJ/pulse
- LLA ESA3000 w/ICCD
- $\tau_{\text{delay}} = 2 \mu\text{s}$
- $\tau_{\text{width}} = 20 \mu\text{s}$
- argon purge
- single shot spectra (all bacteria ablated after one shot)



# Sample Centrifugation



- 1064 nm, 10 ns laser
- 8 mJ/pulse
- LLA ESA3000 w/ICCD
- $\tau_{\text{delay}} = 2 \mu\text{s}$
- $\tau_{\text{width}} = 20 \mu\text{s}$
- argon purge
- single shot spectra (all bacteria ablated after one shot)

Analytical note [Spectrochimica Acta Part B 158 \(2019\) 105629](#)

A simple and efficient centrifugation filtration method for bacterial concentration and isolation prior to testing liquid specimens with laser-induced breakdown spectroscopy<sup>☆</sup>

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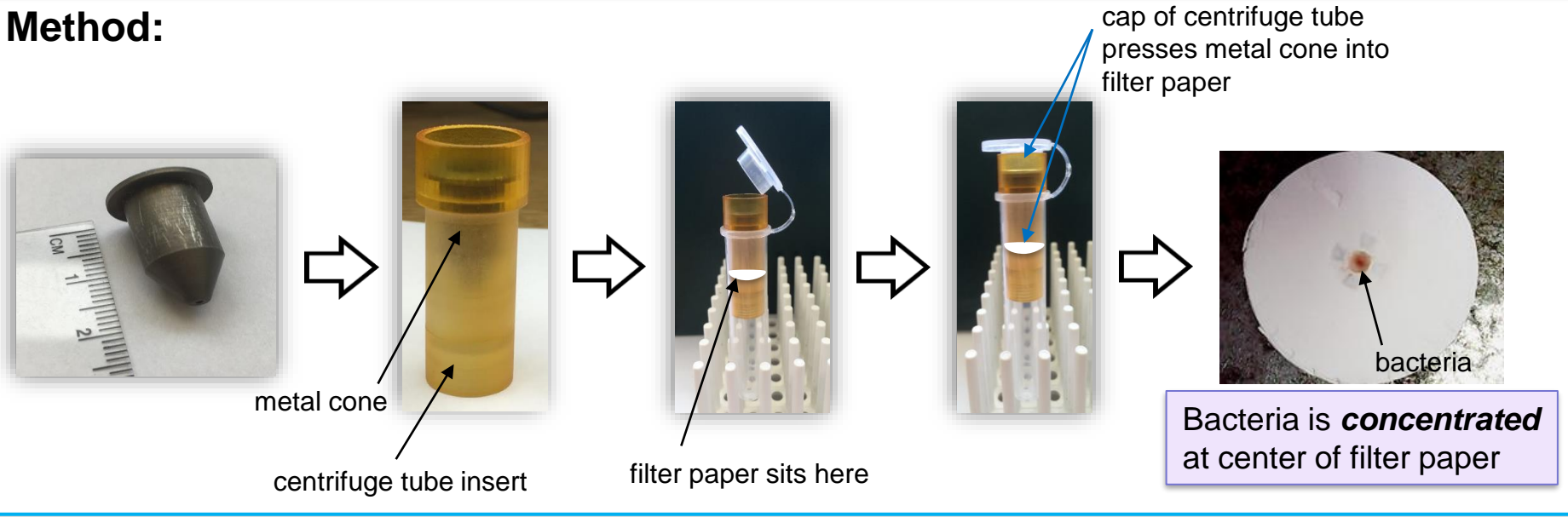


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# Sample Concentration

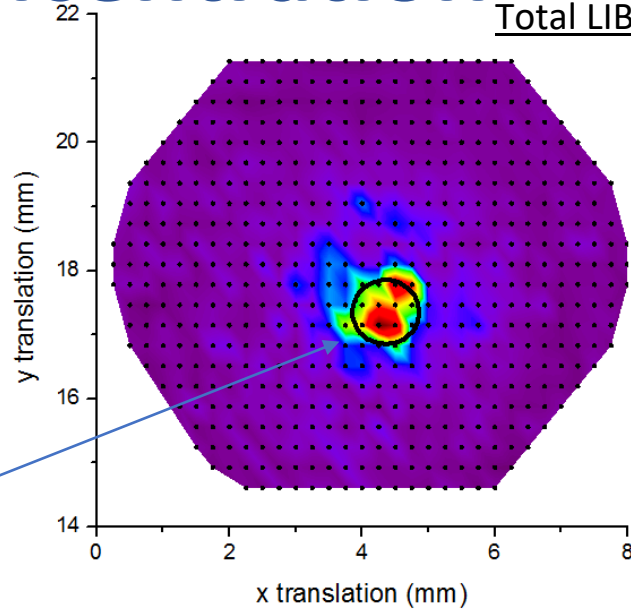
## Method:





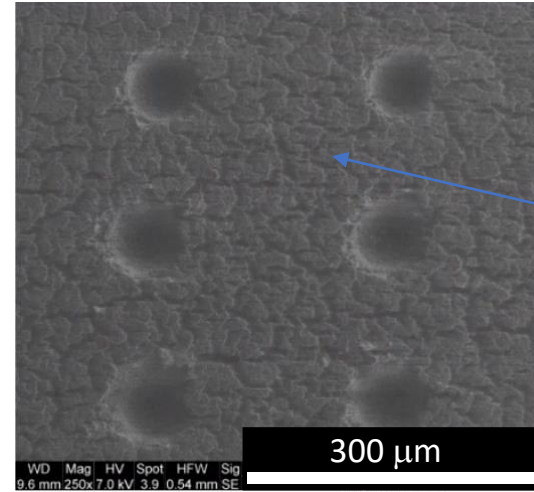
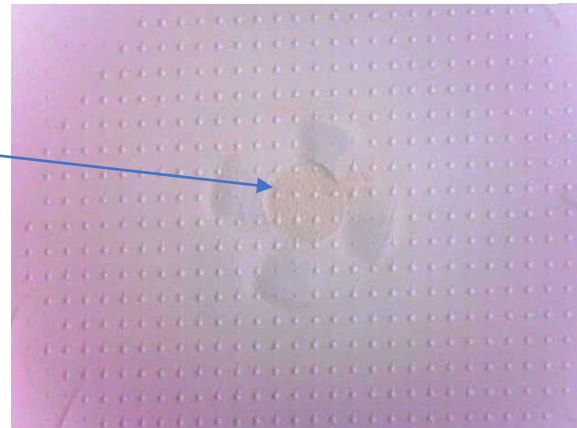
# Sample Concentration

Total LIBS intensity (A.U.)

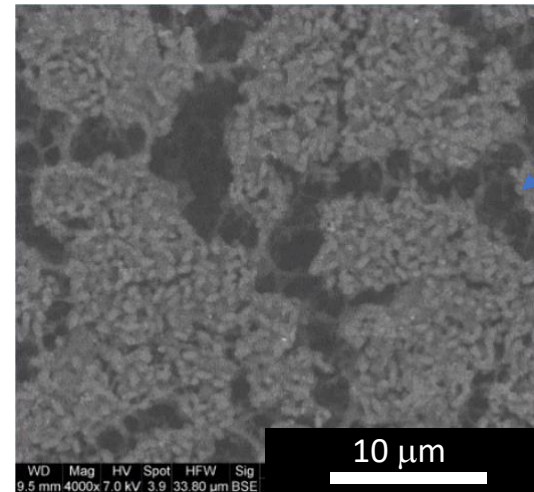


Cone nozzle ~1 mm

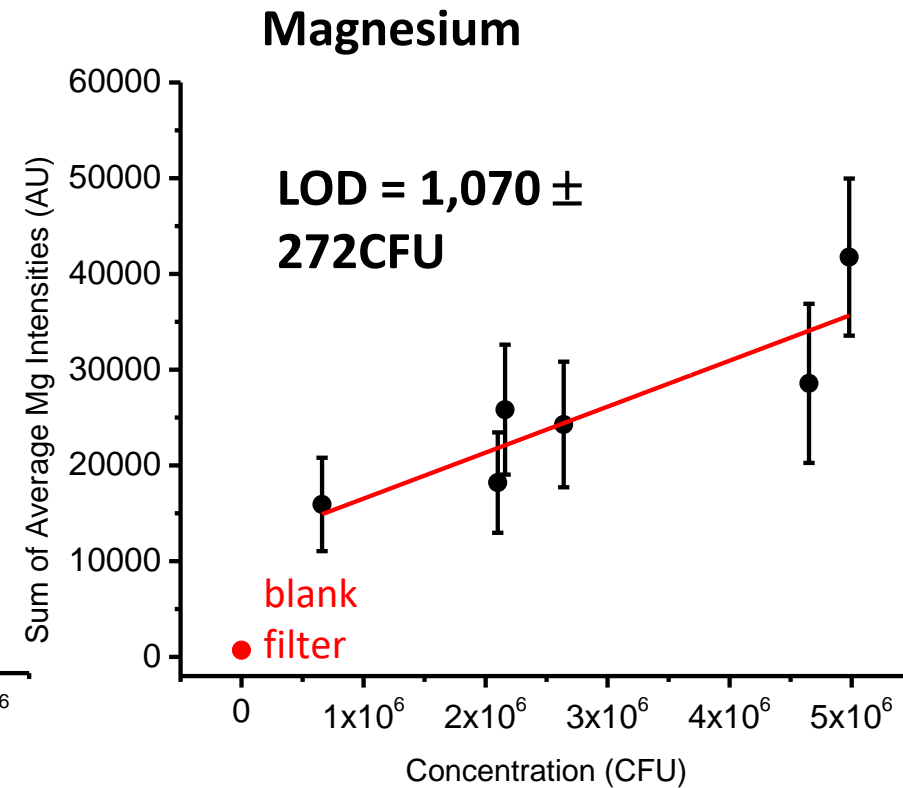
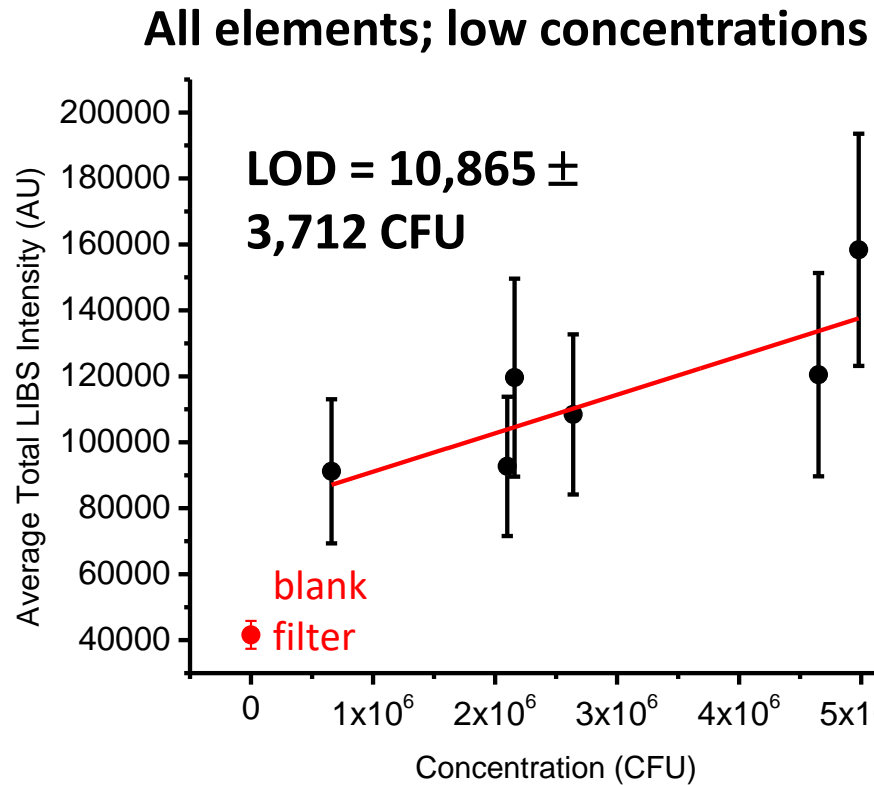
We can reliably acquire 20-30 shots per deposition



Irregular deposition could be problematic



# Sample Concentration



Spectrochimica Acta Part B 157 (2019) 68–75

Concentration of bacterial specimens during centrifugation prior to laser-induced breakdown spectroscopy analysis<sup>☆</sup>

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University of Windsor

# Recent Progress

- i. Biofluids (blood and urine) from hospital
- ii. Full-spectrum chemometrics utilizing a PCA-ANN
- iii. Use of silver to enhance plasma emission





# Recent Progress (i): Blood & Urine

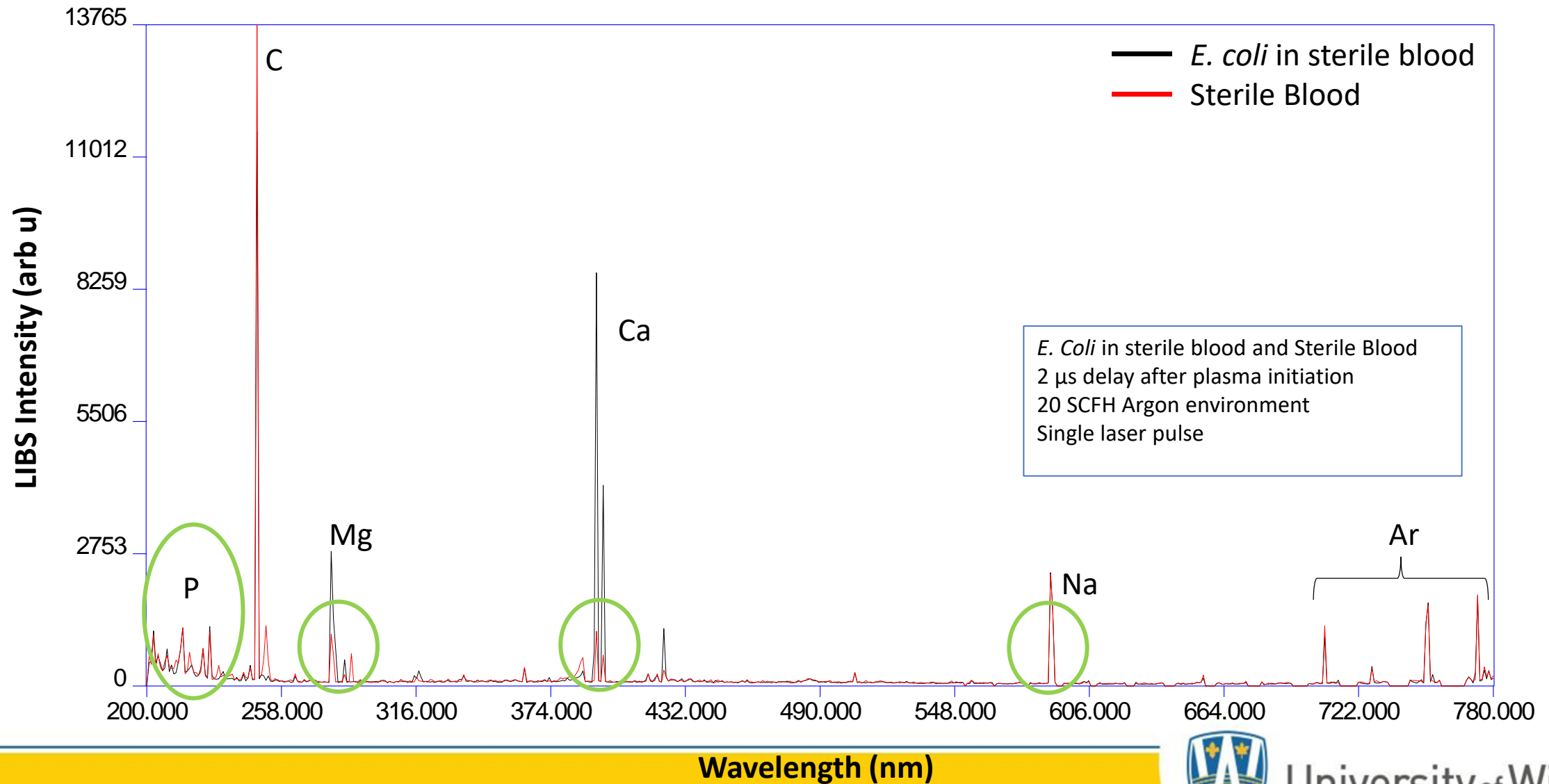
After testing in hospital pathology lab, bacterial negative, specimens sent to us



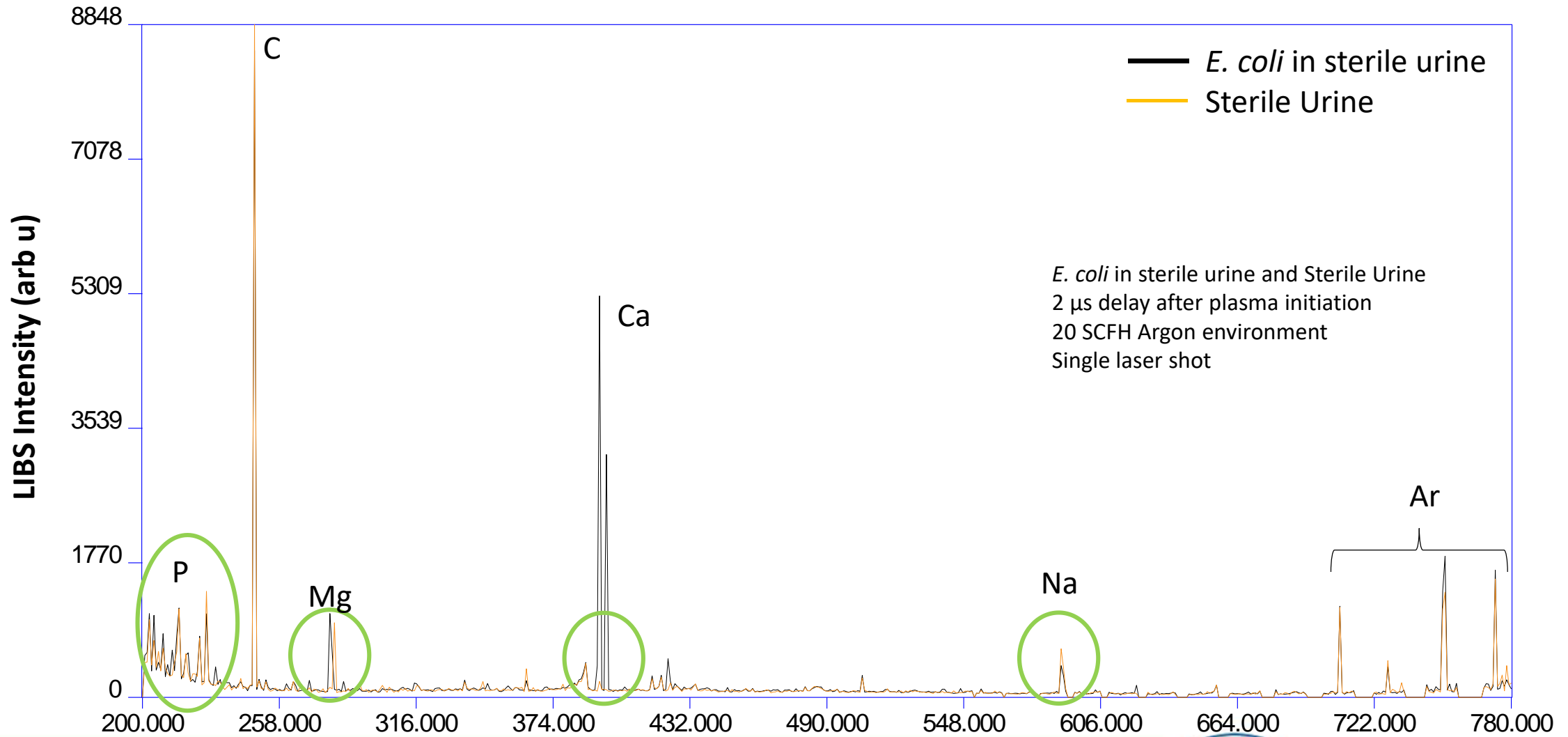
We “spike” the nominally sterile specimens with trace amounts of known bacteria (e.g. *E. coli*) to simulate sepsis and UTI.



# Recent Progress (i): Blood & Urine



# Recent Progress (i): Blood & Urine



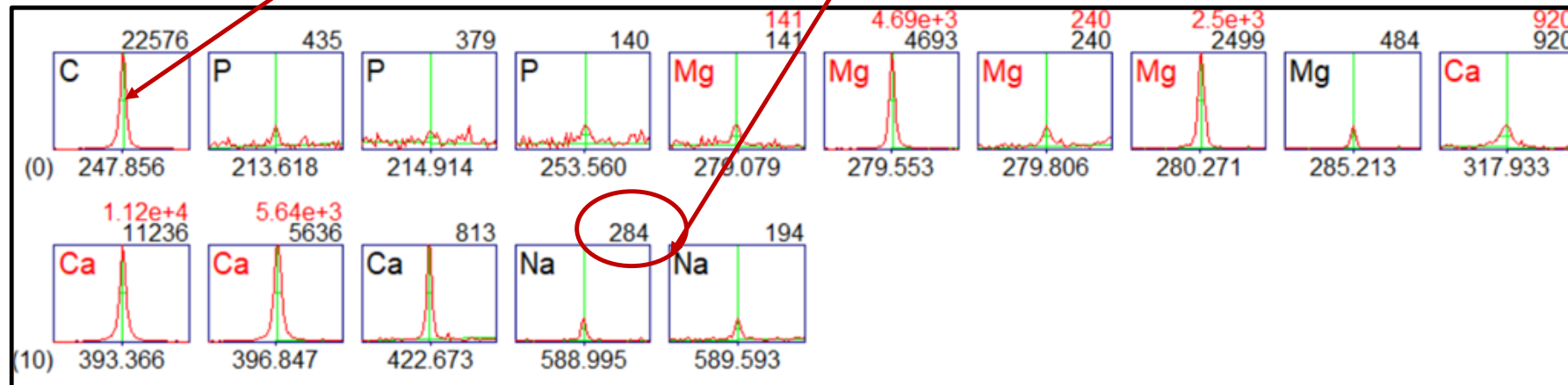


# Recent Progress (i): Blood & Urine

Variable down-selection. Ratio-model 2.5: 15 line areas, 92 ratios = 107 variables

Strong emission of carbon line comes from filter

Emission intensity is the integrated area under the curve

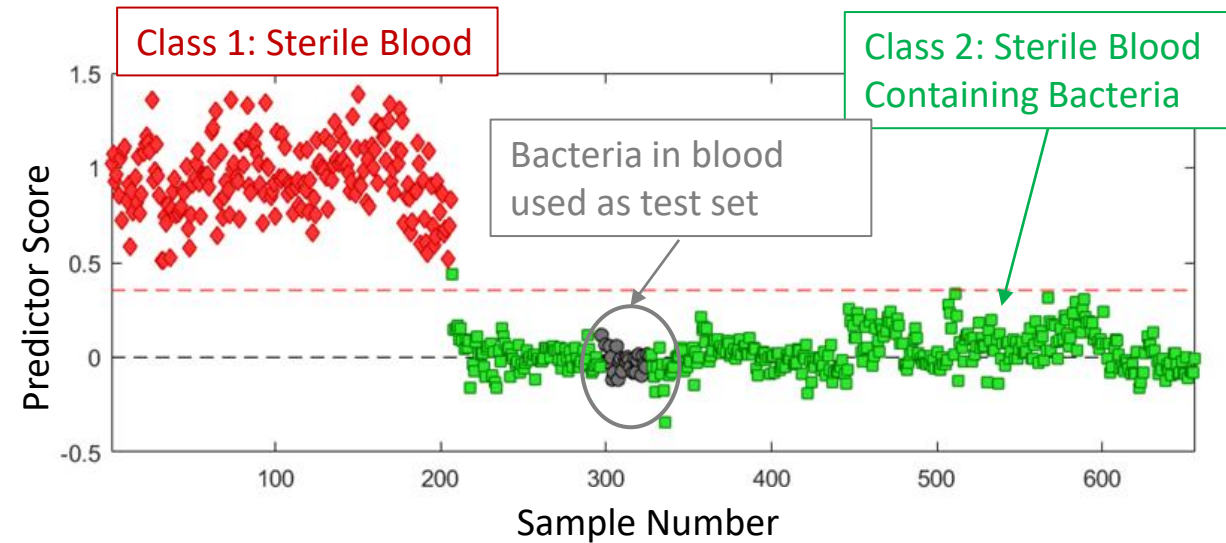


Phosphorus lines are highly indicative of bacteria. Na, Ca, and Mg also indicative of bacteria. All of these elements exist in the bacterial cell membrane.

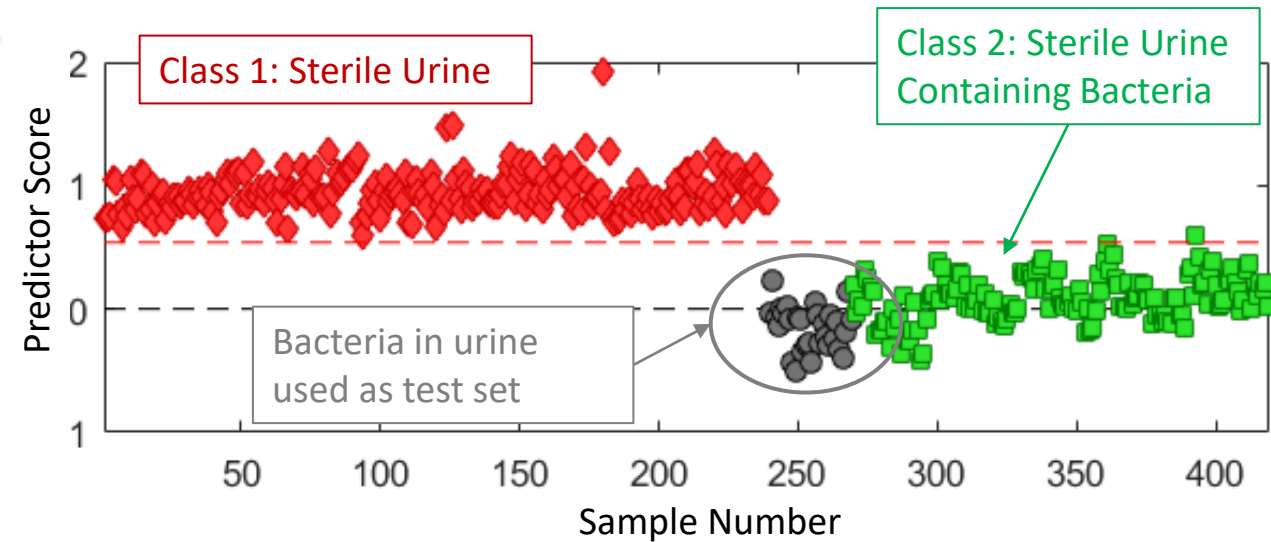


# Recent Progress (i): Blood & Urine

- A **partial least squares discriminant analysis (PLSDA)** test was conducted using external validation to determine if bacteria can be detected in blood and urine
- ✓ We can detect several types of bacteria in blood and urine reliably



**Average Sensitivity: 99.56%**  
**Average Specificity: 100%**

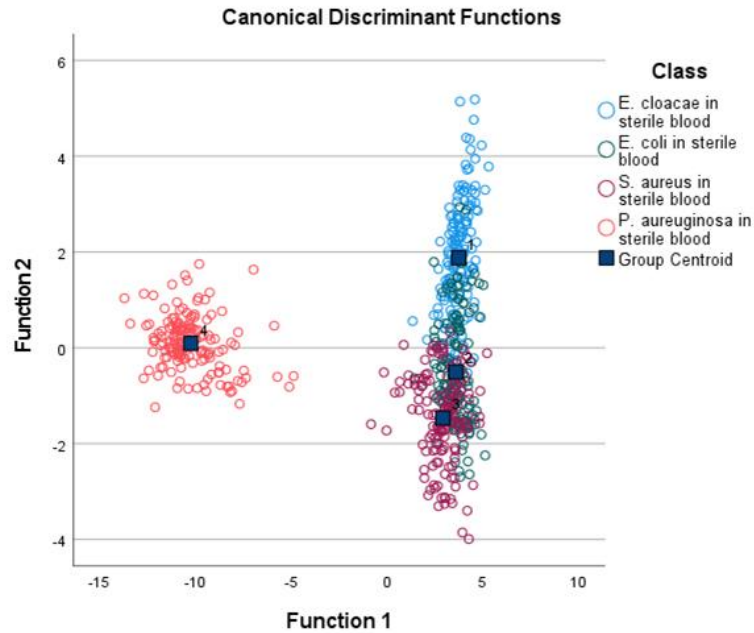


**Average Sensitivity: 98.90%**  
**Average Specificity: 100%**



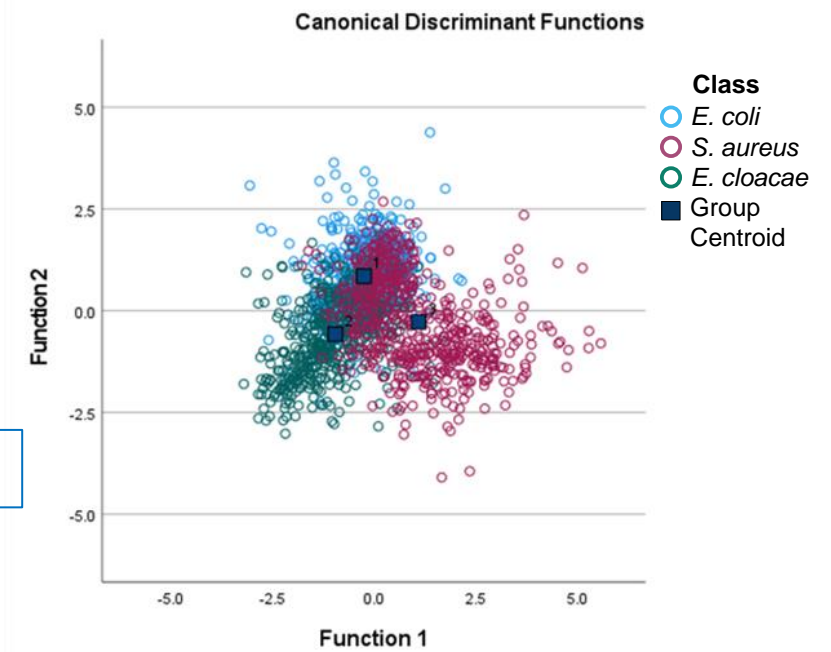
# Recent Progress (i): Blood & Urine

Variable down-selection. Ratio-model 2.5: 15 line areas, 92 ratios = 107 variables



4 bacteria in blood

3 bacteria in water



Sample Type (DFA)	Sensitivity	Specificity
Sterile Blood Containing <i>P. aeruginosa</i>	98.7 %	100 %
Sterile Blood Containing <i>S. aureus</i>	84.0 %	92.9 %
Sterile Blood Containing <i>E. coli</i>	74.7 %	93.6 %
Sterile Blood Containing <i>E. cloacae</i>	78.0 %	92.0 %

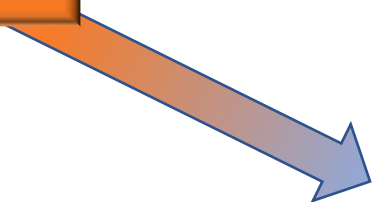
DFA Results			
Sample	Sensitivity (%)	Specificity (%)	Classification Error
<i>E. coli</i>	73.28 %	71.78 %	27.46 %
<i>S. aureus</i>	64.93 %	84.93 %	25.06 %
<i>E. cloacae</i>	60.10 %	92.53 %	23.68 %





# Recent Progress (ii): ANN. First alteration

DFA on RM2.5



ANN on RM2.5

- We implemented a feed forward, one-hidden layer ANN in Python.
- First optimized the # of epochs for a certain number of nodes.
- Used an algorithm that calculates sensitivity/specificity as a function of nodes and patience (100-140, 40).

Sample Type (ANN)	Sensitivity	Specificity
Sterile Blood Containing <i>P. aeruginosa</i>	97.2 %	100 %
Sterile Blood Containing <i>S. aureus</i>	100 %	100 %
Sterile Blood Containing <i>E. coli</i>	100 %	100 %
Sterile Blood Containing <i>E. cloacae</i>	100 %	98.9 %
Sample Type (DFA)	Sensitivity	Specificity
Sterile Blood Containing <i>P. aeruginosa</i>	98.7 %	100 %
Sterile Blood Containing <i>S. aureus</i>	84.0 %	92.9 %
Sterile Blood Containing <i>E. coli</i>	74.7 %	93.6 %
Sterile Blood Containing <i>E. cloacae</i>	78.0 %	92.0 %

ANN Results			
Sample	Sensitivity (%)	Specificity (%)	Classification Error
<i>E. coli</i>	98.0 %	97.7 %	2.13 %
<i>S. aureus</i>	93.3 %	97.2 %	4.28 %
<i>E. cloacae</i>	91.2 %	96.1 %	6.33 %
DFA Results			
Sample	Sensitivity (%)	Specificity (%)	Classification Error
<i>E. coli</i>	73.28 %	71.78 %	27.46 %
<i>S. aureus</i>	64.93 %	84.93 %	25.06 %
<i>E. cloacae</i>	60.10 %	92.53 %	23.68 %



# Recent Progress (ii): ANN. Second alteration

- Started using the “whole spectrum” from 200 nm - 590 nm. 42,000 variables.
- Perform unsupervised PCA first (implemented in Python), reduce to 10 PC's. (~30 minutes).
- Models are trained on 80% of single shot data, 20% reserved for testing. (~15 seconds).

DFA on RM2.5

```
graph TD; A[DFA on RM2.5] --> B[ANN on RM2.5]; B --> C[PCA-ANN on full spectrum*];
```

ANN on RM2.5

PCA-ANN on full spectrum\*

\*The full spectrum spans 200 nm – 760 nm, but no lines of interest > 590 nm



# Recent Progress (ii): ANN. Results.

- When using the 80:20 single shot data, performance is always essentially 100%.
- But this is an incorrect test. External validation is the only accurate test. ⇒ Testing unique specimens/patients.

3 bacteria in water

		actual		
		<i>E. coli</i>	<i>Staph</i>	<i>Myco</i>
predicted	<i>E. coli</i>	436	10	0
	<i>Staph</i>	12	387	1
	<i>Myco</i>	1	7	418

		<i>E. coli</i>	<i>Staph</i>	<i>Myco</i>
sens	0.971047	0.957921	0.997613	
spec	0.987849	0.985023	0.990621	

## *Escherichia coli*

E. COLI	Predicted			Sensitivity
	e. coli	staph	myco	
80321	30	0	0	100
102521	30	0	0	100
22322	28	2	0	93.33333
022322-1	24	6	0	80
32122	30	0	0	100
032122-2	30	0	0	100
41122	30	0	0	100
041122-2	30	0	0	100
041122-3	30	0	0	100
041122-4	30	0	0	100
041122-5	30	0	0	100
041122-6	30	0	0	100
041122-7	30	0	0	100
41322	28	2	0	93.33333
041322-2	26	2	1	89.65517
Sum	436	12	1	97.10468

## *Staphylococcus aureus*

STAPH	Predicted			Sensitivity
	e. coli	staph	myco	
41422	7	23	0	76.66667
81221	0	30	0	100
82421	0	15	0	100
22322	0	30	0	100
022322-2	0	25	5	83.33333
022322-3	0	30	0	100
022322-4	0	30	0	100
22422	0	30	0	100
022422-2	3	27	0	90
022422-4	0	28	1	96.55172
22522	0	30	0	100
022522-2	0	29	1	96.66667
022522-3	0	30	0	100
031822-2	0	30	0	100
Sum	10	387	7	95.79208

## *Mycobacterium smegmatis*

MYCO	Predicted			Sensitivity
	e. coli	staph	myco	
081522-1	0	0	30	100
081522-2	0	0	30	100
081522-3	0	0	30	100
081522-4	0	0	30	100
081522-5	0	0	30	100
081522-6	0	0	30	100
081522-7	0	0	30	100
081522-8	0	0	30	100
081522-9	0	1	29	96.66667
081522-10	0	0	30	100
081622-1	0	0	30	100
081622-2	0	0	30	100
081622-3	0	0	29	100
081622-4	0	0	30	100
Sum	0	1	418	99.76134

Data set is 42,000 x 1272





# Recent Progress (ii): ANN. Results.

- When using the 80:20 single shot data, performance is always essentially 100%.
- But this is an incorrect test. External validation is the only accurate test. ⇒ Testing unique specimens/patients.

4 bacteria in blood

		actual			
		Staph	E. coli	Entero	Pseudo
predicted	Staph	98	17	9	8
	E. coli	23	121	1	1
	Entero	29	0	139	0
	Pseudo	0	12	1	111

		Staph	E. coli	Entero	Pseudo
sens	0.653333	0.806667	0.926667	0.925	
spec	0.919048	0.940476	0.930952	0.971111	

*Staphylococcus aureus*

STAPH	Predicted				Sensitivity
	staph	e. coli	entero	pseudo	
031122-1	30	0	0	0	1
031122-2	1	0	29	0	0.0333333
041422-1	30	0	0	0	1
041422-2	30	0	0	0	1
041422-3	7	23	0	0	0.2333333
Sum	98	23	29	0	0.6533333
ENTERO	Predicted				Sensitivity
	staph	e. coli	entero	pseudo	
032922-1	0	0	30	0	1
032922-2	9	0	21	0	0.7
032922-3	0	0	29	1	0.9666667
032922-4	0	0	30	0	1
032922-6	0	1	29	0	0.9666667
Sum	9	1	139	1	0.9266667

*Enterobacter cloacae*

*Escherichia coli*

E. COLI	Predicted				Sensitivity
	staph	e. coli	entero	pseudo	
032522-1	0	30	0	0	1
032522-2	16	14	0	0	0.4666667
040122-1	0	30	0	0	1
040122-2	1	17	0	12	0.5666667
040122-3	0	30	0	0	1
Sum	17	121	0	12	0.8066667
PSE	Predicted				Sensitivity
	staph	e. coli	entero	pseudo	
071122-1	0	0	30	0	1
071122-2	9	0	21	0	0.7
071122-3	0	0	29	1	0.9666667
071122-4	0	0	30	0	1
071122-6	0	1	29	0	0.9666667
Sum	9	1	139	1	0.9266667

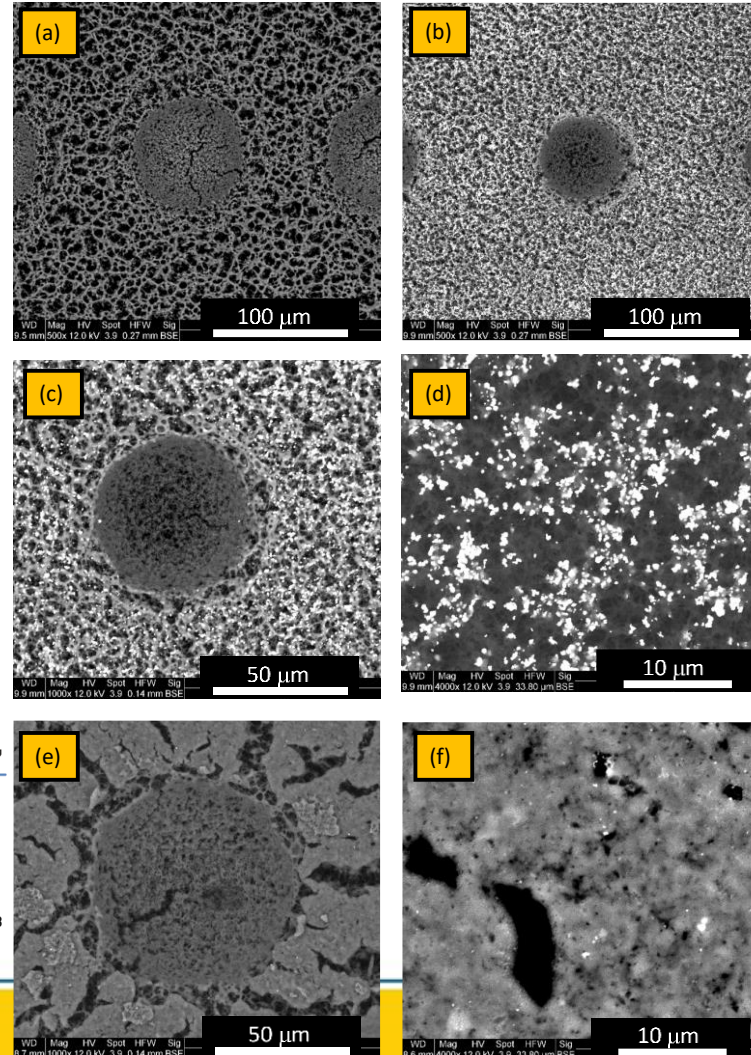
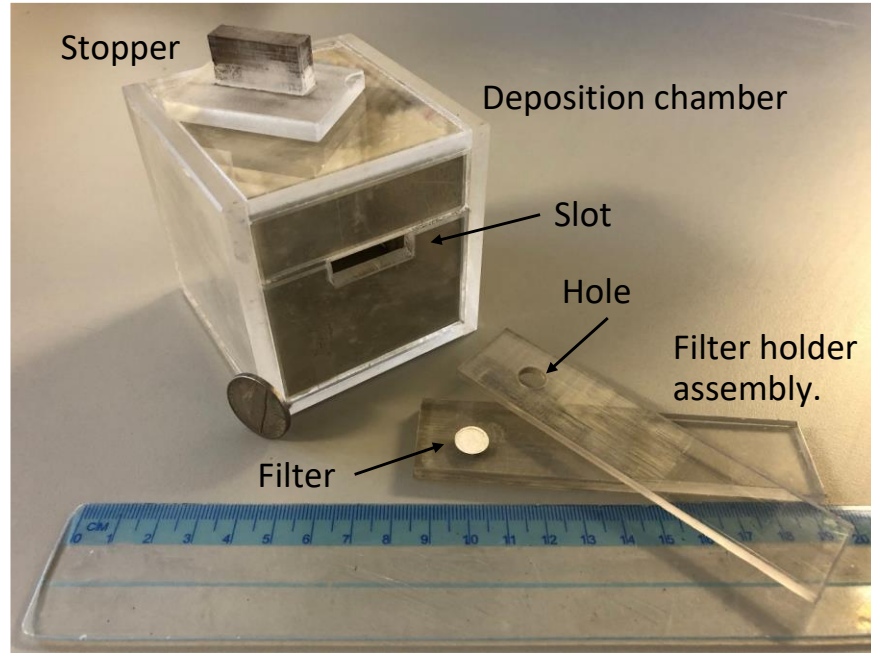
*Pseudomonas aeruginosa*

Our data tends to “fail by filter.”

We are revising our mounting method.



# Recent Progress (iii): Using silver.



- 0.5 - 1.0  $\mu\text{m}$  spheres
- 99.9% Ag
- 3.3 ng/shot

**Table I.** The average enhancement ratio for all emission lines for a given element in a LIBS spectrum of bacterial cells ablated in the presence of silver microparticles.

Bacteria Species	Elements				
	C	P	Mg	Ca	Na
<i>E. coli</i>	1.3	4.6	3.9	5.4	3.9
<i>M. smegmatis</i>	1.2	1.7	2.7	8.4	6.7
<i>E. cloacae</i>	1.2	4.4	6.9	2.2	1.3

Special Issue: Laser Induced Breakdown Spectroscopy

## Silver Microparticle-Enhanced Laser-Induced Breakdown Spectroscopy

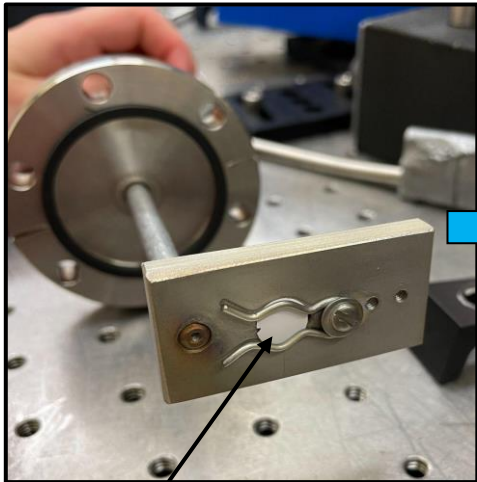
Applied Spectroscopy  
2022, Vol. 76(8) 905–916  
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DOI: 10.1177/00037028221096483  
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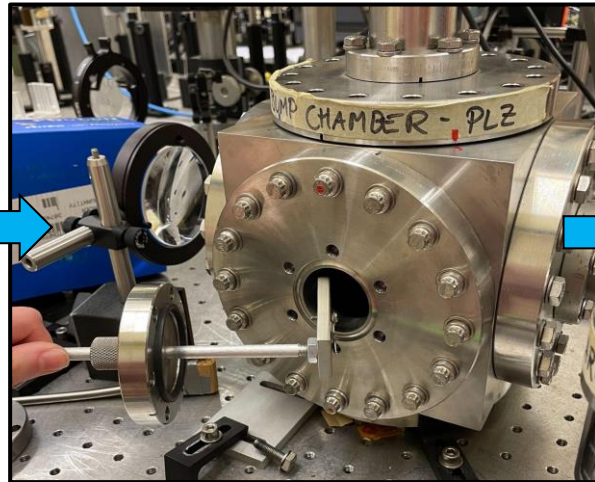
# Pulsed Laser Sputtering

A 1064 nm pulsed laser (60 mJ/pulse) is focused onto a rotating silver target inside a vacuum chamber.

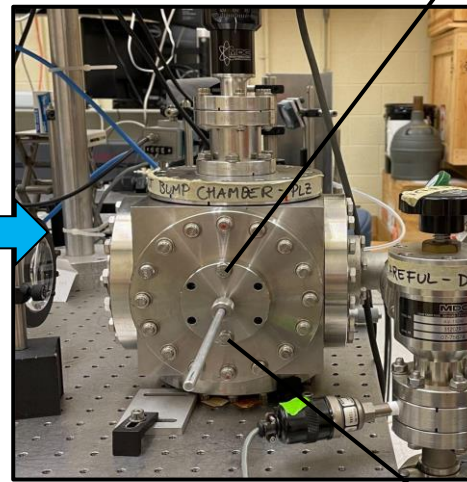
Experimental setup:



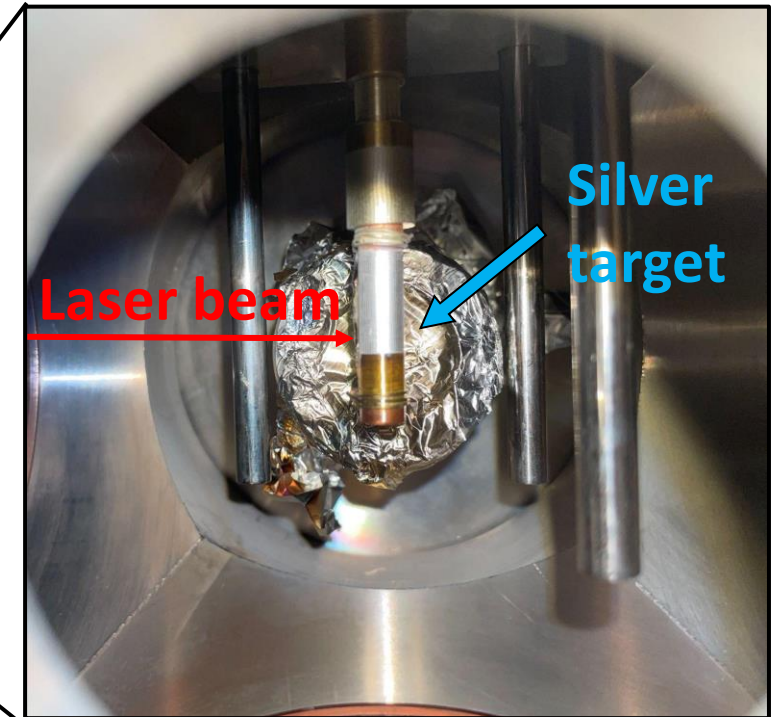
Filter on custom aluminum filter holder



Filter holder being inserted into 10 mTorr evacuated chamber

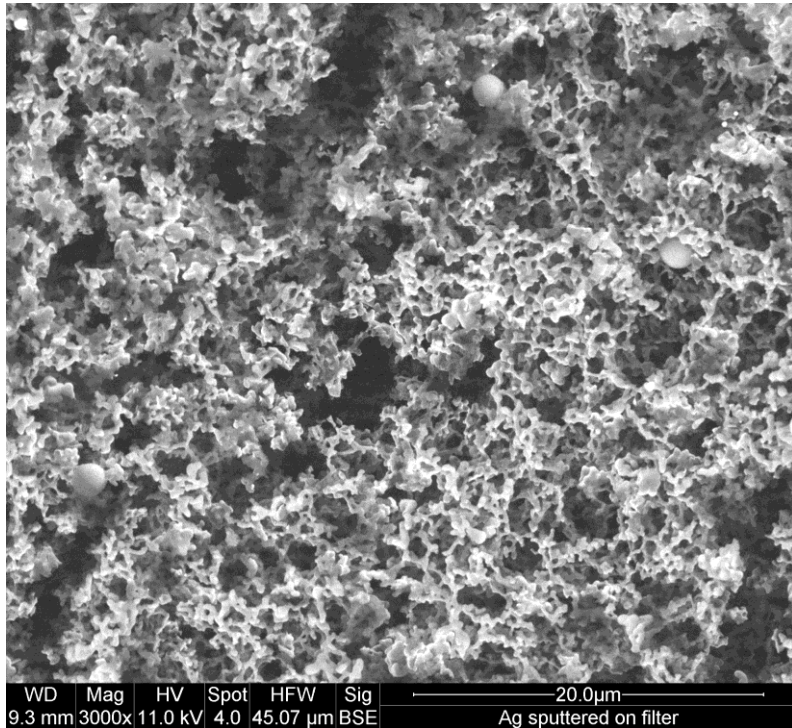


Closed vacuum chamber apparatus

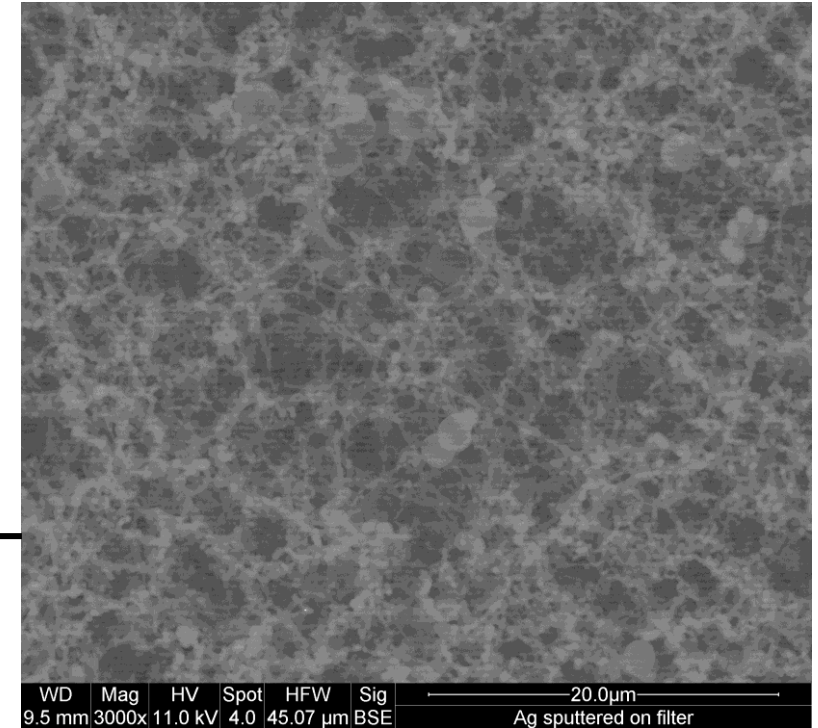
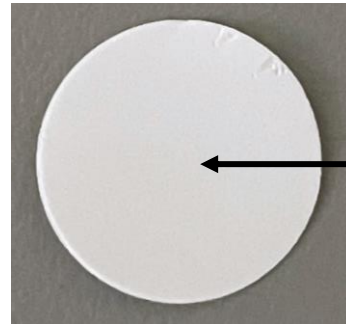
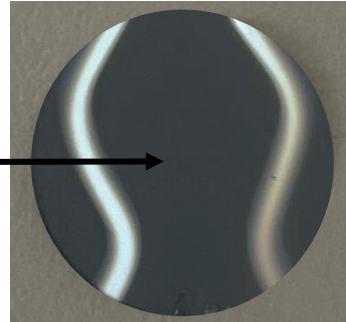


Inside the chamber

# Results: Silver Film Analyzed with Scanning Electron Microscope (SEM)



Filter with 20 min Ag



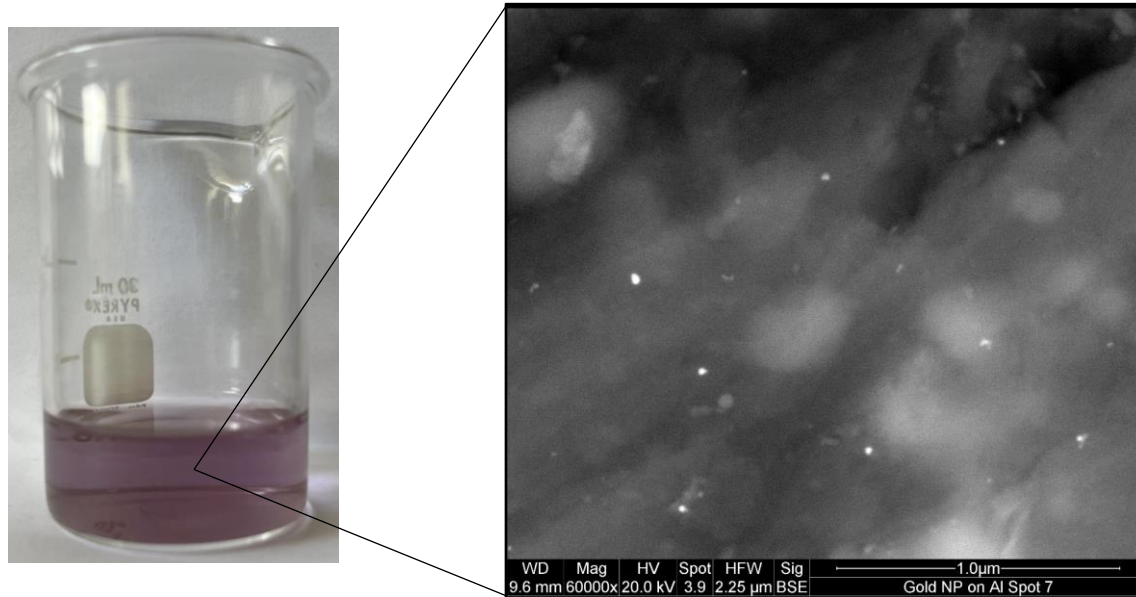
Blank Filter

The structure of the filter is unchanged but it is clear that silver has coated the filter.



# Future Work

- Use nano-particle enhanced LIBS (NELIBS) to increase the emission intensity of our bacteria
- We have successfully produced a gold nanoparticle suspension using pulsed laser ablation in liquid (PLAL)



Au Nanoparticle suspension

## Next Steps:

Investigate (i) laser pulse energy, (ii) volume of liquid, (iii) sputtering time to determine effect on size and spacing of the NP's.

Deposit NP's on filter or suspend with bacteria to investigate NELIBS.

# Acknowledgements

- My students:

- Emma Blanchette
- Emily Tracey
- August Baughan
- Grace Johnson



Summer 2022

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- NSERC USRA, Discovery Grant
- University of Windsor Outstanding Scholars Program



2021-2022

